Fluoxetine Dilates Isolated Small Cerebral Arteries of Rats and Attenuates Constrictions to Serotonin, Norepinephrine, and a Voltage-Dependent Ca$^{2+}$ Channel Opener

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**Background and Purpose**—Recent clinical observations question that the antidepressant effect of fluoxetine (Prozac) can be explained solely with serotonin reuptake inhibition in the central nervous system. We hypothesized that fluoxetine affects the tone of vessels and thereby modulates cerebral blood flow.

**Methods**—A small branch of rat anterior cerebral artery (195±15 μm in diameter at 80 mm Hg perfusion pressure) was isolated, cannulated, and pressurized (at 80 mm Hg), and changes in diameter were measured by videomicroscopy.

**Results**—Fluoxetine dilated small cerebral arteries with an EC$_{50}$ of 7.7±1.0×10$^{-6}$ mol/L, a response that was not affected by removal of the endothelium or application of 4-aminopyridine (an inhibitor of aminopyidine-sensitive K$^+$ channels), glibenclamide (an inhibitor of ATP-sensitive K$^+$ channels), or tetraethylammonium (a nonspecific inhibitor of K$^+$ channels). The presence of fluoxetine (10$^{-6}$ to 3×10$^{-5}$ mol/L) significantly attenuated constrictions to serotonin (10$^{-9}$ to 10$^{-5}$ mol/L) and norepinephrine (10$^{-9}$ to 10$^{-7}$ mol/L). Increasing concentrations of Bay K 8644 (a voltage-dependent Ca$^{2+}$ channel opener, 10$^{-10}$ to 10$^{-6}$ mol/L) elicited constrictions, which were markedly reduced by 2×10$^{-6}$ and 10$^{-5}$ mol/L fluoxetine, whereas 3×10$^{-5}$ mol/L fluoxetine practically abolished the responses.

**Conclusions**—Fluoxetine elicits substantial dilation of isolated small cerebral arteries, a response that is not mediated by endothelium-derived dilator factors or activation of K$^+$ channels. The finding that fluoxetine inhibits constrictor responses to Ca$^{2+}$ channel opener, as well as serotonin and norepinephrine, suggests that fluoxetine interferes with the Ca$^{2+}$ signaling mechanisms in the vascular smooth muscle. We speculate that fluoxetine increases cerebral blood flow in vivo, which contributes to its previously described beneficial actions in the treatment of mental disorders. (*Stroke. 1999;30:1949-1954.*)

**Key Words:** calcium channels • cerebral arteries • dilation • fluoxetine • smooth muscle

Previous studies suggest that 5-hydroxytryptaminergic (serotonergic) pathways in the central nervous system are involved in the pathogenesis of human depression.$^1$ It is thought that by blocking 5-hydroxytryptamine (5-HT) reuptake from nerve terminals, newer antidepressant drugs, such as fluoxetine (Prozac), enhance 5-serotonergic transmission, which is regarded as the primary therapeutic action of these compounds.

Several recent studies suggested that fluoxetine has additional effects that are apparently not related to the inhibition of the neuronal 5-HT reuptake. For example, in neuronal tissue, fluoxetine has been reported to antagonize nicotinic acetylcholine$^2$ and 5-HT$^{3,4}$ receptors and to block sodium$^3$ and voltage-dependent potassium channels.$^5$ Moreover, fluoxetine was reported to inhibit both voltage-dependent and 5-HT–induced increases in [Ca$^{2+}$]i in rat and human cortical neurons.$^7$–$^9$ Fluoxetine has also been shown to affect the function of smooth muscle. It antagonized 5-HT–induced contraction of isolated rat intestinal preparation,$^{10}$ affected potassium channels in isolated intestinal smooth muscle cells,$^{11}$ and inhibited the high potassium–induced contraction of uterine preparations.$^{12}$ In isolated pulmonary arterial smooth muscle cells, fluoxetine inhibited the 5-HT–induced [Ca$^{2+}$]i increase,$^{13}$ suggesting that fluoxetine interferes with the generation of vascular tone. However, the possible effects of fluoxetine on the function of cerebral arteries is not known, and their possible contribution to the therapeutic action of fluoxetine has not been considered.

Thus, in the present study we aimed to characterize the effects of fluoxetine on the myogenic tone of small cerebral arteries of rats. To avoid the masking effect of neural and hormonal regulation of the vascular tone, experiments were conducted on isolated rat cerebral arteries. To elucidate the underlying mechanisms responsible for the vascular action of fluoxetine, the possible roles of endothelium, opening of K$^+$...
channels, inhibition of voltage-dependent Ca\textsuperscript{2+} channels, and the responses to 5-HT and norepinephrine were assessed.

**Materials and Methods**

**Isolation of Arterioles**

Experiments were conducted on isolated rat small cerebral arteries (~195 \, \mu m in diameter at 80 mm Hg). The methods for isolation of vessels have been described in detail previously.\textsuperscript{14} The procedures followed were in accord with institutional guidelines. Male Wistar rats (weighing 350±30 g) were anesthetized with pentobarbitone (75 mg/kg IP). The brain was removed immediately after decapitation and placed in a silicone-lined Petri dish containing cold (0°C to 4°C) physiological salt (PS) solution composed of (in mmol/L) NaCl 110, KCl 5.0, CaCl\textsubscript{2} 2.5, MgSO\textsubscript{4} 1.0, KH\textsubscript{2}PO\textsubscript{4} 1.0, dextrose 10.0, and NaHCO\textsubscript{3} 24.0 and was equilibrated with a gas mixture of 10% O\textsubscript{2} and 5% CO\textsubscript{2}, balanced with nitrogen, at pH 7.4. With microsurgical instruments and an operating microscope, a 1.5-mm segment of a first daughter branch of the anterior cerebral artery running superficially was isolated and transferred into an organ chamber containing 2 glass micropipettes filled with PS solution. One artery was used from each rat. The vessel chamber (15 mL) was continuously supplied with PS solution from a reservoir at a rate of 40 mL/min. After the vessel had been mounted on the proximal micropipette and secured with a Teflon clip, the perfusion pressure was raised to 20 mm Hg to clear the debris from the lumen. Then the other end of the vessel was mounted on the distal pipette. Both micropipettes were connected with silicone tubing to an adjustable PS solution reservoir.

**Experimental Protocols**

Only those vessels that developed spontaneous tone in response to perfusion pressure were used, and thus, no vasoactive agent was added to the PS solution to establish arterial tone. After the equilibration period, the diameter of vessels was measured at 80 mm Hg perfusion pressure under zero-flow conditions. At the conclusion of each experiment, the perfusion pressure was increased to 80 mm Hg for 30 minutes to establish a stable baseline. After the air had been raised to 80 mm Hg for 30 minutes, and vasoactive responses were reassessed. In separate experiments, responses to 5-HT were evaluated in the presence and absence of ketanserin (10\textsuperscript{-6} mol/L), a known 5-HT\textsubscript{1A} receptor antagonist. Also, after removal of the endothelium, changes in diameter to cumulative doses of Bay K 8644 (an opener of voltage-dependent Ca\textsuperscript{2+} channels, 10\textsuperscript{-10} to 10\textsuperscript{-6} mol/L) were assessed before and after preincubation with fluoxetine (2×10\textsuperscript{-6}, 5×10\textsuperscript{-6}, and 10\textsuperscript{-5} mol/L) for 5 minutes. All drugs were added to the vessel chamber, and final concentrations are reported. After responses to each drug subsided, the system was flushed with PS solution.

**Materials**

Fluoxetine was obtained from Research Biochemicals International Co. All other salts and chemicals were obtained from Sigma-Aldrich Co and were prepared on the day of the experiment. A 10\textsuperscript{-2} mol/L stock of glibenclamide was prepared by dissolving the substance in 30% wt/vol (2-hydroxypropyl)-\(\beta\)-cyclodextrin (Cyclolab R&D Ltd), adding a small volume of 40% vol/vol ethanol, and dissolving in buffer. Fluoxetine was dissolved in PS solution. Bay K 8644 was dissolved in ethanol and protected from light; experiments were carried out in the dark. The vehicle did not have vasoactive effects.

**Statistical Analysis**

Dilations were expressed as a percentage of the maximal dilation of the vessel, defined as the passive diameter at 80 mm Hg perfusion pressure in Ca\textsuperscript{2+}-free medium containing 10\textsuperscript{-5} mol/L EGTA and 10\textsuperscript{-4} mol/L SNP. Constrictions were expressed as a percentage of the baseline diameter. The 50% effective concentrations (EC\textsubscript{50}) were calculated from the log-logarithmic regressions of the cumulative dose-response curves of vasoactive agents. In the graphs, the arithmetic mean±SEM values were used. Statistical analyses were performed by ANOVA for repeated measures followed by Tukey's post hoc test or Student's t test. A value of P<0.05 was considered statistically significant.

**Results**

Rat small cerebral arteries developed a spontaneous myogenic tone in response to an increase in the perfusion pressure to 80 mm Hg without the use of any vasoactive agents. The active inner diameter of arterioles was 195±15 \, \mu m. The passive diameter of arterioles obtained in the same conditions but in the absence of Ca\textsuperscript{2+} (see Methods) was 270±17 \, \mu m.

**Responses to Fluoxetine: Role of Endothelium**

In a dose-dependent manner, fluoxetine (10\textsuperscript{-6} to 5×10\textsuperscript{-5} mol/L) elicited substantial dilations in arteries, with an EC\textsubscript{50} of 7.7±1.0×10\textsuperscript{-6} mol/L (Figure 1A). We have also found that removal of the endothelium had no significant effect on the dilation of arteries to fluoxetine (EC\textsubscript{50}=6.9±0.8×10\textsuperscript{-6} mol/L, P=NS; Figure 1A).

**Role of Potassium Channels**

Next, we tested the possible role of K\textsuperscript{+} channels in the vasodilator action of fluoxetine. Fluoxetine-induced dilations of arteries did not change significantly (EC\textsubscript{50}=6.3±1.5×10\textsuperscript{-6} mol/L, EC\textsubscript{50}=7.8±1.7×10\textsuperscript{-6} mol/L, and EC\textsubscript{50}=6.8±1.1×10\textsuperscript{-6} mol/L, respectively; Figure 1B) after preincubation and in the presence of the ATP-sensitive K\textsuperscript{+} channel inhibitor glibenclamide (10\textsuperscript{-5} mol/L), 4-AP (10\textsuperscript{-4} mol/L), or TEA (3×10\textsuperscript{-3} mol/L).

**Responses to 5-HT and Norepinephrine**

5-HT constricted the arteries in a concentration-dependent manner, with an EC\textsubscript{50} of 5.6±2.3×10\textsuperscript{-7} mol/L (Figure 2). In the presence of ketanserin (10\textsuperscript{-6} mol/L), a 5-HT receptor antagonist, 5-HT-induced constrictions were abolished (data not shown). Fluoxetine (10\textsuperscript{-6}, 2×10\textsuperscript{-6}, and 10\textsuperscript{-5} mol/L)
significantly \((P<0.01)\) reduced constrictor responses to 5-HT (Figure 2), whereas \(3\times10^{-3}\) mol/L fluoxetine essentially abolished the response.

In control conditions, norepinephrine constricted arteries in a concentration-dependent manner with an \(EC_{50}\) of \(4.25\pm0.9\times10^{-7}\) mol/L (Figure 3). Increasing doses of fluoxetine (\(2\times10^{-6}\) and \(3\times10^{-6}\)) significantly \((P<0.01)\) reduced responses to norepinephrine, whereas \(10^{-3}\) mol/L completely abolished the response.

Responses to Bay K 8644
We tested the arterial responses to Bay K 8644, a voltage-dependent \(Ca^{2+}\) channel opener. Bay K 8644 (\(10^{-10}\) to \(10^{-6}\) mol/L) constricted arteries in a concentration-dependent manner with an \(EC_{50}\) of \(5.6\pm2.1\) (0.42\(\times10^{-9}\) mol/L) (Figure 4). Constrictions to Bay K 8644 were significantly \((P<0.01)\) reduced in the presence of \(2\times10^{-6}\) or \(10^{-5}\) mol/L fluoxetine (Figure 4), whereas \(3\times10^{-3}\) mol/L completely abolished the response.

Discussion
This study demonstrates for the first time that fluoxetine, a serotonin reuptake inhibitor, dilates isolated rat small cerebral arteries. This dilation is not mediated by 4-AP–, TEA-, or glibenclamide-sensitive \(K^{+}\) channels or endothelium-derived dilator factors. Fluoxetine antagonized arterial constrictions to 5-HT and norepinephrine and had a substantial inhibitory effect on constrictions elicited by opening of voltage-dependent \(Ca^{2+}\) channels.

Fluoxetine Dilates Small Cerebral Arteries
In the present study, fluoxetine elicited a substantial concentration-dependent vasodilation in rat cerebral arteries (Figure 1). The vessels were isolated; thus, the effects of fluoxetine were induced by local factors intrinsic to the arterial wall. The concentrations of fluoxetine, which have a substantial cerebrovascular effect, are close to the upper range of therapeutic plasma concentrations (0.15 to 1.5 \(\mu\)mol/L).\(^{16,17}\) Previous studies reported that similar concentrations of norfluoxetine, the active metabolite of fluoxetine, are also present in the plasma of
fluoxetine-treated patients. In addition, recent data indicate that fluoxetine can be accumulated in the tissues. During chronic fluoxetine treatment, a concentration of fluoxetine 20 times higher than that in plasma has been detected in human brain, a finding that also indicates that fluoxetine effectively crosses the blood-brain barrier. Collectively, these findings suggest that during clinical treatments, the concentration of fluoxetine in the brain can reach levels that can potentially affect cerebral vascular tone and hence blood flow. Further studies are needed, however, to clarify the effect of fluoxetine on cerebral blood flow in patients with major depression. Nevertheless, our findings also suggest that the mechanisms of the medicinal actions of fluoxetine are more complicated than currently believed.

Possible Mechanisms of Action
Because there was no previous knowledge regarding the cerebrovascular effects of fluoxetine, we aimed to characterize the possible endothelial and smooth muscle mechanisms.

First, we aimed to elucidate whether the vasodilation to fluoxetine is mediated by release of factors from the endothelium. Removal of the endothelium, however, did not affect the fluoxetine-induced dilation (Figure 1). Therefore, we conclude that fluoxetine exerts its effect directly on the arterial smooth muscle.

Next, we hypothesized that opening of vascular K⁺ channels may be responsible for the dilation of small cerebral arteries to fluoxetine. Thus, we tested arterial responses to fluoxetine in the presence of glibenclamide (an inhibitor of ATP-sensitive K⁺ channels), 4-AP, or TEA (which inhibits several vascular K⁺ channels). However, the inhibition of these K⁺ channels did not significantly affect the vasodilator action of fluoxetine (Figure 2).

Fluoxetine is known to interfere with 5-HT receptors and/or signal transduction pathways in the central nervous system. Although 5-HT has a complex action on the cerebral vessels, the effect of fluoxetine on responses of isolated cerebral arteries to 5-HT has not been clarified. In the present study, first we confirmed previous findings showing that fluoxetine is a competitive and reversible antagonist of 5-HT₂ receptors. However, the finding that fluoxetine inhibited arterial constriction to norepinephrine also suggests that fluoxetine is not selective for 5-HT receptors and may affect postreceptor sites as well.

Previous studies demonstrated that other 5-HT uptake inhibitors, femoxetine and paroxetine, also antagonized 5-HT–induced contractions of rat intestinal preparations as well. Previous studies also suggested that fluoxetine is not selective for 5-HT receptors and may affect postreceptor sites as well.

Figure 1. Changes in diameter of isolated rat small cerebral arteries in response to cumulative doses of norepinephrine in the absence (control) or presence of 2×10⁻⁶ mol/L, 3×10⁻⁶ mol/L, or 10⁻⁵ mol/L fluoxetine. Each point represents the mean±SEM of 4 to 6 separate experiments.

Figure 2. Changes in diameter of isolated rat small cerebral arteries in response to cumulative doses of Bay K 8644 in the absence (control) or presence of 2×10⁻⁶ mol/L, 10⁻⁵ mol/L, or 3×10⁻⁵ mol/L fluoxetine. Each point represents the mean±SEM of 4 to 6 separate experiments.
inhibiting the influx of extracellular Ca$^{2+}$. This hypothesis is supported by previous in vitro findings that fluoxetine at a similar concentration (10$^{-5}$ mol/L) markedly attenuated the 5-HT-induced rise in [Ca$^{2+}$], in rat pulmonary arterial smooth muscle$^{13}$ and human neuronal cells.$^9$ Thus, the next logical step was to test the effect of fluoxetine on arterial responses to Ca$^{2+}$ channel activation.

In the present study, small cerebral arteries developed spontaneous myogenic constriction in response to an increase in intraluminal pressure. Pressure-induced myogenic response is known to depend primarily on Ca$^{2+}$ influx through voltage-dependent Ca$^{2+}$ channels in the vascular smooth muscle cells.$^{14,28,29}$ Thus, the finding that fluoxetine diluted cerebral arteries (inhibited myogenic tone) in an endothelium-independent manner suggests that fluoxetine interferes with Ca$^{2+}$ signaling mechanisms. To further support this contention, we tested the effects of fluoxetine on cerebral arterial constrictons elicited by Bay K 8644, an opener of norepinephrine but also constrictions to a Ca$^{2+}$ that fluoxetine inhibits not only responses to 5-HT and action of fluoxetine.

This study was supported by grants OTKA T-023863 and T-019206 from the National Science Research Foundation; ETT 524/97 and ETT 469/96 from the Health Science Council, Hungary; and HL-46813 from the National Heart, Lung, and Blood Institute, Bethesda, Md.

References

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Fluoxetine (Prozac) is widely used as an antidepressant in treating patients. The action of fluoxetine is presumably linked to its inhibition of central nervous system neuronal uptake of serotonin, thus making serotonin more available in the brain. Although serotonin is a chemical that affects mood, it is also a potent vasoconstrictor in the cerebral circulation. Despite extensive studies of the clinical pharmacological effects of the drug, the effect of fluoxetine on cerebral circulation remains essentially unexplored. In the accompanying article, Ungvari and coworkers demonstrated that fluoxetine causes dose-dependent vasodilation in the isolated branch of anterior cerebral arteries of rats. They provided the first evidence that fluoxetine diminishes the vasoconstrictor action of serotonin, norepinephrine, and Ca²⁺ channel openers. They concluded that fluoxetine interferes with the Ca²⁺ signaling mechanisms in the vascular smooth muscle. Findings from this in vitro study represent an important initial step in understanding the role of fluoxetine in the cerebral circulation.

Obviously, the vascular effect of fluoxetine needs to be further assessed in in vivo animal models. The anterior cerebral artery in rat is a conduit vessel, and its responses may not reflect those of the smaller downstream arterioles. Therefore, it is not certain that the dilator effect of fluoxetine on large vessels necessarily translates into improved blood flow in the brain in vivo. Another issue is that clinically, a low dose (20 mg/d) of fluoxetine is generally recommended in treating depressed patients. Whether or not this therapeutic dosage could consistently raise the plasma concentration of fluoxetine in patients to micromolar levels remains to be tested. On the basis of the findings of this study, below this level, fluoxetine seems to exert little or no cerebrovascular effect. In humans, it was reported that peak plasma concentrations of fluoxetine can reach ~1 µmol/L after a single oral 40-mg dose. Thus, it is not clear that at the usual therapeutic doses, fluoxetine would have significant cerebrovascular effects.

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doi: 10.1161/01.STR.30.9.1949

*Stroke* is published by the American Heart Association, 7272 Greenville Avenue, Dallas, TX 75231
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Print ISSN: 0039-2499. Online ISSN: 1524-4628

The online version of this article, along with updated information and services, is located on the World Wide Web at:
http://stroke.ahajournals.org/content/30/9/1949

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