Modulation of Basal and Postischemic Leukocyte-Endothelial Adherence by Nitric Oxide

Jeffrey M. Gidday, PhD; T.S. Park, MD; Aarti R. Shah, MS; Ernesto R. Gonzales, BSN

Background and Purpose—Recent studies indicate that leukocytes are important contributors to secondary vascular and parenchymal injury after cerebral ischemia. The present study was undertaken to define nitric oxide (NO)—based mechanisms that regulate leukocyte-endothelial interactions in the cerebral vasculature, how these mechanisms are affected by cerebral ischemia, and whether NO-based therapies can affect postischemic leukocyte dynamics.

Methods—Leukocyte adherence to pial venules of anesthetized newborn piglets was quantified by in situ fluorescence videomicroscopy through closed cranial windows during basal conditions and during reperfusion after 9 minutes of asphyxia. Nitric oxide synthase (NOS) was inhibited by local window superfusion of L-nitroarginine; superfusion of sodium nitroprusside was used to donate NO.

Results—Local inhibition of NOS under resting conditions increased leukocyte-endothelial adherence 2.2-fold and 3.9-fold over baseline values after 1 hour and 2 hours, respectively; this response was completely blocked by cosuperfusion with L-arginine. Cosuperfusion of superoxide dismutase reversed L-nitroarginine–induced leukocyte adherence by 89% and 63% at these respective time points. The extent of acute leukocyte adherence elicited by NOS inhibition was similar in magnitude to that observed during the initial 2 hours of reperfusion after asphyxia. Leukocyte adherence was not additionally increased in asphyxic animals treated with L-nitroarginine. Sodium nitroprusside robustly inhibited asphyxia-induced leukocyte adherence back to control levels.

Conclusions—NO exerts a tonic antiadherent effect in the cerebral microcirculation by inactivation of adherence-promoting superoxide radical formation. Cerebral ischemia is associated with an inhibition of NOS or lower levels of NO, which results in leukocyte-endothelial adherence that can be prevented by NO donors. The latter may be useful therapeutically to prevent the purported vascular and parenchymal dysfunction and injury caused by activated leukocytes in ischemic brain. (Stroke. 1998;29:1423-1430.)

Key Words: cerebral ischemia, global ■ leukocytes ■ nitric oxide ■ reperfusion injury ■ pigs

It is now recognized that the abnormal behavior of leukocytes accompanies many disease states. Accumulating evidence indicates that both focal and global cerebral ischemia can elicit an acute inflammatory response characterized, in part, by leukocytes adhering to microvessel endothelial cells, plugging capillaries, and extravasating into brain parenchyma. On repuffusion, this multistep response progresses from an early stage of coactivation of circulating leukocytes and cerebral endothelial cells, to expression of their respective adhesion molecules, to rolling and sticking of leukocytes to endothelial cells, and, within hours to days after the initial insult, to diapedesis of leukocytes from the intravascular to the extravascular space. Although not all studies support an injurious role for leukocytes in ischemic brain injury, the documented rheological and hemodynamic effects of adherent leukocytes in cerebral vessels and the potent destructive capability of the free radicals and proteases these cells contain strongly suggest that the inadvertent activation of this inflammatory cascade could be an important contrib-
paratively little is known about the control of leukocyte adherence by NO in the cerebral circulation, and results of the sole report to date can counter to the prevailing consensus about NO-based regulation of leukocyte-endothelial interactions in noncerebral tissues. Thus, we undertook the present study to begin to elucidate mechanisms controlling NO-regulated leukocyte-endothelial interactions in the cerebral circulation.

## Materials and Methods

### Animal Preparation and Drug Superfusion

Fifty-six newborn piglets (age, 2 to 5 days) weighing 1.5 to 3.5 kg were used in experimental protocols that were consistent with Public Health Service guidelines and were approved by our institutional animal studies committee. The preparation for in vivo monitoring of leukocyte dynamics has been detailed previously. In brief, after a tracheostomy under ketamine hydrochloride anesthesia (20 mg/kg IM), animals were ventilated with a mix of room air and oxygen, and anesthesia was maintained for the remainder of the experiment with isoflurane (1.0% to 1.5%). End-tidal CO$_2$ and transcutaneous O$_2$ measurements of gas tensions, glucose concentration, pH, and hematocrit. A thermoregulated heating pad and overhead heating lamp were used to maintain core body temperature at 38°C to 39°C, except during the last few minutes of the 9-minute asphyxic period (when cortical temperature could drop 1°C to 2°C, particularly if the animals arrested for more than 1 minute) and during the initial 15 minutes of reperfusion (at which time cortical temperature had returned to baseline after falling no more than 2°C).

After an 18-mm craniotomy and removal of the dura, a closed cranial window made of Plexiglas was placed over the right parietal cortex. Through ports at the edge of the window, intracranial pressure was continuously monitored; juxtaposed ports were used to superfuse drug solutions made up in artificial CSF, as described previously. Buffer or drug solutions were introduced into the window space by superfusion at 1 mL/min for 1 minute, followed by a continuous superfusion rate of 50 μL/min for 2.0 or 2.5 hours, with the use of an automated syringe pump.

### Leukocyte Quantification by In Situ Fluorescence Videomicroscopy

Leukocytes were fluorescently labeled in situ with rhodamine 6G, which stains 100% of circulating leukocytes as assayed by flow cytometry. The loading dose consisted of 2 mL/kg of a filtered 0.006-mg/mL solution administered intravenously over 5 minutes, 30 minutes before the first baseline imaging period commenced. One to 2 minutes before each 60-second imaging period, rhodamine 6G was infused at 800 μL/min to enhance labeling. Leukocyte dynamics in pial venules were recorded to videotape in real time with the use of a Newvicon tube camera mounted on an epifluorescence microscope.
Effects of local NOS inhibition with L-NA and asphyxia/reperfusion on leukocyte adherence in piglet pial venules, as measured by in situ fluorescence videomicroscopy. The increases in the actual number of adherent leukocytes per square millimeter of pial venular area are shown, relative to the number of leukocytes adherent to the venular area before drug administration or asphyxia. Superfusion of L-NA (100 μmol/L; △; n=7) through the cranial window for 2 hours resulted in leukocyte adherence similar to that observed during the initial 2 hours of reperfusion after asphyxia (●; n=9); both conditions resulted in leukocyte adherence significantly greater than that measured in untreated controls (◆; n=13). The adherence-promoting effect of L-NA was reversed by cosuperfusion with a 100-fold molar excess of l-arginine (Δ; n=6). *P<0.05 vs control group at the same time point.

During baseline conditions and during 2 hours of reperfusion after asphyxia. Similarly, no significant differences in maximum or minimum venular diameters, the areas of venular network measured, or baseline arteriolar diameters (32 to 43 μm) were noted between groups. No changes in systemic physiological variables occurred in response to local superfusion of the drugs indicated. Finally, there were no significant differences between groups with respect to the mean number of leukocytes adherent to cerebral venules under baseline conditions (39 to 87/mm²).

In nonischemic control animals (group 1), a slight increase in leukocyte adherence occurred over the 2-hour observation period relative to that measured during baseline conditions (Figure 1). At 1 hour and 2 hours of observation, adherence increased 21% ± 7% and 39% ± 11% above baseline, respectively; only the latter change was significantly greater than baseline values. Leukocyte adherence was significantly increased in animals in which cranial windows were continuously superfused with the NOS inhibitor L-NA (group 2); the increase in adherence was progressive over time, such that adherence at 2 hours of superfusion (163% ± 65% above baseline) was significantly greater than that measured at 1 hour of reperfusion (49% ± 29% above baseline). This L-NA–induced adherence was almost totally reversed by cosuperfusion of a 100-fold molar excess of l-arginine (group 3; Figure 1). Figure 1 also shows that the magnitude and temporal pattern of leukocyte adherence that was elicited by local NOS inhibition was nearly identical to that observed in animals subjected to asphyxia and 2 hours of reperfusion (group 5). When cortical NOS was inhibited in animals rendered asphyxic (group 6), no further increase in leukocyte adherence was observed at any time point relative to animals subjected to asphyxia alone (data not shown).

To begin to address the mechanism of leukocyte adherence after NOS inhibition, we tested the hypothesis that increases in superoxide free radical levels resulting from a loss of NO promoted such adherence. We reasoned that concomitant superfusion of SOD with L-NA would eliminate the increase in leukocyte adherence we witnessed with L-NA alone. Indeed, in these animals (group 4), leukocyte adherence was dramatically attenuated relative to group 2 L-NA–treated animals without SOD (Figure 2). In particular, leukocyte adherence after 1 hour of drug exposure was reduced to levels equivalent to those in untreated controls; by 2 hours of drug exposure, adherence was still significantly reduced relative to animals receiving L-NA alone but also became significantly greater than adherence levels observed in untreated controls at the same time point.

To examine the corollary hypothesis that exogenous NO could attenuate ischemia-induced increases in leukocyte adherence, the NO donor SNP was superfused through the cranial window of asphyxically injured animals at the initiation of reperfusion (group 7). This treatment resulted in a robust and significant reduction in asphyxia-induced leukocyte adherence to levels not significantly different from those in nonasphyxic, untreated controls (Figure 3).

Changes in pial arteriolar diameter in the control group and in animals superfused with either L-NA or SNP are shown in the Table. No significant change in arteriolar diameter was measured in control animals (group 1) over time. Superfusion of L-NA in group 2 animals did not significantly affect pial arteriolar diameter at any time point. Conversely, superfusion...
Nitric Oxide Inhibits Leukocyte Adherence

The present study provides the first evidence in the cerebral circulation that (1) inhibition of NOS during nonischemic resting conditions results in an acute, arginine-reversible adherence of leukocytes to cerebral venules; (2) superoxide radicals are involved in mediating leukocyte adherence after NOS inhibition; (3) inhibition of NOS during ischemia and reperfusion does not exacerbate the extent of leukocyte adherence induced by ischemia/reperfusion alone; and (4) local administration of an NO donor can dramatically attenuate leukocyte adherence after cerebral ischemia. Thus, two lines of complementary evidence gathered herein support an important role for NO in modulating leukocyte-endothelial interactions in the cerebral circulation under both physiological and pathophysiological conditions. Our findings suggest that a balance between the antiadherent effects of NO and the proadherent effects of superoxide radical underlies the changes in leukocyte dynamics we observed with both NOS inhibition and ischemia/reperfusion. These findings are likely to have important implications for anti-inflammatory stroke therapy during the initial hours of reperfusion.

Endothelial cells, leukocytes, and platelets contain both constitutive and inducible NOS isoforms. In addition to influencing tissue perfusion, accumulating evidence gathered from studies of noncerebral vascular beds indicates that NO produced at the blood-endothelial interface also modulates leukocyte adherence, platelet aggregation, and endothelial permeability in a tonic fashion. In resting peripheral microcirculatory beds, for example, leukocytes adhere to venular endothelium after administration of NOS inhibitors. Under similar nonischemic conditions, we confirmed that local inhibition of NOS with L-NA elicited a progressive increase in leukocyte adherence to the pial venular microcirculation over 2 hours of continuous drug presentation and observation. As expected, this effect was arginine reversible. Given the well-established concept that, in most species, NO contributes a tonic dilator effect in the cerebral circulation secondary to its ongoing production by endothelial NOS, our findings indicate that tonically released NO also acts to inhibit the adherence of circulating leukocytes to cerebrovascular endothelium. Our results differ importantly from those in a recent cranial window study in rats, wherein topical L-NA administration (1 mmol/L) did not significantly elevate leukocyte adhesion. However, in the latter study, when the basal level of activation of circulating leukocytes and/or cerebrovascular endothelium was increased mildly with leukotriene B4 superfusion, subsequent L-NA administration promoted significant leukocyte adherence. It is difficult to identify underlying reasons for these discordant observations, given that the magnitude of trauma-induced histamine release, cytokine release, or mast cell degranulation in response to surgical preparation of a cranial window in rats and piglets is probably similar, but species-, anesthesia-, and age-dependent differences in receptor sensitivity, in the regulation of adhesion molecule expression, or in other unidentified parameters could be important. Clearly more studies of the control of leukocyte-endothelial adherence by NO in the cerebral circulation are warranted in other stroke models to resolve these important issues.

The mechanisms whereby tonically produced NO serves to inhibit leukocyte-endothelial interactions are likely to be multifactorial. Changes in vessel shear rate, interactions with superoxide radical, and alterations in adhesion molecule expression are likely candidates. For example, the vessel shear rate resulting from the tonic vasodilative effect of NO would be reduced after NOS inhibition, and leukocyte adherence might then be promoted secondary to a decrease in blood flow. However, the lack of change in pial arteriolar

### Changes in Pial Arteriolar Diameter in Response to Continuous L-NA or SNP Superfusion

<table>
<thead>
<tr>
<th>Experimental Group</th>
<th>Pial Arteriolar Diameter, μm</th>
<th>Percent Change From Baseline</th>
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<tbody>
<tr>
<td>Controls (group 1; n=13)</td>
<td></td>
<td></td>
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<tr>
<td>Baseline</td>
<td>38±4</td>
<td>...</td>
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<tr>
<td>1 h</td>
<td>39±6</td>
<td>1±6</td>
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<tr>
<td>2 h</td>
<td>38±7</td>
<td>2±8</td>
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<tr>
<td>L-NA (100 μmol/L; group 2; n=7)</td>
<td></td>
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<tr>
<td>Baseline</td>
<td>37±3</td>
<td>...</td>
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<tr>
<td>1 h superfusion</td>
<td>36±3</td>
<td>−1±7</td>
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<tr>
<td>2 h superfusion</td>
<td>34±2</td>
<td>−7±5</td>
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<tr>
<td>SNP (40 μmol/L; n=4)</td>
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<tr>
<td>Baseline</td>
<td>43±9</td>
<td>...</td>
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<tr>
<td>1 h superfusion</td>
<td>68±16*</td>
<td>61±8†</td>
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<tr>
<td>2 h superfusion</td>
<td>70±18*</td>
<td>61±6†</td>
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*P<0.05 vs baseline; †P<0.05 vs control group (group 1).
diameter in response to L-NA superfusion in piglets suggests that leukocyte adherence was invoked with little change in local cortical blood flow. This finding is consistent with the predominant dependence of the newborn piglet cerebral circulation on adenosine and prostanoioids for the metabolic regulation of cerebral vascular resistance, with NO-based regulatory systems becoming operative only after maturation.23 Although vasoactive effects of this tonically produced NO are not realized (perhaps because of alterations in the sensitivity of downstream effector pathways in the vascular smooth muscle), our finding that leukocyte adherence was stimulated after NOS inhibition indicates that enough NO is produced at the blood-endothelial interface under baseline conditions in the piglet cerebral circulation to maintain an antiadhesive endothelial surface for circulating leukocytes independent of an effect on shear rate. NO exhibits a similar shear-independent effect on basal leukocyte adherence in the cat mesentery.10

A primary mechanism whereby NO is likely to negatively affect leukocyte adherence is based on the avidity of NO for interacting with superoxide free radical. Endogenous NO competes with SOD to inactivate basally produced superoxide radical24,25; thus, loss of NO after NOS inhibition could lead to increases in the levels of superoxide radical, a well-established proadherent molecule in a variety of microcirculatory beds.26,27 In fact, superfusion of NOS inhibitors increased oxidative stress–sensitive probe fluorescence in rat mesenteric venules before leukocyte adherence.20,22 indicative of increased radical production in response to loss of endogenous NO. Our finding that L-NA–induced adherence to cerebral venules is attenuated dramatically by concomitant superfusion of SOD supports the hypothesis that ongoing NO production balances ongoing oxidant formation and is consistent with similar findings resulting from coadministration of oxygen radical scavengers and NOS inhibitors in non-cerebral tissues.20,26,28

Both endothelial cells and leukocytes could serve as the cellular source of oxygen radicals in the face of declining NO levels during basal conditions. Mitochondrial-rich cerebral endothelial cells produce superoxide as a result of electron transport chain activity during aerobic metabolism, from cyclooxygenase-dependent formation of prostanoioids, and from the activity of xanthine oxidase.20 Elegant intravital microscopy studies21 have demonstrated increases in hydperoxide formation within endothelial cells in response to treatment with NOS inhibitors. In addition, considerable evidence is available documenting the ability of NOS inhibitors to increase free radical production by leukocytes.20 Conversely, NO can reduce free radical formation from activated neutrophils22,30 by inhibiting NADPH oxidase,31 the primary enzymatic source of leukocyte-derived free radicals. The prevention of L-NA–induced adherence with SOD that we and others documented20,28 suggests that superoxide radical may be the particular radical species responsible for promoting adherence, but similar antiadherent effects with catalase20 indicate that other downstream radical species (hydrogen peroxide and/or hydroxyl radical) may be involved as well.

In addition to influencing steady-state oxygen free radical levels, NO may regulate in a direct fashion the expression of endothelial and leukocyte adhesion molecules. There is evidence in the rat ilial mesenteric microcirculation32,33 that NO inhibits the endothelial, cyclic GMP–dependent expression of P-selectin, which in turn promotes rolling of leukocytes on the endothelium at sites of inflammation before their firm adherence. Such adherence is dependent on endothelial cell expression of intercellular adhesion molecule and vascular cell adhesion molecule, which also appear to be downregulated by NO.23,34 Although an NO-induced inhibition of the expression of the leukocyte CD18 adhesion molecule, the co-ligand for endothelial intercellular adhesion molecule, has been demonstrated in vitro with the use of cultured endothelium,35 similar support for this mechanism could not be demonstrated in vivo.36 Finally, NO may also tonically prevent leukocyte adherence in a more indirect way by inhibiting the production of proinflammatory chemoattractants.

There is now considerable evidence indicating that an acute inflammatory response occurs after cerebral ischemia, characterized by a progressive increase in leukocyte adherence and infiltration over the initial hours to days after the insult.1 We demonstrated previously that the severe hypoxia and hypotension accompanying 9 minutes of asphyxia in piglets elicits significant leukocyte adherence even within the initial 2 hours of reperfusion.11 The magnitude and time course of this adherence were nearly identical to those observed in the present study after L-NA superfusion. That no additional increase in leukocyte adherence occurred during the early posts ischemic reperfusion period after NOS inhibition by L-NA suggests that asphyxia–reperfusion resulted in a depletion of endogenous basal levels of NO. As found in other tissues, recent studies in brain37 document an attenuation or absence of NO-dependent vasoreactivity during the initial hours after ischemia, even though reactivity to NO donors remains intact, indicating that endothelial NOS function is impaired after ischemia and/or that NO is efficiently scavenged once it is produced. The robust increase in oxygen free radical formation occurring coincident with posts ischemic reperfusion38 is consistent with the latter possibility but does not explain how NO donors retain their vasoreactivity after ischemia. In either event, the data collectively suggest that a fall in basal NO levels after ischemia underlies, in part, the early leukocyte-endothelial adherence behavior we observed. Our studies only examined the initial 2 hours of reperfusion; thus, the time course over which a significant NO-based antiadherent effect might be reestablished, as well as the effect of large increases in NO production from inducible NOS on posts ischemic leukocyte adherence, remains undefined.

If the above hypothesis that NO exhibits multifunctional antiadherent activity is correct, then supplementing posts ischemic tissue with NO donors or NO precursors would be expected to attenuate the degree of leukocyte sticking after ischemia. Indeed, posts ischemic leukocyte adherence was dramatically reduced in our model when the organic nitrate NO donor SNP was superfused across the cortical surface at the start of reperfusion. Our findings in brain are consistent with results from intravital microscopy studies in the splanch-
nic microcirculation wherein arginine supplementation or administration of a variety of NO donors decreases ischemia-induced leukocyte adherence,32,36 and P-selectin expression13 in a similar fashion.

As alluded to above, direct quenching of superoxide radical is one mechanism whereby pharmacological augmentation of NO levels may attenuate leukocyte adherence after ischemia or after direct exposure to oxygen free radical-generating systems,26 since superoxide radicals potently stimulate adhesion molecule expression and leukocyte adherence. Indeed, superfusion of SOD attenuates ischemia-induced leukocyte adherence in our piglet model similar to SNP (J.M.G. et al, unpublished data, 1997). In addition to direct superoxide inactivation, NO may indirectly reduce superoxide formation as a result of its ability to suppress endothelial xanthine oxidase.40 This enzyme forms superoxide radical in cerebral endothelial cells when hypoxanthine and xanthine are converted to uric acid after ischemia-induced purine catabolism.29 There is also the possibility that an increase in blood flow accounts, in part, for the decrease in postischemic leukocyte adherence after administration of NO donors. With the pial arteriolar dilation elicited by superfusion of SNP, we cannot rule out the possibility that the resultant increase in shear may have contributed to the SNP-induced reduction in leukocyte adherence; parallel control studies employing other common vasodilators like adenosine or prostaecyclin are problematic given their NO dependence or their direct antiadherent effects.

The role of NO in modulating postischemic leukocyte adherence that we demonstrated herein may, in addition to its hemodynamic22 and platelet antiaggregatory14 effects, explain in part the findings that infarcts resulting from middle cerebral artery occlusion are larger in endothelial NOS knockout mice,43 that early administration of NOS inhibitors exacerbates ischemic damage,42,43 and that L-arginine supplementation and NO donors decrease brain injury and improve outcome in a variety of stroke models.12,44,45 Our previous demonstration that blood-brain barrier breakdown in asphyxiated piglets results, in part, from adherent leukocytes13 suggests that reductions in postischemic edema may also be realized with NO-based therapy secondary to reductions in postischemic microvascular permeability; the latter has been documented in the rat mesenteric microcirculation.20

In summary, we have demonstrated that NO inhibits leukocyte adherence to cerebral venules in a tonic fashion by inactivating basally produced superoxide radical, that the antiadherent effect of NO is lost during the initial hours of reperfusion after ischemia as a result of an impairment in NO and/or an increase in free radical formation, and that NO supplementation can reverse ischemia-induced leukocyte adherence. Mechanistically, these effects of NO are much more complex than our end points indicate, and future work can elucidate how they are likely to vary depending on the nature of the ischemic insult, the relative extent of NO and oxygen free radical production, the status of many coexistent homeodynamic variables, and the time at which the inflammatory response to ischemia is examined.

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In 1993, Kubes and coworkers introduced the concept that NO functions as an important regulator of leukocyte adherence, possibly via roles in modulating local superoxide or hydroxyl radical levels and/or effects on expression of P-selectin or CD18. This role for NO in the tonic regulation of leukocyte adherence in the mesenteric circulation of the rat did not seem to operate in rat cerebral circulation, as reported by Lindauer et al; thus, the antiadhesion effects of NO were thought to play a more limited role in brain versus microvascular beds outside of the central nervous system. However, Lindauer et al did report that NOS inhibitors augment leukocyte adherence in LTB4-stimulated rat pial microcirculation. In the accompanying article by Gidday et al, we are shown that NO exerts tonic basal antiadherent effects in the pial microcirculation of newborn piglets and that loss of endogenous local NO production after asphyxia may play an important role in promoting the adherence of leukocytes. Thus, loss of NO could potentially magnify secondary damage produced by the acute inflammatory response. Factors related to species (rat versus piglet versus humans), age (mature versus immature), or model (focal or global ischemia versus asphyxial cardiopulmonary arrest) could potentially limit the importance of this observation. Nevertheless, it appears that another layer of complexity has been added to the potential roles for NO in the evolution or prevention of secondary damage in the injured brain—in this case, another putative favorable effect. This work brings to light a fundamental issue that deserves additional discussion.

Numerous recent studies in models of both ischemic and traumatic brain injury are demonstrating both detrimental and beneficial effects of an incredibly complex and highly interactive local inflammatory response. Some of the NO-mediated effects in this setting, related to the inducible form of NOS (iNOS), are considered part of traditional "inflammation." However, to be more encompassing, both NO and a variety of inflammatory participants seem to share this bipolar role in the injury response in brain, and we will take the liberty of discussing them together as part of a "tissue-injury response" rather than an "inflammatory response." Both detrimental and beneficial aspects of NO and many components of this tissue-injury response have been reported. This includes a variety of participants, such as cytokines, adhesion molecules, NF-κB, and NO. Adding to the complexity, a variety of factors appear to determine whether beneficial or detrimental aspects dominate for any given component of this tissue-injury response. Included in this extensive list are factors such as the experimental model or specific clinical condition involved, the site of mediator.
production (ie, neuron, astrocyte, or microcirculation), the timing of the event in question or of the therapeutic intervention, and a variety of other contributors. Certainly for NO, both beneficial and detrimental effects seem to be operating in the injured brain. NO derived from neuronal NOS during the excitotoxic response to ischemia/reperfusion may lead to peroxynitrite formation, PARS activation, and neuronal death.

Under different redox conditions, however, NO-mediated nitrosylation of the NMDA receptor or caspases may attenuate neuronal death. Similarly, microcirculatory effects of NO may help provide flow in the ischemic penumbra and, as we are shown here by Gidday et al, attenuate local inflammation in injured brain.

Despite a number of studies targeting selected aspects of the tissue-injury response after ischemic or traumatic brain injury, the only approaches to date that have paid dividends in the clinic (although still somewhat controversial) are augmentation of reperfusion with thrombolytics in stroke6 (ie, good, old-fashioned plumbing) or the application of a broad-spectrum therapy such as hypothermia in traumatic brain injury.7 Is the inflammatory (or tissue-injury) response both too cybernetic and too bipolar to yield a therapeutic approach that will translate into a clinical breakthrough? Are there purely deleterious aspects of this response to target with inhibitors? Are there components that should be targeted and enhanced because they are exclusively and/or powerfully beneficial? Despite what appears to many to be an “old story,” our knowledge of how NO and many other inflammation-related aspects of the tissue-injury response, we have yet to begin to answer the grade school questions of “who, what, when, where, why, and how.”

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