Anesthetic Modulation of Cerebral Hemodynamic and Evoked Responses to Transient Middle Cerebral Artery Occlusion in Cats

Mark A. Helfaer, MD, Jeffrey R. Kirsch, MD, and Richard J. Traystman, PhD

We measured cerebral blood flow and somatosensory evoked potentials during transient focal cerebral ischemia in cats to compare the effects of four commonly used anesthetic regimens: ketamine/fentanyl/N2O (fentanyl), pentobarbital, ketamine/α-chloralose (α-chloralose), and ketamine/halothane/N2O (halothane). Six cats in each group were subjected to 60 minutes of left middle cerebral artery occlusion followed by 120 minutes of reperfusion. Although the amplitude of the initial somatosensory evoked potential wave complex was highest in the α-chloralose group (58.6 ± 16.5 μV) and smallest in the halothane group (27.5 ± 5.7 μV), amplitude fell by 75% in all groups upon occlusion. Baseline cerebral blood flow varied substantially between groups (e.g., in the right intersylvian gyrus: fentanyl, 96 ± 12; pentobarbital, 30 ± 6; α-chloralose, 24 ± 3; and halothane, 76 ± 11 ml/min/100 g). Occlusion decreased cerebral blood flow to subcortical (e.g., left caudate) structures in all groups (fentanyl, 29 ± 11%; pentobarbital, 45 ± 12%; α-chloralose, 27 ± 13%; and halothane, 18 ± 5% of baseline). Postischemic hyperemia occurred in the cortical regions of cats anesthetized with pentobarbital or α-chloralose that had reduced cerebral blood flows during occlusion but not in cats anesthetized with fentanyl (cerebral blood flow during occlusion not different from that of cats anesthetized with pentobarbital or α-chloralose) or halothane. After 120 minutes of reperfusion, cerebral blood flow had returned to baseline values in all groups. Recovery of cerebral blood flow and somatosensory evoked potential amplitude at that time did not differ among groups. We conclude that anesthetics alter baseline cerebral blood flow and baseline somatosensory evoked potentials as well as the cerebral blood flow pattern during reperfusion after middle cerebral artery occlusion independent of insult severity. (Stroke 1990;21:795-800)
Materials and Methods

We used 24 adult female cats weighing 2.6–3.6 kg. Each cat was randomly assigned to one of four anesthetic groups. The six cats in the fentanyl group were induced with 50 mg/kg i.m. ketamine and 0.02 mg/kg i.m. acepromazine and maintained on 3–5 μg/kg/min i.v. fentanyl and 70% N2O in 30% O2. The six cats in the pentobarbital group were induced with 30 mg/kg i.p. pentobarbital and maintained on 5 mg/kg/hr i.v. pentobarbital. The six cats in the α-chloralose group were induced with 50 mg/kg i.m. ketamine and maintained on α-chloralose (50 mg/kg i.v. bolus and 25 mg/kg/hr i.v. infusion) and 70% N2O in 30% O2 until the end of the surgical preparation, when the N2O was replaced with N2. The six cats in the halothane group were induced with 50 mg/kg i.m. ketamine and 0.02 mg/kg i.m. acepromazine and maintained on <0.75% halothane in 70% N2O and 30% O2.

After induction of anesthesia, all cats were orally intubated and mechanically ventilated with a small animal ventilator (Model 661, Harvard Apparatus, South Natick, Massachusetts) to achieve normocapnia. Supplemental O2 was administered to avoid hypoxia. Mean±SEM rectal temperature was maintained at 38±0.5°C using a heating pad and heat lamp. A catheter was placed in the femoral vein and advanced into the inferior vena cava for the administration of fluids and drugs. A catheter was inserted into the descending aorta via a femoral artery for monitoring mean arterial blood pressure. A catheter was placed in the left atrium via a left thoracotomy for the injection of radiolabeled microspheres. A catheter was placed in the descending aorta via a femoral artery for withdrawal of the reference blood sample during microsphere injection. After the insertion of all catheters, the cat was turned prone and its head was stabilized so that the external auditory meatus was approximately 5 cm above the level of the heart.

Arterial blood pressure was monitored continuously with a Statham P-23 pressure transducer (Oxnard, California) and recorded on a Gould Brush recorder (Cleveland, Ohio). PaO2, PaCO2, and arterial pH were measured with Radiometer BMS3 electrodes and analyzer (Copenhagen, Denmark). Oxygen content and saturation and hemoglobin concentration were determined with a CO-oximeter (model 282, Instrumentation Laboratories, Lexington, Massachusetts).

Regional CBF (rCBF) was measured using the radiolabeled microsphere technique.15,17 For each measurement, approximately 105 microspheres (15±1.0 μm in diameter; Du Pont–New England Nuclear Products, Boston, Massachusetts) were injected into the left atrium and the reference blood sample was withdrawn simultaneously at a rate of 1.94 ml/min from the femoral artery. At the end of each experiment, the cat was killed with KCl and the brain was removed for processing. After formalinization, the brain was sectioned to determine rCBF to the right and left caudate nucleus, the brainstem, the right and left thalamus, the right and left insylvian and ectosylvian gyri (cortex in the MCA distribution), the right and left lingula gyri (cortex in the posterior cerebral artery distribution), and the right and left precruciate gyri (cortex in the anterior cerebral artery distribution).

Somatosensory evoked potentials and brainstem auditory evoked responses were measured and recorded as previously described.15,17 Briefly, stimulating needle electrodes were placed in each cat percutaneously in the vular surface of both forelegs in a location that caused a distinct digital twitch. The stimulus lasted 200 μsec; 256 stimuli were delivered at a rate of 5.9/sec and averaged. Upper and lower band-pass filters were 5 and 1,500 Hz, respectively. High-amplitude electrical artifacts were automatically rejected by the computer. The peripheral nerve was stimulated only for the purpose of data collection (approximately 45 seconds for each measurement). The active electrode and reference system yields a consistent waveform in cats. The waveform complex consists of a small positive wave (P2) followed by a large negative wave (N2) and a large positive wave (P3). The amplitude of the initial complex (P,N2) was measured and recorded for each time. Baseline latencies of N2 were anesthetic-dependent (for fentanyl 11–15, for pentobarbital 13–18, for α-chloralose 15–19, and for halothane 12–15 msec).

We measured brainstem auditory evoked responses to document that MCA occlusion changed the amplitude of the somatosensory evoked potential independent of changes in electrical transit through the brainstem. A small earpiece connected to an NIC-1007 click stimulus generator (Nicolet Instruments, Madison, Wisconsin) was secured in the auditory canal of each cat. Alternating clicks at a rate of 11.9/sec for 200 msec were used as the sound stimulus. There were 256 measurements lasting 10 msec each. The stimulus intensity was 95 dB for each measurement.

In each group CBF, somatosensory evoked potentials, brainstem auditory evoked responses, and blood gases were measured before MCA occlusion (baseline, at least 30 minutes after the end of surgery), after 30 and 60 minutes of ischemia, and after 15, 60, and 120 minutes of reperfusion. To initiate focal cerebral ischemia, a Weck micro–cerebral
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**FIGURE 1.** Graph of regional cerebral blood flow to left (L) caudate nucleus of cats before, during, and after L middle cerebral artery occlusion (LMCAO). Cats were anesthetized with fentanyl (o, n=6), pentobarbital (□, n=6), α-chloralose (△, n=6), or halothane (○, n=6). *p<0.05 different from time 0 for all groups at 30 minutes and for fentanyl, α-chloralose, and halothane groups at 60 minutes.

Aneurysm clip (Edward Weck, Inc., Research Triangle, North Carolina) was placed on the left MCA via a transorbital approach. Briefly, the left orbit was exonerated and the bone surrounding the optic foramen was removed using a diamond bit on a high-speed water-cooled drill. The dura was carefully removed, and the MCA was exposed close to its origin from the internal carotid artery. Correct clip placement was assumed when a >75% reduction in P_{\text{N}} amplitude occurred within 5 minutes. After 60 minutes of ischemia, reperfusion was initiated by removing the clip from the MCA.

All values are reported as the mean±SEM. Because CBF data sometimes had a skewed distribution and the SD varied among times and among groups, the CBF data were subjected to logarithmic transformation before testing for differences as a function of time within each group. Repeated-measures analysis of variance (ANOVA) was used to define differences in each measurement over time within each group. One-way ANOVA was used to define the effect of anesthetic at each time for each measurement between groups. The Newman-Keuls test was used for post-hoc comparisons. Unless otherwise specified, p<0.05.

**Results**

Although there were small changes in Paco\textsubscript{2} (for fentanyl), Pao\textsubscript{2} (for pentobarbital and α-chloralose), and hemoglobin concentration (for fentanyl and pentobarbital) during the experiment, there were no differences among groups at any time for arterial pH (7.32–7.36), Paco\textsubscript{2} (32–35 mm Hg), Pao\textsubscript{2} (115–135 mm Hg), hemoglobin concentration (9.5–11.3 g/dl), or mean arterial blood pressure (80–107 mm Hg).

Anesthetics affected baseline CBF. Global CBF was higher for the fentanyl (66±5 ml/min/100 g) and halothane (63±6 ml/min/100 g) groups than for the pentobarbital (27±4 ml/min/100 g) and α-chloralose (28±1 ml/min/100 g) groups.

MCA occlusion reduced CBF variably, depending on the anesthetic used. rCBF to the left caudate nucleus decreased significantly for all four groups (Figure 1). On the contrary, rCBF to the intersylvian gyrus decreased significantly only in the fentanyl, pentobarbital, and halothane groups (Figure 2). Absolute rCBF to the intersylvian gyrus during occlusion was higher for the halothane group than for the other groups, but there was no difference among the fentanyl, α-chloralose, and pentobarbital groups.

MCA occlusion also significantly decreased rCBF to the left ectosylvian gyrus for the fentanyl, α-chloralose, and halothane groups (Figure 3), the left lingula gyrus for all four groups (data not shown), the left precruciate gyrus for the pentobarbital, α-chloralose, and halothane groups (data not shown), and the left thalamus for the fentanyl and halothane groups (data not shown). rCBF to the brainstem and the right-sided brain regions were not decreased by MCA occlusion (data not shown).

Five of six cats in the pentobarbital group and four of six in the α-chloralose group had reduced rCBF to the intersylvian gyrus during MCA occlusion (for

**FIGURE 2.** Graph of regional cerebral blood flow to left (L) intersylvian gyrus of cats before, during, and after L middle cerebral artery occlusion (LMCAO). Cats were anesthetized with fentanyl (o, n=6), pentobarbital (□, n=6), α-chloralose (△, n=6), or halothane (○, n=6). *p<0.05 different from time 0 for fentanyl, pentobarbital, and halothane groups at 30 and 60 minutes.

**FIGURE 3.** Graph of regional cerebral blood flow to left (L) ectosylvian gyrus of cats before, during, and after L middle cerebral artery occlusion (LMCAO). Cats were anesthetized with fentanyl (o, n=6), pentobarbital (□, n=6), α-chloralose (△, n=6), or halothane (○, n=6). *p<0.05 different from time 0 for fentanyl, α-chloralose, and halothane groups at 30 and 60 minutes.
pentobarbital 32±19% and for α-chloralose 22±19% of baseline). All nine of these cats demonstrated reactive hyperemia during reperfusion (for pentobarbital 697±122% and for α-chloralose 801±492% of baseline). After 15 minutes of reperfusion cats in the pentobarbital and halothane groups did not demonstrate hyperemia relative to their high baseline values.

In all regions, rCBF eventually returned to baseline values. In the pentobarbital, α-chloralose, and halothane groups, rCBF returned to baseline in all regions by 60 minutes of reperfusion. In the fentanyl group, rCBF in the left thalamus did not return to baseline until 120 minutes of reperfusion; it had returned to baseline in all other regions by 60 minutes of reperfusion. No group demonstrated hyperemia at 120 minutes of reperfusion. Expressed as a percentage of baseline, there were no differences among groups in the recovery of rCBF for any region at 120 minutes of reperfusion.

The latencies of waves I, III, and V of the brainstem auditory evoked response were well controlled, and MCA occlusion was proximal to perforators and thus will not produce the same distribution of ischemia as would a more proximal occlusion. The presence of hyperemia did not correlate with the severity of ischemia since cats anesthetized with fentanyl had CBF reduced to the same extent as cats anesthetized with pentobarbital or α-chloralose but did not exhibit hyperemia. No group demonstrated delayed hypoperfusion after 120 minutes of reperfusion. There were no differences in the recovery of somatosensory evoked potential amplitude among the groups.

Somatosensory evoked potentials have been used both clinically and experimentally to noninvasively measure the adequacy of CBF. We used the forepaw somatosensory evoked potential as a noninvasive indicator that MCA occlusion reduced CBF.

The extent of CBF diminution with MCA occlusion and the exact distribution of ischemia is the subject of considerable disagreement. Important potential modulators of the CBF response to MCA occlusion and reperfusion are systemic factors, differences in anesthetics, and differences in location of the MCA clip. For example, it is likely that a clip on the distal MCA will not occlude the lenticulostriate perforators and thus will not produce the same distribution of ischemia as would a more proximal occlusion. In our study, physiologic parameters were well controlled, and MCA occlusion was proximal to the lenticulostriate arteries in all cats so that we could evaluate the role of anesthetics alone.

In spite of reduced amplitudes of the somatosensory evoked potential during reperfusion, latencies of the brainstem auditory evoked response were unchanged from baseline. This suggests that the etiology of somatosensory evoked potential amplitude reduction results from higher brain structures. Cortical (gray matter) rCBF levels below which amplitude is substantially (>25%) reduced have been reported to be in the range of 6–16 ml/min/100 g. In our study, we achieved such values in the cortex supplied by the MCA with all anesthetics except halothane. Several investigators have suggested that the loss of somatosensory evoked potentials during MCA occlusion correlates better with subcortical (thalamic,
white matter) than with cortical ischemia. Likewise, in our studies, a reduction in somatosensory evoked potential amplitude to 25% of baseline was closely associated with a reduction in subcortical (i.e., caudate nucleus) but not cortical rCBF. rCBF to the thalamus was reduced during MCA occlusion in cats anesthetized with fentanyl or halothane but not pentobarbital or α-chloralose.

Isolated MCA occlusion causes partial focal cerebral ischemia because collateral vessels continue to provide blood flow. The fact that MCA occlusion caused larger reductions in subcortical than in cortical rCBFs may indicate a more extensive collateral circulation in the cortex rather than a decreased vulnerability to ischemia of gray than white matter.23 Other physiologic variables that may affect this model (such as PaCO₂, hemoglobin concentration, and body temperature24) were well controlled.

We chose a model of transient focal ischemia with reperfusion over one of permanent ischemia so that we could evaluate the pattern of reperfusion with different anesthetics. We did not consistently observe reactive hyperemia followed by hypoperfusion, which has been reported in models of transient global ischemia.23 Some investigators have demonstrated reactive hyperemia after 1 hour of MCA occlusion in animals anesthetized with pentobarbital23 or α-chloralose2 but not with different durations of ischemia.26 Other investigators, however, have been unable to demonstrate hyperemia following any duration of focal ischemia.22 In our study, five of six cats anesthetized with pentobarbital and four of six anesthetized with α-chloralose had reduced cortical rCBF after 60 minutes of ischemia; all nine cats demonstrated hyperemia during reperfusion. It is possible that our inability to demonstrate reactive hyperemia in cats anesthetized with fentanyl or halothane relates to the times we chose to measure CBF. Specifically, reactive hyperemia may occur before 15 minutes of reperfusion and, therefore, we may have missed it. As in our study, others have demonstrated that in cats, reactive hyperemia (when present) is sustained for at least 15 minutes of reperfusion following either global28 or focal26 ischemia.

One possible explanation for the lack of hyperemia in cats anesthetized with fentanyl or halothane is their high baseline CBF. One proposed mechanism for hyperemia after focal ischemia is an alteration in the ratio of blood pressure to hematocrit. This explanation is supported by Coyer et al,4 who demonstrated that volume expansion and hemodilution provided a means of significantly elevating rCBF in the gray matter. However, this explanation cannot account for our data, for we maintained blood pressure and hemoglobin concentration constant in all groups. Delayed hypoperfusion did not occur with any of our anesthetics and has not been demonstrated by other experimenters after transient focal ischemia.23,22,26 Likewise, when CBF was expressed as a percentage of baseline values, there was no difference in recovery among groups. Differences among groups in absolute CBF values during reperfusion can be accounted for by previously described effects of anesthetics on CBF. For example, animals anesthetized with halothane have greater CBFs than animals anesthetized with pentobarbital.7,11

In summary, our data suggest that monitoring somatosensory evoked potentials is more useful in diagnosing subcortical than cortical ischemia in the distribution of the MCA. It is clear from our data that the regional cerebrovascular response to transient MCA occlusion depends on the anesthetics used. This may make it difficult to compare studies from different laboratories unless the studies were done in an identical fashion. Differences in the exact distribution of CBF reductions during MCA occlusion (more regions have decreased rCBFs with fentanyl and halothane) and the presence of hyperemia during reperfusion may be due to different baseline CBFs but does not appear to be due to differences in the intensity of the ischemic insult among groups.

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References

4. Coyer PE, Michele JJ, Lesnick JE, Simeone FA: Cerebral blood flows and tissue oxygen levels associated with maintenance of the somatosensory evoked potential and cortical neuronal activity in focal ischemia. Stroke 1987;18:77–84


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M A Helfaer, J R Kirsch and R J Traystman

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