Acute Toxicity of a Nuclear Magnetic Resonance Cerebral Blood Flow Indicator in Cats

Craig A. Branch, PhD, James R. Ewing, MS, Susan C. Fagan, PharmD, David A. Goldberg, MD, and K.M.A. Welch, MD

We studied trifluoromethane as a potential gaseous indicator in nuclear magnetic resonance measurements of cerebral blood flow. We considered the effects of trifluoromethane on cerebral blood flow in 17 cats and on the electroencephalogram and electrocardiogram in nine cats and compared these with the effects of the more toxic compound chlorodifluoromethane in five cats. Inhaled at 60%, trifluoromethane had no effect on cerebral blood flow, the cerebral metabolic rate for oxygen, or oxyhemoglobin content. At 70%, trifluoromethane sensitized the cats' hearts to epinephrine, but to a much lesser degree than 40% chlorodifluoromethane, and produced only moderate changes in cerebral electrical activity as measured by the electroencephalogram. We found trifluoromethane to be suitable for use in animals, but its toxicity needs to be studied further before it can be used in humans for the measurement of cerebral blood flow. (Stroke 1990;21:1172-1177)

Gaseous halocarbons have great potential in nuclear magnetic resonance (NMR) indicator-dilution measurements of cerebral blood flow (CBF). However, in animals acute exposure to halocarbons can alter pulmonary compliance and resistance, can depress myocardial contractility, arterial blood pressure, and coronary blood flow, and can sensitize the myocardium to epinephrine-induced arrhythmias. Extended exposure can cause pulmonary hemorrhage and congestion. A sparse literature indicates that these effects are apparently absent for trifluoromethane (FC-23). One report demonstrated no change in cardiac sensitivity to epinephrine during FC-23 inhalation, although small changes in sensitivity might not have been discerned because of the large doses of epinephrine administered. In another report no toxic symptoms were observed in dogs, although one author stated that during inhalation of FC-23 he experienced "definite analgesia and impairment of consciousness."

Our initial studies of CBF measured with NMR in cats employed chlorodifluoromethane (CFC-22) because its high blood solubility, relatively low toxicity, and low cost were well suited to the development of NMR methodologies. With these methodologies proven, CFC-22 was discarded in favor of FC-23 because we found that 40% CFC-22 increased CBF by approximately 50%. Despite an additional fluorine atom in FC-23, its signal-to-noise ratio in NMR experiments was poorer than that of CFC-22 because it is roughly one third as soluble as the latter in blood. Thus, it was necessary to administer FC-23 at the highest possible concentration to attain bicompartmental curve fits to the clearance data so that relative weights of the cerebral compartments could be estimated.

Recently we reported multicompartmental NMR measurements of CBF using FC-23 as an indicator, along with the preliminary finding that FC-23 had no effect on CBF in nine cats. Because of the limited statistical significance of our results using so few animals, as well as because of recent reports to the contrary, we studied CBF and the cerebral metabolic rate for oxygen (CMRO₂) in additional cats. We also studied the acute cardiac toxicity of FC-23 and its influence on the electroencephalogram (EEG) when administered at the concentration and duration required for NMR measurement of CBF. For comparison, we also evaluated the effects of CFC-22.
Materials and Methods

We studied the effect of 60% FC-23 on CBF in 17 cats using the Kety-Schmidt N2O clearance technique.23,24 Cats were anesthetized with 1.5 mg/kg i.m. xylazine and 15 mg/kg i.m. ketamine, tracheotomized, paralysed with 0.08 mg/kg i.v. pancuronium bromide, and mechanically ventilated. A femoral artery and vein were cannulated for measurements of arterial blood pressure, blood gases, and N2O content and for the administration of fluids, respectively. The sagittal sinus was cannulated rostrally with a 24-gauge catheter for measurements of cerebral venous N2O and O2 contents.

Each measurement of CBF was preceded by the administration of 5 mg/kg i.v. thiopental sodium 5 minutes before the administration of 2 mg/kg i.v. ketamine. This combination produces a stable CBF approximately 80% of normal levels.1,24 Cats were ventilated with 30% O2+5% N2O+65% N2 for 1 hour before the measurement of control CBF by N2O clearance. The N2O was then replaced with N2, and the measurement of CBF began. The measurement of CBF during FC-23 inhalation was accomplished by replacing 60% of the N2 with FC-23 10 minutes before and during the N2O clearance period. Arterial and cerebral venous blood samples (0.3 mL aliquots) were taken before and at 0.5, 1, 1.5, 2, 3, 4, 5, 7, 9, 12, 15, 30, and 45 minutes after N2O clearance into 1-mL plastic syringes. Each sample was immediately sealed, packed in crushed ice, and analyzed within 30 minutes for N2O content in a (modified) trace N2O monitor (Sensors Inc.; Traverse Medical Monitors, Saline, Mich.). We calculated CBF using a standard, model-independent approach.23,24 We assumed a blood:brain partition coefficient of 1.08 for N2O. We monitored PaO2, PaCO2, and arterial pH, and if necessary we adjusted the respiratory parameters to maintain gas tensions constant throughout each CBF measurement. Blood withdrawn for N2O determinations was replaced between CBF measurements, and the order of FC-23 administration was alternated to control for order effects. We analyzed the CBF data using repeated-measures analysis of covariance designed to detect a change in CBF of 20% from control with an α of 0.25 and a power of 0.9 (β of 0.10).

In 10 of these 17 cats, the effect of 60% FC-23 on cerebral venous and arterial oxygen saturation and hemoglobin concentration were measured with a CO-oximeter (model IL482, Instrumentation Laboratory, Lexington, Mass.) at three times during each CBF measurement, and we calculated the oxyhemoglobin content and CMRO2. We evaluated changes in oxyhemoglobin content and CMRO2 using paired t tests.

In nine additional cats, we studied the central nervous system (CNS) and central circulatory effects of 70% FC-23. These cats were prepared similarly to those in which CBF was measured, but the sagittal sinus was not cannulated. Anesthesia and paralysis were achieved with 2 mg/kg ketamine and 0.06 mg/kg pancuronium bromide administered every 30 minutes. Lead II EEG, electrocardiogram (ECG), and cardiac sensitivity to epinephrine were studied during the control condition (inhalation of 30% O2+70% N2) and during inhalation of 70% FC-23; in five of these 9 cats these measurements were repeated during the inhalation of 40% CFC-22. Each study followed a ketamine injection by 15 minutes, and the administration of FC-23 or CFC-22 began 10 minutes before the study. EEG, ECG, and blood pressure were recorded before, during, and after the infusion of 0.1 μg/kg epinephrine in two cats (both received CFC-22) or 1 μg/kg epinephrine in seven cats (three received CFC-22) over 10 seconds. We analyzed the changes in response to epinephrine using a paired t test. Referential EEG was monitored in the right hemisphere via frontal, occipital, and lateral subdural needle electrodes. All records were visually reviewed, and in the last five cats studied we analyzed 20–30 4-second epochs of EEG using a fast-Fourier transform algorithm and averaged the spectral power into five frequency “bins” (0–4 [delta], 4–8 [theta], 8–13 [alpha], 13–30 [beta], and 30–60 [gamma] Hz). To each bin we added 1.0 and transformed the result using the natural logarithm. We compared spectral power between the control and FC-23 conditions using a paired t test.

Results

Kety-Schmidt N2O clearance measurements of CBF in 17 cats are presented in Table 1. Control CBF did not differ significantly from that during inhalation of 60% FC-23 either before (p>0.25) or after (p>0.43) adjusting for PaCO2. No significant differences were detected in CMRO2 or arterial oxyhemoglobin content between the control and FC-23 conditions (p>0.5).

Visual review of the EEG tracings from all nine cats revealed increased activity in the theta bin after 10 minutes of 70% FC-23 inhalation (Figure 1). Theta and alpha activity in both the frontal and occipital leads increased significantly (p<0.05) during FC-23 inhalation (Figure 2). Spectral power decreased nonsignificantly in the frontal and lateral (data not shown) leads at higher frequencies. In two cats, 10 minutes of 40% CFC-22 inhalation produced suppression-burst activity indicative of deep anesthesia. In the other three cats, signal amplitude above

<p>| Table 1. Effect of 60% FC-23 on CBF and CMRO2 in Cats by Kety-Schmidt N2O Clearance |
|---------------------------------|-----------------|-----------------|</p>
<table>
<thead>
<tr>
<th>Condition</th>
<th>CBF (ml/100 g/min)</th>
<th>PaCO2 (mm Hg)</th>
<th>CMRO2 (ml/100 g/min)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control</td>
<td>(n=17)</td>
<td>(n=17)</td>
<td>(n=17)</td>
</tr>
<tr>
<td>38±10</td>
<td>31.6±2.8</td>
<td>3.3±0.9</td>
<td></td>
</tr>
<tr>
<td>FC-23</td>
<td>41±13</td>
<td>32.3±3.3</td>
<td>3.4±0.8</td>
</tr>
</tbody>
</table>

FC-23, trifluoromethane; CBF, cerebral blood flow; CMRO2, cerebral metabolic rate for oxygen. Data are mean±SD.
FIGURE 1. Trends in electroencephalogram recorded from referential occipital (left) and frontal (right) leads during control condition (above) and inhalation of 70% trifluoromethane (FC-23) (below) in cats. Increased theta (4–8 Hz) and alpha (8–13 Hz) activity are apparent in either lead during FC-23 inhalation.

approximately 4 Hz was greatly decreased (not shown).

Heart rate before epinephrine, arterial blood pressure, and the pressor response to 1 µg/kg epinephrine during 70% FC-23 inhalation did not differ significantly from that during the control condition (Table 2). Inhalation of 40% CFC-22 significantly decreased both blood pressure ($p<0.05$) and the pressor response to epinephrine ($p<0.05$). Infusion of 0.1 µg/kg epinephrine in two cats produced little or no effect on blood pressure or heart rate during the control condition or inhalation of 70% FC-23 or 40% CFC-22. No arrhythmias were induced by this dose of epinephrine during either the control condition or inhalation of 70% FC-23. However, during 40% CFC-22 inhalation in one cat 0.1 µg/kg epinephrine elicited a severe arrhythmic reaction (Table 3).

During the control condition, none of seven cats experienced any abnormal alterations in ECG after 1.0 µg/kg epinephrine infusions. The characteristic response to this dose of epinephrine consisted of an initial rise in heart rate, followed by a rise in blood pressure and subsequent bradycardia, with blood pressure gradually returning to normal. During 70% FC-23 inhalation, 1.0 µg/kg epinephrine elicited variable responses (Table 3). Three cats responded as during the control condition, one demonstrated a mild response, and three suffered severe arrhythmic responses (Figure 3). In general, during inhalation of 70% FC-23 arrhythmia in response to epinephrine was transient, lasting only several seconds, in contrast to that during 40% CFC-22 inhalation, when arrhythmia lasted up to 10 times as long.

Two cats suffered prolonged periods of arrhythmia after the withdrawal of CFC-22 (Figure 4). Approximately 1 minute after discontinuing the CFC-22, both animals began a period of spontaneous multifocal ventricular tachycardia with ventricular group beats lasting 2–3 minutes. The onset of arrhythmia coincided with the rise in blood pressure to near control levels.

Discussion

In cats, CFC-22 increases CBF by 50%, eliminating this agent as a possible NMR indicator. In

<table>
<thead>
<tr>
<th>Condition</th>
<th>Control</th>
<th>70% FC-23</th>
<th>40% CFC-22</th>
</tr>
</thead>
<tbody>
<tr>
<td>Heart rate (min⁻¹)</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Change</td>
<td>60±30</td>
<td>58±23</td>
<td>Obscured by arrhythmia</td>
</tr>
<tr>
<td>Blood pressure (mm Hg)</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Systolic</td>
<td>139±22</td>
<td>134±21</td>
<td>95±25*</td>
</tr>
<tr>
<td>Change</td>
<td>62±27</td>
<td>61±22</td>
<td>42±6.3*</td>
</tr>
<tr>
<td>Diastolic</td>
<td>111±20</td>
<td>105±21</td>
<td>74±26*</td>
</tr>
<tr>
<td>Change</td>
<td>36±20</td>
<td>35±18</td>
<td>34±10</td>
</tr>
</tbody>
</table>

FC-23, trifluoromethane; CFC-22, chlorodifluoromethane.

*p<0.05 different from control by paired t test.
Our EEG data indicate that inhalation of 70% FC-23 causes a mild CNS stimulatory effect (increased power in the theta and alpha bins) while possibly depressing power at higher frequencies. However, the variance of our data is great due to the few cats studied. To better define the effect of FC-23 on the CNS, a more complete study of the FC-23 dose–response relation for EEG needs to be undertaken. We are currently studying this effect further in primates.

Some inhalational anesthetics induce cardiac arrhythmias, and halocarbons increase cardiac sensitivity to the arrhythmogenic potential of epinephrine. Lesions within the pulmonary or cardiovascular systems also increase the potential for arrhythmias during the inhalation of halocarbons. Since an NMR technique to measure CBF may eventually be used in patients with strokes or other disease states that might elevate endogenous catecholamine levels, the tendency of these halocarbons to increase cardiac sensitivity to epinephrine is of great importance. With these factors in mind, we investigated the sensitizing potential of FC-23 in cats.

Except for elevating the ST segment of the ECG, the response to epinephrine during 70% FC-23 inhalation appears to be an enhancement of typical epinephrine-induced cardiac activity. We observed ST segment elevations only following epinephrine infusions during inhalation of 70% FC-23 or 40% CFC-22. Such ST segment elevations have also been observed during inhalation of other halocarbons and probably indicate myocardial ischemia. Generally, halocarbons are believed to influence coronary blood flow by reducing left ventricular perfusion pressure and opposing sympathetic and parasympathetic activities in the systemic arterial bed. A similar mechanism has been proposed to explain the depression of coronary blood flow following exposure to methylene chloride.

Branch et al

Trifluoromethane Toxicity in Cats

Table 3. Responses Elicited by Epinephrine Infusions During Inhalation of FC-23 or CFC-22 by Cats

<table>
<thead>
<tr>
<th>Cat</th>
<th>Fluorocarbon</th>
<th>Response</th>
</tr>
</thead>
<tbody>
<tr>
<td>0.1 μg/kg epinephrine</td>
<td></td>
<td></td>
</tr>
<tr>
<td>1</td>
<td>FC-23</td>
<td>Normal sinus rhythm</td>
</tr>
<tr>
<td>2</td>
<td>CFC-22</td>
<td>Normal sinus rhythm</td>
</tr>
<tr>
<td>2</td>
<td>FC-23</td>
<td>Normal sinus rhythm</td>
</tr>
<tr>
<td>3</td>
<td>CFC-22</td>
<td>VPCs, ventricular bigeminy, elevated ST</td>
</tr>
<tr>
<td>1.0 μg/kg epinephrine</td>
<td></td>
<td></td>
</tr>
<tr>
<td>3</td>
<td>FC-23</td>
<td>VPCs, elevated ST</td>
</tr>
<tr>
<td>4</td>
<td>CFC-22</td>
<td>VPCs, multifocal VT, elevated ST</td>
</tr>
<tr>
<td>4</td>
<td>FC-23</td>
<td>VPCs, fusion beats, JER, elevated ST, sinus bradycardia</td>
</tr>
<tr>
<td>5</td>
<td>CFC-22</td>
<td>VPCs, ventricular bigeminy</td>
</tr>
<tr>
<td>6</td>
<td>FC-23</td>
<td>Normal sinus rhythm</td>
</tr>
<tr>
<td>7</td>
<td>FC-23</td>
<td>1 VPC, elevated ST</td>
</tr>
<tr>
<td>7</td>
<td>CFC-22</td>
<td>VPCs, sinus bradycardia, JER, elevated ST</td>
</tr>
<tr>
<td>8</td>
<td>FC-23</td>
<td>Normal sinus rhythm</td>
</tr>
<tr>
<td>9</td>
<td>FC-23</td>
<td>VPCs, VT</td>
</tr>
</tbody>
</table>

Contrast, in 17 cats we found no evidence that 60% FC-23 alters CBF, nor did it affect CMRO₂ in 10 cats. To our knowledge, there exists only one other report of the effect of FC-23 on CBF. That report studied regional CBF using microspheres in six cats before and during inhalation of 70% FC-23 and suggested that regional CBF in gray tissues increased while that in white tissues decreased during inhalation of FC-23. Opposing trends could result in no change in mean CBF as measured with the Kety-Schmidt N,O clearance technique or the increase in CBF observed during inhalation of 70% FC-23 could be absent during inhalation of 60% FC-23, although we find neither possibility likely. It is more likely that the effect of a small sample size, combined with order and anesthetic effects had greater influence on those results. Apparently, the potential for FC-23 to cause regional changes in CBF needs to be examined more closely.
respiratory irritation, then any alterations in vagal tone could potentiate arrhythmias. Thus, the acute rise in blood pressure following epinephrine infusion, which causes a reflex increase in vagal tone, leads to the increased incidence of arrhythmia we observed.

Pretreatment of animals with β-blockers decreases cardiac sensitivity to halocarbons, although arrhythmias still occur at higher concentrations of the more toxic halocarbons. Halothane directly stimulates adrenergic β receptors in bronchial smooth muscle, and it is likely that trichlorofluoromethane does so too. Trichlorofluoromethane induces peripheral vasodilation, while CFC-22 may affect smooth muscle tone, as evidenced by its effect on CBF. These findings suggest that some halocarbons directly stimulate β receptors.

The absence of cardiac arrhythmogenic tendencies in healthy animals does not necessarily clear a halocarbon for use as an NMR indicator. Rabbits exposed to 30% or 40% CFC-22 for extended periods demonstrate subpleural hemorrhage, excessive mucus, and evidence of pulmonary emphysema and congestion. However, at low inspiration levels for short periods no irreversible effects were seen. Similar observations have been made in our laboratory following extended periods of 40% CFC-22 inhalation. However, animals inhaling 60% FC-23 for as long as 90 minutes failed to exhibit pulmonary damage.

Rats respond to chlorofluorocarbons with a decrease in heart rate, while dogs and primates respond to the same agents with increases in heart rate. This effect is probably attributable to the occurrence of α receptors in the rat heart and represents a species-specific effect. On the other hand, cardiac arrhythmias can be elicited in dogs breathing 5% CFC-22 but not in monkeys or mice breathing 20% CFC-22. This type of variability probably represents a difference in species sensitivity to the gases, a probable contributing factor to which is the difference in

![Figure 3](https://stroke.ahajournals.org/)

**Figure 3.** Electrocardiogram (above) and arterial blood pressure (below) from two representative cats during control condition (left) and during inhalation of 70% trifluoromethane (70% FC-23) before (center) and after (right) infusion of 1 μg/kg epinephrine. Following epinephrine infusion in one cat (top) 20-second period of arrhythmia characterized by ventricular premature contractions and elevated ST segment occurred. Similar infusions of epinephrine before and after FC-23 inhalation in this cat were uneventful. In the other cat (bottom) 30-second arrhythmia consisted of ventricular premature contractions alternating with sinus beats and elevated ST segment.

![Figure 4](https://stroke.ahajournals.org/)

**Figure 4.** Electrocardiogram (above) and arterial blood pressure (below) in cat shown in top of Figure 3 immediately after discontinuing 40% chlorodifluoromethane (CFC-22) inhalation. Ventricular group beats appeared 1 minute after switch to 30% O2 + 70% N2 and continued for 2 minutes. Even after 6 minutes, isolated ventricular premature contractions were still observed.
species sensitivity to epinephrine. Since FC-23 is a demonstrated (but relatively mild) cardiac sensitizing agent, further study of this sensitizing potential (preferably in primates) are necessary before administering it to humans who may already suffer from vascular or pulmonary disease.

Acknowledgments

We wish to gratefully acknowledge Dr. Robert Simkins for his assistance in interpreting our electroencephalography records and Mr. Shazad Butt for his extensive technical assistance.

References

30. Govier WC: Myocardial alpha adrenergic receptors and their role in the production of a positive inotropic effect by sympathomimetic agents. J Pharmacol Exp Ther 1968;159:82–90

Key Words • cerebral blood flow • nuclear magnetic resonance • cats
Acute toxicity of a nuclear magnetic resonance cerebral blood flow indicator in cats.
C A Branch, J R Ewing, S C Fagan, D A Goldberg and K M Welch

Stroke. 1990;21:1172-1177
doi: 10.1161/01.STR.21.8.1172

The online version of this article, along with updated information and services, is located on the World Wide Web at:
http://stroke.ahajournals.org/content/21/8/1172

Permissions: Requests for permissions to reproduce figures, tables, or portions of articles originally published in Stroke can be obtained via RightsLink, a service of the Copyright Clearance Center, not the Editorial Office. Once the online version of the published article for which permission is being requested is located, click Request Permissions in the middle column of the Web page under Services. Further information about this process is available in the Permissions and Rights Question and Answer document.

Reprints: Information about reprints can be found online at: http://www.lww.com/reprints

Subscriptions: Information about subscribing to Stroke is online at: http://stroke.ahajournals.org//subscriptions/