Evaluation of 2, 3, 5-Triphenyltetrazolium Chloride as a Stain for Detection and Quantification of Experimental Cerebral Infarction in Rats

JOSHUA B. BEDERSON, M.D.,* LAWRENCE H. PITTS, M.D.,*† SABELLE M. GERMANO, M.D.,* MERRY C. NISHIMURA, A.B.,* RICHARD L. DAVIS, M.D.,† AND HENRY M. BARTKOWSKI, M.D., PH.D.*

SUMMARY We have evaluated the use of 2, 3, 5-triphenyltetrazolium chloride (TTC) as an histopathologic stain for identification of infarcted rat brain tissue. The middle cerebral artery (MCA) of 35 normal adult rats was occluded surgically. At various times after surgical occlusion, rats were sacrificed and brain slices were obtained and stained with TTC or hematoxylin and eosin (H & E); the size of the area of infarcted tissue stained by each method was quantified. In rats sacrificed 24 hr after occlusion of the MCA, the size of the area of infarction was 21 ± 2% of the coronal section for TTC, and 21 ± 2% for H & E (mean ± S.D., N = 13). The size of areas of infarction determined by either staining method was not significantly different in area by the paired t test, and a significant correlation between sizes determined by each method was found by linear regression analysis (r = 0.91, slope = 0.89, and the y intercept = 4.4%). Staining with TTC is a rapid, convenient, inexpensive, and reliable method for the detection and quantification of cerebral infarction in rats 24 hr after the onset of ischemia.

THE ABILITY to determine the location and extent of areas of infarction in cerebral tissue after ischemic injury is essential for the assessment of medical or surgical interventions, for the confirmation of clinical findings, and for the evaluation of new diagnostic techniques. Traditional histopathologic methods based on staining with hematoxylin and eosin (H & E) can accurately and reliably distinguish infarcted from normal tissue, but the method is costly and time-consuming. Infarction is identified at microscopic examination by shrinkage and dark staining of neurons and swelling with vacuolization of perineural and perivascular glial elements. The shrinkage of tissue during preparation for microscopic examination may affect estimates of the size of the area of infarction. Despite the fact that ultrastructural changes appear early after ischemic injury (see below), areas of cerebral infarction can be determined reliably by standard histopathologic techniques only several hours after the onset of ischemia. Other methods that could improve the detection and measurement of areas of ischemic infarction in cerebral tissue are needed.

2,3,5-Triphenyltetrazolium hydrochloride (TTC), first synthesized in 1894, was used originally to test the viability of seeds. Since 1958, TTC has been used as a stain to detect ischemic infarction in mammalian tissue. While this water soluble salt is not a dye, it is reduced by certain enzymes in normal tissue to a deep red, fat soluble, light-sensitive compound (formazan) that turns normal tissue deep red and thereby clearly delineates abnormal areas. TTC can act as a proton acceptor for many pyridine nucleotide-linked enzyme systems such as the dehydrogenases that, with the cytochromes, form an integral part of the inner mitochondrial membrane and make up the electron transport chain.

TTC has been used extensively to stain myocardial tissue obtained from humans and experimental animals. TTC staining reflects accurately the extent of irreversible myocardial ischemic damage in dogs and it has been reported that TTC can stain areas of injury to cerebral tissue in the cat. We report here the results of a comparative study of the use of TTC and H & E to stain cerebral tissue in rats in which infarction was produced by occlusion of the middle cerebral artery (MCA) for various times.

Methods

Thirty-five normal adult male Sprague Dawley rats weighing 300–400 gm, which were allowed free access to food and water before and after all procedures, were weighed and placed in an ether jar until they were immobilized. Anesthesia was induced with chloral hydrate dissolved in normal saline (35 mg/100 gm, intraperitoneally) and was maintained during the procedure by supplemental doses as needed. Body temperature was monitored with a rectal thermometer and was maintained within normal limits with a heating pad.

Using an operating microscope under high magnification, the left MCA was exposed transcranially without damage to the zygomatic bone. The brain was retracted gently to gain proximal exposure and the MCA was occluded with microbipolar forceps on a low power setting using continuous saline irrigation. Areas of infarction of different sizes were produced by occlusion of the MCA 1 to 3 mm proximal to the olfactory tract and extending to the inferior cerebral vein; the artery was transected to insure that recanal-
TTC STAINING FOR CEREBRAL INFARCTION/Bederson et al 1305

zation did not occur. The temporalis muscle and skin were sutured and rats were returned to cages until they either recovered from anesthesia or were sacrificed. The procedure was performed in 20 to 25 minutes; brains were retracted for less than 5 minutes.

After various survival times, rats were sacrificed by an intracardiac injection of sodium pentobarbital (75 mg of a 390 mg/cc solution). Rats were divided into four groups based on the time of sacrifice after occlusion. Time of sacrifice after occlusion for each group was: Group 1, 1 to 2 hours (N = 6); Group 2, 3 hours (N = 3); Group 3, 5 to 6 hours (N = 3); and Group 4, 24 hours (N = 23).

Within 3 min of sacrifice, brains were removed, two coronal slices were made at 5 and 7 mm from the frontal pole, and brain slices were immersed in a 2% solution of TTC in normal saline at 37°C for 30 min, after which sections were fixed in 10% phosphate-buff ered formalin for photography and H & E staining.

To assess the effects of formalin and time on fading of TTC and the estimation of the size of the area of infarction, sections from 10 brains from Group 4 rats were stored in formalin for 10 to 40 days before photography and sectioning. The other specimens (N = 13) were photographed and sectioned 1 to 5 days after fixation. The rostral surface of the TTC-stained sections was photographed using color slide film (Ektachrome, tungsten 160 ASA). For comparison with H & E staining, histologic sections were prepared from the surface of the same slices and stained with H & E.

Projection tracings of the H & E and TTC stained sections from the same slice were made at different times by an independent observer who was not aware of the group from which the slices were obtained. Cortex and basal ganglia were outlined separately and the area of infarction was determined both by computer image analysis and by differential weighing of cut out areas of the tracing. Essentially identical results were obtained by both methods. The area of infarction in the cortex and the basal ganglia was expressed as the percent of the whole coronal section. Histologic sections were reviewed by one of us (R.D.) to confirm infarction size. Data for TTC and H & E sections were paired and examined for the possible existence of correlations.

Results

TTC stained normal areas of brain deep red but did not stain infarcted tissue. In tissue from Group 4 rats (24 hr after occlusion), the border between normal and infarcted tissue was well-defined and could be detected easily by gross inspection. The deep red color of TTC-stained sections disappeared during alcohol dehydration of the formalin-fixed brain slices. H & E slides prepared from these slices could not be distinguished from slices not stained with TTC, which allowed an unbiased assessment of the H & E slides. Tissue from Group 4 rats was characterized by a central area of coagulative necrosis surrounded by a zone of peripheral spongiosis. Generalized tissue pallor defined the limits of the area of infarction in each specimen. Cor-

![Figure 1](http://stroke.ahajournals.org/)

**Figure 1.** A (upper). TTC-stained coronal brain slice 24 h after occlusion of the MCA (upper). B (lower). H & E stained section from the surface of the brain slice shown in 1A.
Table 1  Infarction Size Determined by TTC and H & E Staining as the Percentage of Coronal Section

<table>
<thead>
<tr>
<th>Group 4: 24 hours after occlusion (N = 23)</th>
<th>TTC</th>
<th>H &amp; E</th>
</tr>
</thead>
<tbody>
<tr>
<td>basal ganglia plus cortex</td>
<td>21.2±2.4</td>
<td>21.3±2.0</td>
</tr>
<tr>
<td>basal ganglia</td>
<td>11.8±0.6</td>
<td>11.8±0.9</td>
</tr>
<tr>
<td>cortex</td>
<td>10.4±1.6</td>
<td>9.8±1.41</td>
</tr>
<tr>
<td>Group 3: 5-6 hours after occlusion (N = 3)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>basal ganglia plus cortex</td>
<td>16.3±5.1</td>
<td>18.0±1.7</td>
</tr>
<tr>
<td>basal ganglia</td>
<td>9.6±2.5</td>
<td>10.9±2.0</td>
</tr>
<tr>
<td>cortex</td>
<td>6.7±0.7</td>
<td>7.1±1.2</td>
</tr>
<tr>
<td>Group 2: 3 hours after occlusion (N = 3)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>basal ganglia plus cortex</td>
<td>7.0±0.9</td>
<td>14.1±0.8‡</td>
</tr>
<tr>
<td>basal ganglia</td>
<td>6.3±1.0</td>
<td>8.6±0.91</td>
</tr>
<tr>
<td>cortex</td>
<td>0.7±0.7</td>
<td>5.4±0.7*</td>
</tr>
<tr>
<td>Group 1: 1-2 hours after occlusion (N = 6)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>basal ganglia plus cortex</td>
<td>10.5±1.7</td>
<td>7.2±1.5</td>
</tr>
<tr>
<td>basal ganglia</td>
<td>6.6±0.6</td>
<td>5.3±1.2</td>
</tr>
<tr>
<td>cortex</td>
<td>3.9±1.4</td>
<td>1.8±0.6</td>
</tr>
<tr>
<td>24 hours after occlusion, with delay (N = 4§)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>basal ganglia plus cortex</td>
<td>26.0±3.4</td>
<td>20.4±3.6‡</td>
</tr>
</tbody>
</table>

Mean ± S.E.; * = p < 0.025; † = p < 0.005; ‡ = p < 0.01.
§Ten to 40 days before photography and sectioning.

FIGURE 2. Infarction size after 24 h survival determined by TTC and H & E staining for basal ganglia and cortex in each specimen (r = 0.91, slope = 0.89, y intercept = 4.4%).

Variation in the size of the area of infarction between TTC and H & E stained sections was the result primarily of the difference in the areas of infarction in the basal ganglia; despite the fact that there was no significant difference in size by the paired t-test within the entire lesion (basal ganglia plus cortex) and for either basal ganglia or cortex alone. Linear regression analysis showed that a significant correlation existed between data obtained for both stains (r = 0.91, slope = 0.89, y intercept = 4.4%) that did not differ statistically from a line with a slope = 1 and y intercept = 0 (H0; slope = 1 and y intercept = 0, p = 0.16, NS) (fig. 2).

In photographs of TTC-stained slices from 10 Group 4 rats that were held in formalin for 10 to 40 days, color had faded visibly from the margins of normal and infarcted tissue, which made the lesion (area not stained) appear larger. For these slices, the mean size of the infarcted area was 26.1 ± 3.4% for TTC and
20.4 ± 3.6% for H & E (paired t-test; \( p < 0.005 \)) (table 1).

**Discussion**

We found that it was difficult to compare results for H & E and TTC-stained sections in rats sacrificed 6 h or less after the onset of cerebral ischemia. TTC and H & E were equally effective for the determination and quantification of infarction 24 h after the onset of ischemia.

Infarction can be detected in TTC-stained infarcted coronary artery tissue in dogs as early as 1 to 2 h after occlusion, although color differences between normal and infarcted tissue are subtle. Three to 6 h after occlusion of the coronary artery, the border between TTC-stained normal and infarcted tissue becomes distinct and the border is delineated before the development of definite histologic evidence of infarction, and the time at which delineation occurs coincides with the rate of reduction of levels of dehydrogenase activity after occlusion of the coronary artery. Depletion of dehydrogenase is an irreversible process in myocardium and electron micrographs show that TTC reliably localizes infarcted cells only after extensive, irreversible ischemic damage has occurred. Thus, TTC-staining can detect ischemic changes in myocardium earlier than conventional methods, but by 12 to 24 h after coronary occlusion, the sensitivity of TTC and H & E are essentially equivalent and correlate significantly.

Depending on regional physiologic factors that affect cerebral tissue, ultrastructural changes may appear after 15 to 30 min of ischemia. Because the rate of cerebral oxygen consumption (CMRO₂) is reduced in the immediate postischemic period in rat brain, staining with an agent such as TTC that is based on the presence of intact enzymes of the electron transport chain might be expected to delineate infarction at an earlier stage than traditional histologic methods. However, irreversible injury to mitochondria in brain cells may not occur until after a long period of ischemia. Moreover, because coupling between electron transport and \( O_2 \) consumption in ischemic brain cells is altered, a decrease in CMRO₂, may not immediately reflect decreased dehydrogenase activity. Finally, the time necessary to cause irreversible damage to oxoreductases may be far longer than the duration of ischemia, as suggested by the results of this study. This may explain in part why unequivocal changes observed in tissue stained with TTC paralleled rather than preceded the irreversible changes seen in tissue stained with H & E.

TTC effectively and accurately detected areas of infarction 24 h after the onset of ischemia. Because TTC can be used to stain tissue much more rapidly, easily, and cheaply than techniques based on H & E — 30 min vs. several days/rat and \( \$9.00 \) vs \( \$5.00 \) per rat for TTC and H & E, respectively — it is a useful method for the detection of infarction in cerebral tissue. However, it is not clear at this time whether TTC can be used for the evaluation of other types of cerebral injury. TTC staining is an excellent research method that can be used to confirm the size and location of areas of infarction, and may be valuable to determine the presence and location of infarction in autopsy samples.

**Acknowledgment**

We thank Neil Buckley for preparing and editing the manuscript.

**References**

5. Stowell RA: Effect on tissue volume of various methods of fixation, dehydration, and embedding. Stain Technology 16: 67, 1941
Evaluation of 2,3,5-triphenyltetrazolium chloride as a stain for detection and quantification of experimental cerebral infarction in rats.

J B Bederson, L H Pitts, S M Germano, M C Nishimura, R L Davis and H M Bartkowski

Stroke. 1986;17:1304-1308
doi: 10.1161/01.STR.17.6.1304

Stroke is published by the American Heart Association, 7272 Greenville Avenue, Dallas, TX 75231
Copyright © 1986 American Heart Association, Inc. All rights reserved.
Print ISSN: 0039-2499. Online ISSN: 1524-4628

The online version of this article, along with updated information and services, is located on the World Wide Web at:
http://stroke.ahajournals.org/content/17/6/1304

Permissions: Requests for permissions to reproduce figures, tables, or portions of articles originally published in Stroke can be obtained via RightsLink, a service of the Copyright Clearance Center, not the Editorial Office. Once the online version of the published article for which permission is being requested is located, click Request Permissions in the middle column of the Web page under Services. Further information about this process is available in the Permissions and Rights Question and Answer document.

Reprints: Information about reprints can be found online at:
http://www.lww.com/reprints

Subscriptions: Information about subscribing to Stroke is online at:
http://stroke.ahajournals.org//subscriptions/