Factors Influencing the Frequency of Fluorescence Transients as Markers of Peri-Infarct Depolarizations in Focal Cerebral Ischemia

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Background and Purpose—Peri-infarct depolarizations (PIDs) that occur in ischemic boundary zones of the cerebral cortex of experimental animals have been shown to promote rather than simply to indicate the evolution of the lesion and are especially prominent in the rat. To study the influence of one factor, species, on PID incidence, we compared the frequency of PIDs in a primate species, the squirrel monkey, with that in the cat after middle cerebral artery occlusion. Plasma glucose was reviewed as a possible cause of interexperiment variability in the cat experiments.

Methods—In open-skull experiments under chloralose anesthesia, changes in cortical fluorescence believed to indicate NADH/NAD redox state, as markers of PIDs, were recorded by serial imaging of the cortical surface in vivo for 4 hours after middle cerebral artery occlusion.

Results—Fluorescence transients occurred in squirrel monkeys at a frequency (mean ± SD) of 0.7 ± 0.8 hours⁻¹ (n = 5), which was not significantly less than in that observed in cats (1.3 ± 1.6 hours⁻¹, n = 8). Data from the cat experiments indicated a relationship between number of transients (dependent) and plasma glucose, with a striking increase in PID frequency in association with values of mean postocclusion plasma glucose <4.1 mmol/L (Mann-Whitney U = 15.0, P = 0.034); this observation agrees well with other published findings.

Conclusions—Transient changes in fluorescence strongly suggestive of peri-infarct depolarizations, either transient or terminal, occur and propagate in the ischemic cerebral cortex of a nonhuman primate. The results also suggest that the relationship of frequency of peri-infarct depolarizations with plasma glucose requires further examination, to confirm the finding and to determine a safe lower limit for a target range for control of plasma glucose if insulin is used in the management of patients with cerebral ischemia. (Stroke. 2000;31:214-222.)

Key Words: spreading cortical depression ■ NADH ■ middle cerebral artery occlusion ■ depolarization ■ hypoglycemia
primate brain, the present experiments were designed to examine and compare the frequencies of occurrence of fluorescence transients in the cat and in a nonhuman primate, the squirrel monkey. During analysis of the data, in a search for sources of interexperiment variability in PID frequency, we found evidence that PID frequency may be highly sensitive to quite modest reductions in plasma glucose. There is considerable published evidence to support and explain this finding, which, if confirmed, would have important implications for the management of plasma glucose levels in patients with acute traumatic or ischemic brain injury.

Materials and Methods

Animals and Housing
Adult squirrel monkeys (Saimiri sciureus) weighing 0.9 to 1.4 kg were housed in one group, and adult colony-bred male cats weighing 2.8 to 4.0 kg were housed in groups of 3 to 9 in UK Home Office–approved caging. The environment was maintained at 19°C to 22°C and a relative humidity of 55±3%, respectively, with a 14-hour/10-hour light/dark cycle (light on from 6 AM to 8 PM). Food and water were available ad libitum, and food was withdrawn 16 hours before surgery.

Surgical Preparation and MCAO
In a protocol approved by the Home Office, halothane was used to induce anesthesia in cats and squirrel monkeys (4% in a mixture of 70% N₂O and 30% O₂), and each animal was then initially allowed to breathe halothane (2%) spontaneously via a face mask. Rectal temperature was maintained at 37°C (36.5°C to 38°C) with a heating blanket (Harvard Apparatus). The left femoral vein was cannulated for fluid administration, and the left femoral artery was cannulated for the continuous monitoring of arterial blood pressure and for repeated blood sampling for serial measurements of arterial blood glucose concentration, PaO₂, PaCO₂, [HCO₃⁻], and pH. Arterial blood (200 μL) was collected from the left femoral artery of the animals for immediate analysis at 30-minute intervals during the experiment by use of an IL1304 analyzer (Instrumentation Laboratories). Blood glucose was measured electrochemically with Exactech strips (Medisense, Coleshill). A tracheostomy was performed, and the animal was intubated and ventilated mechanically with paconumonium (bolus: 0.02 mg · kg⁻¹ · maintenance: 0.06 mg · kg⁻¹ · h⁻¹) for neuromuscular blockade. Induction anesthesia was replaced with intravenous anesthesia (2%). Rectal and paraffin pool temperatures, blood pressure, glucose, and pH recorded during the experiment were analyzed by MANOVA. Significance of any relationship between fluorescence transient incidence and mean ischemic plasma glucose was tested by a number of approaches, as described in Results.

Results

Physiological Variables
In cats, PaCO₂ (33.8±3.9 to 33.4±5.1 mm Hg, mean±SD), arterial pH (7.35±0.5 to 7.34±0.3), and HCO₃⁻ (18.7±1.3 to 18.2±2.2 mmol/L) changed very little during the experiments, although there was a trend toward reduction in mean arterial pressure in cats, from 77±24 to 60±17 mm Hg at 4 hours after occlusion (despite supplemental infusions of colloid volume expander). Mean maximum and minimum brain surface temperatures in cats were 37.0±0.5°C and 36.3±1.1°C, respectively. All variables remained stable in squirrel monkeys. Plasma glucose data in cats are described below.

Propagated Fluorescence Transients
The times required for surgical preparation and for imaging during ischemia precluded imaging for substantial periods of time before MCAO. The mean sampling duration was 11 minutes in both species, and no transients were seen.

The incidence of transients after MCAO in cats and squirrel monkeys are shown in the Table, together with their distribution in cats by gyrus and an assessment of whether the transient originated on the gyrus or had propagated from a neighboring gyrus. Transients occurred in squirrel monkeys at a frequency of 0.7±0.8 hours⁻¹ (n=5), which was not significantly less than that observed in cats (1.3±1.6 hours⁻¹);
There was great variability in the incidences, both in cats and in squirrel monkeys. In cats, the range was from 1 to 20 in 4 hours; in 1 monkey, 8 PIDs were recorded over 4 hours (Figure 2); in another, none were observed.

In cats, it was possible to describe the initial location and subsequent propagation of fluorescence transients by reference to the convenient gyral anatomy and established topography of infarction and penumbra in the MCAO stroke model. In the 8 cat experiments, an aggregate of 10 new transients occurred on the marginal gyrus, 18 on the suprasylvian gyrus, and 17 on the ectosylvian gyrus (11 in experiment 7, within a relatively short period between 70 and 140 minutes after MCAO, when plasma glucose was in the range of 3.4 to 3.8 mmol/L). In the great majority, the direction of propagation was centrifugal from the core, or circumferential on the gyrus of origin, but 7 transients were seen to spread toward the core area (Figure 1).

In squirrel monkeys, the cortical topography cannot be defined as readily as in cats, but the general onset, resolution, and propagation characteristics of the transients observed were similar to those in cats. In the experiment in which the most transients were seen, we observed propagation of a transient around the site of delayed fluorescence recovery from a previous transient (Figure 2); it was evident that this area subsequently repolarized, because it was invaded later by a third transient (Figure 2, f through h).

Plasma Glucose
In cats, plasma glucose was measured at MCAO and hourly thereafter. Although the values for SD for plasma glucose do not appear wide, values remained low in experiments 6 and 7, with mean values of 3.48 and 3.78 mmol/L, respectively; the highest incidence of PIDs occurred in these 2 experiments (Table; Figure 3). The relationship of PID number with plasma glucose was examined in cats (Table; Figure 3), and a possible threshold dependence of fluorescence transient number on plasma glucose was clearly evident. The relationship failed to reach statistical significance when analyzed by linear or logarithmic regression. However, in the light of existing published work suggesting that perilesion depolarizations might be promoted by reduced availability of glucose to the brain (please see Discussion), we dichotomized the experiments according to a putative mean plasma glucose threshold for increased transient frequency of 4.10 mmol/L; frequency of transients was significantly higher in experiments in which mean plasma glucose at and after MCAO was <4.10 mmol/L (Table; Figure 3) (Mann-Whitney U=15.0, P=0.034).

Discussion
Since spontaneous transient increases in extracellular potassium were first described in the cortical ischemic penumbra after MCAO in baboons and cats, a very extensive investi-
Negative effort has been devoted to exploring their nature. It is widely agreed that such cation transients are markers of focal neuronal depolarizations propagating across the cerebral cortex, now usually designated peri-infarct depolarizations (PIDs). Although sharing certain features (depolarization, cortical propagation, and transient increases in extracellular potassium and decreases in extracellular calcium) with Leão’s cortical spreading depression (CSD), PIDs differ from CSD in 3 critical aspects. First, the intense hyperemic transient of CSD is not seen in association with PIDs; this is most readily attributable to proximal vascular occlusion, with incomplete compensation by collateral flow. Second, and as a consequence of diminished hyperemia, the cortical tissue PO2 transient, which is positive in CSD, is reversed to a transient decrease in PIDs. Third, whereas CSD is not associated with neuronal damage, there is a linear relationship of infarct size with number of PIDs, and PID number is the determining variable in this relationship.

It is now very clear from a large volume of literature that CSD is readily elicited in the (lissencephalic) rat brain but less readily in the gyrencephalic cat or primate brains. It is, however, unclear whether the same comparison can be applied to PIDs, and this study was undertaken to confirm that PIDs do indeed occur in the ischemic primate brain and to compare the frequencies of PIDs in the cat and primate brains. The relevance of this issue to human disease states and their treatment is discussed below. We found clear evidence for PIDs in squirrel monkeys subjected to MCAO, with a frequency that is not significantly different from that in cats; possible reasons for this apparent similarity are discussed later.

An important observation was the considerable interexperimental variability in PID frequency in both species. Although it suggests a statistically significant dependence of PID frequency on mean postocclusion plasma glucose level in cats, the experiments were not originally designed to examine this issue. However, we believe both that attention should be focused on this issue because of its relevance to the proper application of one proposed treatment regimen (glucose-insulin-potassium) in clinical management of patients with stroke and that the question needs to be examined in further, specifically designed experiments.

Detection of PIDs by Fluorescence Imaging

In considering our results, some discussion of the method we have adopted for PID detection and the interpretation of the resulting data is first necessary. CSD is associated with a transient oxidation of the NAD/H couple, resulting in depression of NADH fluorescence (emission maximum 450 nm); thus, depression of this fluorescence may be used as a surrogate marker of CSD in the normally perfused cortex. Using an imaging method for NADH fluorescence (rather than single-point detection with a fluorometer), we have described 2 patterns of fluorescence change after MCAO: either sustained increases in fluorescence or transient increases (or sometimes decreases; please see below) that resolve toward baseline, sometimes with an undershoot, over periods of some 2 to 10 minutes. We interpret sustained

### Occurrence of New and Propagated Fluorescence Transients in Relation to Mean Plasma Glucose (Cats Only) at and After MCAO

<table>
<thead>
<tr>
<th>Fluorescence Transients by Gyrus</th>
<th>Marginal</th>
<th>Suprasylvian</th>
<th>Ectosylvian</th>
<th>Mean Plasma Glucose, mmol/L±SD</th>
<th>Total New Transients</th>
<th>New Transients per Hour</th>
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<tr>
<td>Cats</td>
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<td>New</td>
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<td>4.96±1.17</td>
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<td>5.8±0.88</td>
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<td>5</td>
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<td>2</td>
<td>2</td>
<td>6.5±1.81</td>
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<td>7</td>
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<td>8</td>
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<td>Squirrel monkeys</td>
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<td>0.7±0.8</td>
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*New* indicates transient that originates on the gyrus; *Pr*, propagated from a neighboring gyrus; and *CL*, confidence limits. Where the electrode recorded a value of plasma glucose as “low,” a value of 2.0 mmol/L was assumed for purposes of calculation.
fluorescence increases as most probably coinciding with terminal depolarization, i.e., development of core conditions, well characterized previously by increases in extracellular potassium or negative changes in DC potential (sustained in both cases). The second pattern we observed was of multiple transient increases in fluorescence in the suprasylvian and middle/posterior marginal gyri, although on some occasions, a transient that propagates into the anterior marginal gyrus (anterior cerebral artery territory) will there reverse its polarity to a primary decrease in fluorescence. Although our method does not provide a quantitative measure of changes in NAD/NADH redox potential, the topographical pattern of transient changes is consistent both with reduction of the couple in the penumbra, where flow recruitment in response to the transient is restricted, and with oxidation, where a transient has propagated into normally perfused cortex. The reversal in polarity is closely comparable to the opposite polarities of tissue PO2 transients in CSD versus PIDs. Thus, the imaging method we used here not only marks the occurrence of a depolarization but also tracks its propagation and, from the polarity, indicates a distinction between PID (with the implicit risk of promoting tissue damage) and, in normally perfused cortex, an NADH oxidation transient that may reasonably be interpreted as CSD. Because the imaging method samples almost the entire penumbra (in cats), it is a more comprehensive sampling tool than single or dual intracortical electrodes. The longest interval between acquisition sequences was \(4\) minutes; because propagation rates of transients are in the range of \(1\) to \(3\) mm/min and duration at a given cortical site is \(1\) to \(3\) minutes, and because imaging continued for \(95\%\) of occlusion duration, it is unlikely that any transient that commenced during an interval between sequences would escape detection.

**Occurrence and Topography of Fluorescence Transients**

No transients were seen during the time available in these experiments for observation before MCAO. In an unpublished review of 32 earlier experiments with the same anesthetic method and ion-elective electrodes (often at 2 sites) for detection of depolarizations, we found a mean incidence of 0.4 events/hour per experiment before MCAO. However, in the majority of those experiments, intracortical hydrogen polarography electrodes (diameter 125 \(\mu\)m) were in use (the focal cortical trauma associated with needle or electrode insertion is a classic method for the induction of CSD). The question arises as to whether CSD might have
been induced in the present studies by possible trauma during creation of the preparation; however, scrupulous attention was paid to craniotomy technique in these experiments. In particular, exposure of the MCA, with opening of the adjacent arachnoid, secured significant drainage of cerebrospinal fluid and hence allowed the brain to fall away from the dura, conferring additional protection.

The incidence of fluorescence transients in cats after MCAO in the present series was 1.3 events per hour; our review of an earlier series (n=32) yielded a value of 1.26 per hour. Much of the aggregate number of fluorescence transients seen on the ectosylvian gyrus in the present work is accounted for by experiment 7, and in the remaining experiments, we attribute the low transient incidence on this gyrus to early terminal depolarization, undetected at the time of occlusion (because of the need for visible light during and immediately after the surgical MCAO procedure).

Factors Affecting the Frequency of Transients
The frequency of transients was very variable in these experiments, and brief mention must first be made of factors that are already recognized as influencing PID frequency. The incidences of potassium-evoked CSDs in the normally perfused cat brain and of PIDs in cats (with MCAO) are appreciably reduced by halothane,11 which may be due in part to the capacity of this agent to uncouple glial gap junctions.25 We therefore restricted any use of halothane after initial induction of anesthesia to rare, transient supplementation of chloralose, at a maximum inspired concentration of 0.75%. Pool temperature was rigorously controlled in the present cat experiments.9,26 Despite our considerable efforts to achieve uniform conditions, PID frequency varied widely between individual animals in a species (eg, Table, cats); the experience of Gill et al6 with rats undergoing MCAO was similar to our observations in hypoglycemia and by hypoglycemia, and given the dependence of infarct size on PID number and the likely dependence of glutamate homeostasis on tissue glucose, it is possible that their observations in hypoglycemia could be explained on the basis of increased numbers of PIDs.

Plasma Glucose
We found a statistically significant dependence of fluorescence transient frequency on plasma glucose level, and there is already considerable evidence that supports and explains this finding. Nedergaard and Astrup4 showed that the rate of PIDs in the penumbra after MCAO in rats was 3.8±1.8 at “normoglycemia” (9.3 mmol/L) and 0.3±0.4 at hyperglycemia (32.5 mmol/L), and they observed increased glucose phosphorylation in the same region; they suggested that tissue glycopenia was likely to be present, and the high level of plasma glucose required to reduce PID frequency is worthy of note. The present results suggest that the same principle may apply in cats, but at levels of plasma glucose likely to be encountered in clinical practice (especially if an attempt is made to control hyperglycemia to reduce brain acidosis). Given that ATP yield from anaerobic utilization of glucose is one nineteenth of that available from aerobic oxidation, anaerobic glucose utilization must be expected to increase and to become rate-limited by glucose availability. Mies and Paschen27 showed that after a wave of CSD in the normally perfused rat brain in vivo, the tissue glucose pool remained depressed for ≥160 seconds. There is also evidence to link diminished tissue glucose availability with destabilization of glutamate (and by implication cation) homeostasis in the extracellular space, which is a possible cause of PID initiation. Swanson et al28 showed that microdialysis of glucose into globally ischemic deep gray matter could reduce ischmic glutamate release to 20% of the value seen with glucose-free dialysate. De Courten-Myers and colleagues29 found that MCAO infarct size in cats was increased both by hyperglycemia and by hypoglycemia, and given the dependence of infarct size on PID number and the likely dependence of glutamate homeostasis on tissue glucose, it is possible that their observations in hypoglycemia could be explained on the basis of increased numbers of PIDs.

A Species Hierarchy for PID Frequency, and Its Biological Basis?
What is the basis for the suggestion that PIDS might be less frequent in humans than in the MCAO models? There is experimental evidence that species-related hierarchies may exist for more than 1 relevant variable. First, regarding PID frequency, Nedergaard and Astrup4 recorded 5.1±2.3 PIDs in their observation period of 80 minutes (3.8 PIDs per hour) in rats (MCAO, pentobarbital anesthesia). In only 1 cat experiment have we observed a frequency in the range of 3.5 to 5 transients per hour; this was in an experiment (number 7, Table) in which plasma glucose was low, and the mean value for the present study group was 1.3 PIDs per hour. Our earlier, unpublished review of 32 MCAO experiments in cats (detection of PIDs with potassium-sensitive electrodes) also yielded a value of 1.3/h. The present experiments represent, so far as we are aware, the first specific attempt to establish a value for PID frequency in primate MCAO experiments. That we have been unable to demonstrate a difference in PID frequency between cats and squirrel monkeys may be due to interexperiment variability but possibly also to the fact that the sizes of the squirrel monkey and cat brains are similar. In consequence, their glial:neuronal ratios and PID frequencies may be similar (please see below).

A second variable for which a hierarchy may exist is the flow threshold for homogenous ischemic cell change or massive, sustained potassium release after MCAO. In rats, a flow value of 24 mL · 100 g⁻¹ · min⁻¹ is required for homogenous infarction.31 In cats, the flow threshold for sustained, major potassium ion release is ~16 mL · 100 g⁻¹ · min⁻¹,2 whereas in the baboon, the corresponding value lies in the range of 8 to 11 mL · 100 g⁻¹ · min⁻¹.

Third, in view of the role of the glia in homeostasis of extracellular concentrations of both potassium and glutamate, the ratio of glia to neurons in the cortex is a further potentially relevant variable for which a species hierarchy has been proposed, and it is possible that interspecies variation in this ratio accounts for the differences between cats and rats discussed above. Tower and Young30 described a hierarchy for glial:neuronal ratio in a broadly based group of mammalian species and demonstrated a striking, linear relationship of glial:neuronal ratio with brain size, the hierarchy being: mouse, rat, guinea pig, rabbit, cat, dog, monkey (macaque),
ox, horse, human, elephant, fin whale. Thus, it may be increasing brain size rather than membership in the primate order that reduces PID frequency.

How might an increased glial:neuronal ratio confer such potential benefits as lower PID frequency and infarction flow thresholds? The glia are essentially the only location of glycogen in the cortex,^{12,33} are fully capable of anaerobic glycolysis in vitro^{14} and generation of pyruvate or lactate, and contribute substantially to homeostasis of potassium^{15} and glutamate^{16} in the extracellular space. These functions must necessarily become critical (and an increased glial:neuronal ratio an advantage) at the particular stage of progressive focal ischemia when anaerobic metabolism has become the sole source of ATP and glycolytic rate, now enhanced, outpaces glucose availability.

It must be recognized that the 3 sets of findings on which the discussion above is based lack the strength of results from a single, specifically designed study, but it seems that any consideration of species differences in PID frequency must take account of the issue of brain size in relation to glial:neuronal ratio.

**PDIs in Humans: Implications for Treatment of Acute Brain Injury**

Were extracellular ion/neurotransmitter homeostatic capacity indeed related to brain size and glial:neuronal ratio and the proposed hierarchy a reality, a rather lower PID frequency might be predicted for humans than for the experimental species. Evidence for the occurrence of PDIs in humans is extremely limited, but this is possibly due to lack of appropriate methods for detection. There is one recent report^{12} of transient changes in NADH, extracellular potassium, and laser Doppler flow in the frontal cortex of 1 patient of 14 with severe head injury. We and others agree the transients reported are suggestive of a CSD-like phenomenon, but the significance remains unclear; the finding appears to have been a rarity, and the changes observed may have been preterminal. However, in the patient concerned, the PID detection system was sited over the right frontal convexity, whereas the traumatic lesion was left parietal, and the data may therefore underestimate the frequency of perilesion PDIs.

The principal conclusion from this study is that cortical fluorescence transients believed to be markers of PDIs occur in the nonhuman primate brain. This supports the earlier report by Branston et al^{1} of spontaneous extracellular potassium transients in the MCAO penumbra in baboons (commented on at the time as resembling spreading depression), thus confirming that the primate brain, when “injured,” is capable of supporting propagated CSD-like phenomena. There are 2 aspects of the findings with potential implications for the clinical management of patients with acute brain injury. First, if suppression of PDIs is the main mechanism of neuroprotection by the EAAA in experimental models of brain injury, and if the occurrence of PDIs in humans is uncertain, it is perhaps not surprising that phase 3 clinical studies of neuroprotection with EAAA should have proved unsuccessful to date. It follows that more precise definition of the conditions under which PDIs occur in the human brain is needed, possibly leading to designation of the presence of PDIs as a criterion for entry of a patient into a clinical trial of EAAA. Second, interest is being expressed in the establishment of a clinical trial of control of reactive hyperglycemia with insulin in patients with acute stroke,^{37} and a recent review of this issue recommends a target plasma glucose in the range of 80 to 150 mg% (4.4 to 8.3 mmol/L).^{38} The lower limit of this range is adjacent to the plasma glucose threshold for an increase in PID frequency suggested by our results, implying that the safe target range for control of plasma glucose in brain injury in humans may be narrower than is currently envisioned. There is therefore a need for an experimental study specifically designed to examine the relationship between plasma glucose and PID frequency that is suggested by our findings.

**Conclusions**

We have found clear evidence that cortical fluorescence transients, believed to indicate peri-infarct depolarizations, occur in the nonhuman primate brain during focal cerebral ischemia. Furthermore, we found considerable interexperimental variation in PID frequency within the 2 species we studied, so that the possibility of a modest difference in PID frequency between cats and nonhuman primates is not excluded by our results.

Our data suggest that the risk of a substantial increase in frequency of PDIs associated with focal cerebral ischemia rises when plasma glucose falls toward 4 mmol/L. This suggestion is consistent with other studies and may offer at least a partial explanation for other published data relating low plasma glucose with increased infarct size. If confirmed by deliberate plasma glucose reduction in specifically designed experiments, the finding would influence the application of the therapeutic concept of control of plasma glucose in patients with acute brain injury. It also follows that any rigorous interspecies comparison of “natural” PID frequency must be controlled for plasma glucose as well as for anesthesia, temperature, and other factors affecting excitatory neurotransmission, such as pH.

Clear information on the occurrence, frequency, and properties of PDIs in human disease states would be an important addition to our understanding of the pathophysiology of ischemic (and traumatic) brain injury in humans and would provide important guidance on the value of continued efforts to develop neuroprotection strategies based on the use of EAAAs. The scientific case for their use in acute brain lesions in humans remains to be either established or dismissed.

**Acknowledgments**

We thank SmithKline Beecham, HeadFirst, and the Golden Charitable Trust for financial support. We are grateful to Prof Brian Everitt (Institute of Psychiatry) for his helpful statistical advice and analysis.

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the number of PIDs observed have protective effects during focal ischemia. However, these studies do not establish a cause-and-effect relationship for PIDs and increase injury. The presence of PIDs may simply be a marker of a more severe ischemic injury. The study of Busch et al., lends more support to this idea by demonstrating that repeated episodes of cortical spreading depression induced by potassium are associated with a larger volume of tissue injury. However, because these studies involved induced spreading depression, the role that depolarizations which occur spontaneously after focal ischemia play in worsening injury is still not entirely clear.

In attempting to identify factors that might explain the variability of PID frequency among individual animals, the authors also observed an interesting association between plasma glucose concentration, and the frequency of PIDs in the cat studies. The animals with low plasma glucose had a significantly higher frequency of PIDs after middle cerebral artery occlusion. Because the numbers of animals in the study are small, and the studies were not really designed to examine this relationship, the association must be considered speculative. Although additional studies are needed to definitively answer this question, this issue of optimal plasma glucose concentration is very timely. Hyperglycemia has long been associated with a higher mortality rate and poorer neurological recovery after stroke. In a recently published analysis of 1259 patients involved in the trial of ORG 10172, a higher plasma glucose concentration on admission was significantly associated with a poorer neurological recovery at 3 months. This relationship was especially strong for the subgroup of patients with nonlacunar stroke. A recent pilot trial has demonstrated the feasibility of reducing plasma glucose concentrations after acute stroke with glucose potassium insulin infusion. Because trials are conducted to study the efficacy of this treatment strategy for stroke, it will be important to consider that there may also be adverse consequences for a plasma glucose concentration that is too low for the ischemic brain.

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Stroke. 2000;31:214-222
doi: 10.1161/01.STR.31.1.214

Stroke is published by the American Heart Association, 7272 Greenville Avenue, Dallas, TX 75231
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Print ISSN: 0039-2499. Online ISSN: 1524-4628

The online version of this article, along with updated information and services, is located on the World Wide Web at:
http://stroke.ahajournals.org/content/31/1/214

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