Brain Polyphosphoinositide Metabolism During Focal Ischemia in Rat Cortex

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Using a rat model of stroke, we examined the effects of focal cerebral ischemia on the metabolism of polyphosphoinositides by injecting $^{32}$Pi into both the left and right cortices. After equilibration of the label for 2-3 hours, ischemia induced a significant decrease ($p<0.001$) in the concentrations of labeled phosphatidyl 4,5-bisphosphates (66-78%) and phosphatidylinositol 4-phosphate (64-67%) in the right middle cerebral artery cortex of four rats. The phospholipid labeling pattern in the left middle cerebral artery cortex, which sustained only mild ischemia and no permanent tissue damage, was not different from that of two sham-operated controls. However, when $^{32}$Pi was injected 1 hour after the ischemic insult, there was a significant decrease ($p<0.01$) in the incorporation of label into the phospholipids in both cortices of four ischemic rats compared with four sham-operated controls. Furthermore, differences in the phospholipid labeling pattern were observed in the left cortex compared with the sham-operated controls. The change in labeling pattern was attributed to the partial reduction in blood flow following ligation of the common carotid arteries. We provide a sensitive procedure for probing the effects of focal cerebral ischemia on the polyphosphoinositide signaling pathway in the brain, which may play an important role in the pathogenesis of tissue injury. (Stroke 1991;22:495-498)
to the sagittal suture and 1.5 mm dorsal to the coronal suture for $^{32}$Pi injections into the right and left MCA cortices. The $^{32}$Pi (20 $\mu$Ci in the form of H$_2$PO$_4^-$, carrier free, 285 Ci/mg; New England Nuclear, Boston, Mass.) was delivered with a 10-μl microsyringe at a depth of 3 mm beneath the calvaria. The $^{32}$Pi injections were made at precise time intervals either before or after the ischemic insult.

The procedure to induce severe focal cerebral ischemia leading to a consistently large infarct in the right MCA cortex has been described in detail. Ligation of the right MCA and temporary occlusion of both CCAs for 30 minutes or longer consistently resulted in coagulation necrosis of the cortex in the right MCA territory but sparing of the subcortical structures and the left hemisphere. Cerebral blood flow was less than 20% of control in the ischemic right MCA cortex and approximately 60–80% of control in the left MCA cortex. At precise time intervals after the ischemic insult, the rats were decapitated and the heads were dropped into liquid nitrogen for rapid freezing of the brain tissue. The left and right MCA cortices were identified and punched out using cylindrical tubing 1 cm in diameter, and subcortical tissue was dissected out with a scalpel.

Brain tissues were homogenized in a buffer containing 0.32 M sucrose with 1 mM ethylenediaminetetraacetic acid and 50 mM Tris-HCl (pH 7.4). Aliquots of the tissue homogenate were taken for counting of radioactivity. Lipids from the tissue homogenate were extracted by a stepwise procedure with neutral and acidified chloroform:methanol (2:1, vol:vol) as described previously. Lipids were separated by the two-dimensional high-performance thin-layer chromatography procedure as described by Sun and Lin. After solvent development, lipid spots on the plate were visualized by exposure to iodine vapors and then scraped into scintillation vials for counting of radioactivity by a Beckman LS 5800 spectrometer (Carlsbad, Calif.). Due to variations in the amount of $^{32}$Pi delivered into the brain tissue, labeling of the phospholipids among individual samples was assessed by normalizing the radioactivity against that of phosphatidylethanolamine (PE). We have previously observed that, using this procedure to prelabel brain phospholipids, the labeling of PE was not affected by the ischemic insult. Another reason to use this procedure to assess the data is that labeled inositol phosphates released from poly-PI appear in the aqueous phase. Consequently, this leads to a net decrease of radioactivity in the organic phase and a concomitant increase in the aqueous phase. Data were evaluated by analysis of variance followed by a post hoc test for assessing the significance of the differences.

The first experimental protocol examined whether poly-PI in the MCA cortex were susceptible to hydrolysis due to decapitation-induced global cerebral ischemia. In this protocol, the right and left cerebral cortices were labeled with $^{32}$Pi and the label was allowed to equilibrate in the brain for 2.7 hours prior to the ischemic treatment. The prelabeled rats (five in each group) were decapitated and the head was either frozen immediately in liquid nitrogen or incubated at 37°C for 3 minutes (to simulate an ischemic condition) prior to freezing in liquid nitrogen. The right and left MCA cortices were obtained from each group for phospholipid analysis.

In the second experimental protocol, rats were injected with $^{32}$Pi 2 hours before the induction of focal cerebral ischemia by ligation of the right MCA and both CCAs for either 30 minutes ($n=4$) or 2 hours ($n=4$). Two sham-operated controls received $^{32}$Pi in a similar fashion and at identical time intervals except that the right MCA and CCAs were isolated but not occluded. After decapitation, both the left and right MCA cortices in the controls and ischemic rats were processed for analysis of the phospholipids.

In the third experimental protocol, $^{32}$Pi was injected after four rats had been subjected to focal cerebral ischemia for 1 hour. Four sham-operated controls and the ischemic rats were decapitated 1 hour after $^{32}$Pi labeling, and brain samples were analyzed as described above.

## Results

When the left and right cortices were prelabeled with $^{32}$Pi for 2.7 hours and the brain tissue was frozen immediately after decapitation, we observed no obvious differences in the phospholipid labeling pattern between the two MCA cortices (data not shown). There were some variations (approximately 35%) in the amount of label delivered into the brain tissue, but this variance did not alter the phospholipid labeling pattern. Nevertheless, 3 minutes of global cerebral ischemia elicited significant decreases compared with control in the labeling of PIP$_2$ (64%), phosphatidylinositol 4-phosphate (PIP) (58%), and phosphatic acids (60%) in both the left and right MCA cortices (Figure 1). In addition, some decreases in the labeling of phosphatidylcholines (12%) and ethanolamine plasmalogens (31%) were also observed in the ischemic brain.

The phospholipid labeling patterns obtained from the left and right MCA cortices of the sham-operated controls and rats that sustained focal cerebral ischemia by ligation of the right MCA and both CCAs for 30 minutes are shown in Figure 2, top. Two hours after the injection of $^{32}$Pi into the brain, labeled PE comprised approximately 7–8% of the total phospholipid radioactivity. There were no differences in the phospholipid labeling pattern between the left or right MCA cortices of the sham-operated controls and the left MCA cortex of the ischemic rats. However, 30 minutes of ischemia resulted in a significant decrease ($p<0.001$) in the labeling of PIP$_2$ (66%) and PIP (64%) in the ischemic right MCA cortex, which sustained severe injury. The labeling of other phospholipids in the ischemic right MCA cortex was not appreciably altered. When the duration of the ischemia was extended to 2 hours, there was a similar decrease in the labeling of PIP$_2$ and PIP (Figure 2, bottom) and a small but significant increase (143%).
FIGURE 1. Bar graph of effect of decapitation-induced global cerebral ischemia (crosshatched bars) on phospholipid labeling patterns in rat middle cerebral artery (MCA) cortices. $^{32}$Pi was injected as described in text, and label was equilibrated in brain for 2.7 hours prior to decapitation. See text for details of ischemic treatment and lipid analysis. Radioactivity of individual phospholipids are expressed against that of phosphatidylethanolamine (PE) as mean±SD (n=10, five rats in each group). Because there were no obvious differences in labeling patterns between left and right MCA cortices in either control (open bars) or ischemic brains, values are not presented separately. a, p<0.05 different from sham-operated controls by analysis of variance. Mean±SD radioactivity of lipid extract obtained from 20 MCA cortices (left and right) was 7.4±3.4×10⁵ dpm. PIP₂, phosphatidylinositol 4,5-bisphosphates; PIP, phosphatidylinositol 4-phosphate; PA, phosphatidic acids; PI, phosphoinositides; PC, phosphatidylcholines; PEP, ethanolamine plasmalogens.

in the labeling of phosphatidylinositols compared with that in the left MCA cortex and that in the sham-operated controls.

When the rats were subjected to focal cerebral ischemia for 1 hour prior to the injection of $^{32}$Pi, measurement of the radioactivity in brain homogenates from the MCA cortices of sham-operated controls and the left and right MCA cortices of ischemic rats gave values of 1.0±0.33×10⁶ (n=4), 1.8±0.46×10⁵ (n=6), and 7.3±4.1×10⁴ (n=5) cpm, respectively. Thus, although the left MCA cortex of the ischemic rats sustained only mild ischemia with no obvious pathological consequence, there was a significant decrease (p<0.01) in the uptake of labeled precursor during ischemia. When phospholipids in the left MCA cortex of the ischemic rats were analyzed, we were surprised to find twofold increases in the labeling of PIP and PIP₂ compared with the sham-operated controls (Figure 3). On the other hand, the labeling of poly-PI in the ischemic right MCA cortex showed large variances and was significantly less than that in the left MCA cortex.

Discussion

Previous studies from our laboratory⁴ as well as from others⁵ have demonstrated a rapid decrease in the level of poly-PI in brain during the early phase of global cerebral ischemia. In most instances, poly-PI degradation is marked by a concomitant increase in the concentrations of diacylglycerols⁵⁻⁶ and inositol phosphates,⁸ suggesting that poly-PI is hydrolyzed through the receptor-mediated phospholipase C pathway.² To facilitate transport of the labeled precursor to the MCA cortex, $^{32}$Pi was injected into both the left and right cortices. As indicated by the results of the first
precursor by the right MCA cortex, corroborating the brain cortex after ligation of the CCAs and right clearly indicate that pory-PI breakdown is a response large reduction of blood flow in the tissue. Although sham-operated controls) in uptake of the labeled specific to tissue sustaining severe ischemic insult. region was apparently not hydrolyzed. These results ligation of the left CCA), labeled poly-PI in this brain mild ischemia (20-40% reduction of blood flow due to ligated of the right MCA together with temporary ischemic injury and >80% reduction of blood flow. Although precursor was injected into the brain cortex after ligation of the right MCA cortex, which sustained severe ischemic damage. Stimulation of this signaling process is known to result in an increase in the cytoplasmic free calcium, which in turn may contribute to the underlying mechanism of tissue injury.

References


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