Correlation Between Myeloperoxidase-Quantified Neutrophil Accumulation and Ischemic Brain Injury in the Rat

Effects of Neutrophil Depletion

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Background and Purpose Neutrophils have been implicated in the pathogenesis of ischemia-reperfusion injury. The aim of the present study was to evaluate the correlation between neutrophil infiltration into ischemic tissues and brain injury after transient focal ischemia.

Methods We evaluated the effects of depletion of circulating neutrophils by administration of an antineutrophil monoclonal antibody (RP3) on brain edema formation, infarct size, and neutrophil infiltration (myeloperoxidase [MPO]-quantified) in rats with 1 hour of middle cerebral artery (MCA) occlusion.

Results In the cerebral cortex perfused by the anterior cerebral artery (ACA area), there was a significant increase in MPO activity only 24 hours (P<.05) after reperfusion. In the cerebral cortex perfused by the middle cerebral artery (MCA area) and caudate putamen, MPO activity was significantly increased at 12 (MCA area, P<.01; caudate putamen, P<.05), 24 (MCA area, P<.05; caudate putamen, P<.01), and 72 hours (MCA area, P<.01; caudate putamen, P<.05) and returned to near-normal level by 168 hours after reperfusion. Brain MPO activity after transient MCA occlusion correlated well with the appearance of neutrophils. Depletion of neutrophils by RP3 treatment completely inhibited the increase in MPO activity in the ischemic brain after 24 hours of reperfusion. In addition, treatment with RP3 significantly attenuated the posts ischemic increase in brain water content at 24 hours after reperfusion. RP3 also significantly reduced the size of infarct area.

Conclusions These results indicate that the increase in brain MPO activity after transient focal ischemia virtually reflects the neutrophil infiltration and that neutrophil infiltration into the ischemic brain is implicated in posts ischemic brain injury. (Stroke. 1994;25:1469-1475.)

Key Words cerebral ischemia • neutrophils • neuronal damage • rats

Recent evidence strongly suggests that neutrophils play an important role in the development of ischemia-reperfusion injury by releasing various chemical mediators such as proteases, active oxygen species, and lipid-derived mediators in various tissues. Depletion of circulating neutrophils and inhibition of neutrophil adhesive interactions can ameliorate myocardial injury after ischemia-reperfusion. In the central nervous system, neutrophil accumulation has been observed during ischemia and after reperfusion. Postischemic brain injury is attenuated by rendering the animals neutropenic with antineutrophil serum. It has also been reported that administration of monoclonal antibodies against cell adhesion molecules, such as CD11/CD18 located on neutrophils and intercellular adhesion molecule-1 (ICAM-1) located on endothelial cells, attenuates ischemic brain injury.

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Clinically and experimentally, ischemic injury accompanies acute inflammatory responses, which are characterized by neutrophil infiltration and edema formation. The degrees of neutrophil infiltration into inflammatory tissues have been estimated by histopathologic observations, counting the number of neutrophils after tissue digestion, or by radioactive means. Recently, Barone et al reported that neutrophils infiltrate into the infarcted cortex after middle cerebral artery (MCA) occlusion (MCAO), and they quantified the activity of myeloperoxidase (MPO), an enzyme localized in the azurophilic granules of these cells. However, few studies have evaluated the time course of MPO activity in ischemic brain or analyzed the correlation between MPO activity and brain injury after cerebral ischemia. Thus, the aims of the present study were to evaluate the time course of posts ischemic MPO activity and infiltration of neutrophils and to analyze the correlation between MPO activity and brain injury after transient MCAO.

We show here that the temporal profile of MPO activity correlated well with that of neutrophil infiltration and development of brain edema formation after transient focal ischemia, and that depletion of circulating neutrophils by treatment with antineutrophil monoclonal antibody significantly reduced not only the in-
creased in MPO activity in ischemic brain areas but also postischemic brain injury.

Materials and Methods

Transient focal brain ischemia in the area perfused by MCA was induced as reported previously.21-23 Briefly, adult male Wistar rats (270 to 320 g), specific-pathogen-free, were anesthetized with a gas mixture of 70% N2O, 30% O2, and 2% halothane. After median incision of the neck skin, the right external carotid artery (ECA) was carefully dissected. An 18 mm-long 4-0 nylon thread (Nitcho Kogyo Co, Ltd), precoated with silicon (Xantpren, Bayer Dental) mixed with a hardener (Optosil Activator, Bayer Dental) to increase the thickness of the distal half, was inserted from the lumen of the ECA to that of the right internal carotid artery (ICA) to occlude the origin of the right MCA. Body temperature was maintained at 37°C through the caudate putamen of the rat for tissue sampling in the cerebral cortex perfused by the anterior cerebral artery (ACA area); B and E, frontoparietal cortex, somatosensory area, supplied by middle cerebral artery (MCA area); C and F, caudate putamen. The shaded area represents typical ischemic areas.

FIG 1. Diagram of anatomic regions in a coronal section through the caudate putamen of the rat for tissue sampling in the measurement of brain water content and myeloperoxidase activity. A and D, Frontoparietal cortex, motor area, supplied by anterior cerebral artery (ACA area); B and E, frontoparietal cortex, somatosensory area, supplied by middle cerebral artery (MCA area); C and F, caudate putamen. The shaded area represents typical ischemic areas.

The method used to quantify MPO activity from rat brain samples was similar to that recently described by Barone et al24,25 but with minor modifications. The modified MPO assay for brain tissue was conducted as follows. Brain samples were thawed on ice, and wet weight in grams was measured. Each sample was homogenized (1:20, wt/vol) in 5 mmol/L potassium phosphate buffer (pH 6.0, 4°C) using an Ultra-Turrax (Janke & Kunkel Gmb & Co) for three on/off cycles at 5-second intervals and centrifuged at 2000 g (30 minutes, 4°C). The supernatant was discarded, and the pellet was washed again as described above. After decanting the supernatant, the pellet was extracted by suspension in 0.5% hexadeuteriobromophenolmammonium bromide (HTAB; Sigma Chemical Co) in 50 mmol/L potassium phosphate buffer (pH 6.0, 25°C) for approximately 2 minutes at an original tissue wet weight-to-volume ratio of 1:10. The samples were immediately frozen on dry ice. Three freeze/thaw cycles were then performed with sonications (10 seconds, 25°C; Powersonic Model 50, Powersonic Inc) between cycles. After the last sonication, the samples were incubated at 4°C for 20 minutes and centrifuged at 12 500g (15 minutes, 4°C). MPO activity in the supernatant was assayed as described previously by Bradley et al.26 Briefly, 0.1 mL of supernatant was mixed with 2.9 mL of 50 mmol/L potassium phosphate buffer, pH 6.0, containing 0.167 mg/mL o-dianisidine dihydrochloride (Sigma Chemical Co) and 0.0005% hydrogen peroxide (Wako Pure Chemical Co). The change in absorbance at 460 nm was measured with a spectrophotometer (UV-2200, Shimadzu Inc). One unit of MPO activity is defined as that which degrades 1 μmol of peroxide per minute at 25°C.

Brain water content and MPO activity were also measured in complete sham-operated rats.

For histopathology, rats were killed after 6 hours, 12 hours, 24 hours, 3 days, and 7 days of reperfusion after 1 hour of transient MCA occlusion. Brains were perfused transcardially with 100 mL of physiological saline (25°C at a pressure of 100 mm Hg), fixed in formalin, and prepared for embedding in paraffin by routine histological procedures. After embedding, 5-μm coronal sections were cut on a microtome and stained with hematoxylin and eosin. The profile and degree of neutrophil infiltration into the brain were determined and photographed under light microscopy.

The generation of anti-rat neutrophil monoclonal antibody, RP3, was described elsewhere.27 The ascitic fluid containing a high titer of RP3 was used as an antibody source in the experiment. RP3 selectively reacts and depletes rat neutrophils but not other nucleated blood cells on in vivo administration.28-30 Depletion of neutrophils with RP3 inhibited the late phase response of inflammatory edema in rats.31

To assess the influence of depletion of circulating neutrophils on neutrophil infiltration, brain edema formation, and size of infarct area after ischemia, rats were given 3 mL of the ascitic fluid containing RP3 or vehicle (saline, 3 mL per animal) intraperitoneally, 24 hours before and immediately after ischemic insult. The dose of RP3 chosen in this study is a sufficient dose to induce almost complete depletion of neutrophils from 6 hours to over 24 hours after administration and does not affect other blood cells.13,24-25 Brain water content and MPO activity after 24 hours of reperfusion were measured as described above. To measure the size of infarct area, rats were perfused with physiological saline containing 0.2% heparin after 24 hours of reperfusion. Brains were removed, cut into 1-mm coronal sections, and immersed in 2% triphenyltetrazolium chloride (TTC) solution at 37°C for 30 minutes. The colorless areas with TTC staining, which reflect mitochondrial damage, were quantified as infarct areas by an image analyzer system (Kontron M14, Zeiss). The number of neutrophils was counted before RP3 treatment, 24 hours after the first RP3 treatment (immediately before ischemic insult), and 24 hours after the second RP3 treatment (immediately before death). Rats treated with phosphate-buffered saline served as vehicle controls.
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FIG 2. Photomicrographs of cerebral cortex stained with hematoxylin and eosin after 12 (A), 72 (B), and 168 hours (C) of reperfusion following 1 hour of MCA occlusion. A, Neutrophils (small arrowheads) infiltration into the ischemic cortex was observed after 12 hours of reperfusion. B, Neutrophils (small arrowheads) and monocytes (large arrowheads) were observed in the infarcted region after 72 hours of reperfusion. C, After 168 hours of reperfusion, neutrophils disappeared, and a large amount of monocytes (large arrowheads) filled the infarcted region.

Each experiment group consisted of more than 5 animals. All values are presented as mean±SEM. For statistical analysis, Dunnett’s multiple range test and unpaired t test were used.

Results

Sections stained with hematoxylin and eosin under high magnification revealed the infiltration of neutrophils in the ischemic hemisphere after reperfusion, as shown in Fig 2. Few neutrophils were observed in the ischemic hemisphere after 6 hours of reperfusion after 1 hour of MCA occlusion. However, after 12 and 24 hours of reperfusion, a large number of neutrophils were observed in the ischemic hemisphere (Fig 2A). After 72 hours of reperfusion, many neutrophils and monocytes were observed in the infarcted region (Fig 2B). After 168 hours of reperfusion, neutrophils disappeared, and a large number of monocytes were still noticeable in the

infarcted region (Fig 2C). Fig 3 illustrates the time course of neutrophil counts in the ischemic hemisphere (per 5-μm section). We could not detect neutrophils in both ischemic and contralateral hemispheres until 6 hours after reperfusion following 1 hour of MCA occlusion. The number of neutrophils in the ischemic hemisphere began to increase from 12 hours (36.2±6.3), reached a peak between 24 hours (60.2±5.8) and 72 hours (68.6±4.7), and almost-infiltrated neutrophils disappeared 168 hours after reperfusion following 1 hour of MCA occlusion.

Fig 4 illustrates the time course of changes in brain MPO activity after reperfusion following 1 hour of MCA occlusion. In the ACA area, there was a significant increase in MPO activity only at 24 hours (0.068±0.013 U/g wet tissue, P<.05) after reperfusion following 1 hour of MCA occlusion (Fig 4A). In the MCA area, MPO activity began to rise from 12 hours (0.040±0.018 U/g wet tissue, P<.01), reached a peak at 24 hours (0.270±0.074 U/g wet tissue, P<.01), and returned to near-normal level (sham-operated, 0.021±0.003 U/g wet tissue) by 168 hours. In the caudate putamen, MPO activity began to rise from 12 hours (0.075±0.029 U/g wet tissue, P<.05), reached a peak at 72 hours (0.080±0.029 U/g wet tissue, P<.01), and returned to normal level (sham-operated, 0.010±0.002 U/g wet tissue) by 168 hours. In the caudate putamen, we observed a significant increase in MPO activity even at 168 hours (0.027±0.002 U/g wet tissue, P<.01) after reperfusion (Fig 4C). Brain MPO activity after transient MCA occlusion correlated well with the appearance of neutrophils.

Fig 5 illustrates the time course of the changes in brain water content after reperfusion following 1 hour of MCA occlusion. Brain water content continued to rise significantly until 72 hours after reperfusion, and then returned to near-normal level (sham-operated) by 168 hours after reperfusion (Fig 5A, 5B, and 5C). Brain edema formation correlated well with MPO activity, reflecting neutrophil infiltration.
Fig 4. Graphs showing myeloperoxidase (MPO) activity of anterior cerebral artery areas (A), middle cerebral artery (MCA) areas (B), and caudate putamen (C) after 6, 12, 24, 72, and 168 hours of reperfusion following 1 hour of MCA occlusion. Each point is mean±SEM of 5 to 12 animals. *P<.05, **P<.01 vs sham-operated rats (unpaired t test).

Fig 5. Graphs showing water content of anterior cerebral artery areas (A), middle cerebral artery areas (B), and caudate putamen (C) after 6, 12, 24, 72, and 168 hours of reperfusion following 1 hour of MCA occlusion. Each point is mean±SEM of 5 to 11 animals. ○ indicates occluded side; ●, nonoccluded side. *P<.05, **P<.01 vs sham-operated rats (unpaired t test).

Discussion

The present study indicates that (1) the MPO activity after transient focal ischemia correlated well with the
The invasion of neutrophils into the ischemic areas is implicated in the development of postischemic brain injury. As to the mechanisms of depletion of in vivo neutrophils by RP3, there is little information. Sekiya et al reported that peritoneal macrophages that showed phagocytosis of neutrophils were observed after administration of RP3. Phagocytosis of antibody-coated neutrophils by macrophages may be a candidate in the mechanism of neutrophil depletion. In our present study, we administered saline as the vehicle. We should have used nonimmune serum as control to remove the possibility that neutrophil depletion. In our present study, we administered saline as the vehicle. We should have used nonimmune serum as control to remove the possibility that neutrophil depletion by antibody is not the cause of the neuroprotective effect of the antibody. Nevertheless, these results are strongly indicative of a contribution of neutrophils to brain injury under ischemia/reperfusion conditions. However, there is a possibility that depletion of neutrophils alters hemorheological factors that may contribute to the mechanism of protection. Further work is required to clarify the effect of neutrophil depletion on blood viscosity.

**Fig 7.** Graphs showing the effects of depletion of circulating neutrophils with RP3 on increased brain myeloperoxidase (MPO) activity (A), increased brain water content (B), and infarct size (C) after 24 hours of reperfusion following 1 hour of middle cerebral artery (MCA) occlusion. ACA indicates frontoparietal cortex supplied by anterior cerebral artery (ACA area); MCA, frontoparietal cortex supplied by middle cerebral artery (MCA area). *P<.05, **P<.01 vs vehicle (Dunnett’s multiple range test).
The roles of neutrophils in the pathogenesis of cerebral ischemia and stroke include the no-reflow phenomenon caused by vessel plugging and release of vasoconstrictive mediators, enhancement of vascular permeability, parenchymal injury by hydrolytic enzyme release, lipid mediator synthesis, and active oxygen species production. We assume that the depletion of neutrophils results in the reduced release of chemical mediators in the ischemic area and lightens the no-reflow phenomenon, thus suppressing postischemic brain injury.

Increased adherence of circulating neutrophils to the vascular endothelium is an essential early event in the initiation of an inflammatory response after a period of ischemia-reperfusion. However, the precise mechanism of the neutrophil adherence under the ischemia-reperfusion condition is still poorly understood. Recent reports have indicated that ischemia-reperfusion treatment increases CD11/CD18-ICAM-1-dependent neutrophil adherence to endothelial cells and that platelet-activating factor (PAF) and PAF-induced active oxygen species (H2O2 and superoxide anion) are responsible for this hyperadhesiveness. PAF is synthesized in the brain during ischemia or convulsions, and beneficial effects of PAF receptor antagonists have been demonstrated in various models of cerebral ischemia.

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Acknowledgments

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The article by Matsuo et al is an important milestone in the elucidation of the role of inflammation in ischemic brain injury. Although the accumulation of leukocytes into ischemic (permanent or transient) brain has been repeatedly demonstrated, the significance of leukocyte accumulation on the final outcome of stroke (histologically and functionally) is far from being settled. The generation of the RP3 anti-rat neutrophil antibody has been extremely valuable in studying this issue because it provides a specific means to selectively deplete neutrophils with a monoclonal antibody. J Leukoc Biol. 1990;48:259-265.

The significance of leukocyte adherence to the brain capillaries results in microhemodynamic alterations, many critical issues remain unanswered. First, it is not known whether “distancing” leukocytes from the brain capillaries results in microhemodynamic alterations that enable critical flow patterns, especially in the penumbra region; it would be of great interest to explore rheological factors at the capillary level in neutrophil-depleted rats. Second, it would be of critical importance to evaluate the therapeutic opportunities available in a clinical time frame (ie, less than 2 hours after ischemia), because depletion of leukocytes in a preventive mode (either by an RP3-like strategy or an antibody-mediated approach) carries liabilities such as host-defense impairment and is therefore unrealistic for clinical use. Third, the mediators generated by the leukocytes that contribute to brain injury should be better understood. In this respect, one must keep in mind that reciprocal interactions between the leukocytes and brain cells (glia and neurons) may result in modified neurochemical profiles of either or both cells such that factors released from leukocytes may stimulate glia or neurons to release toxic factors (eg, cytokines) that then amplify the inflammatory process and aggravate the distended environment.

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