Noninvasive Measurement of Cerebral Blood Flow With 99mTc-Hexamethylpropyleneamine Oxime Single-Photon Emission Computed Tomography and 1-Point Venous Blood Sampling

Yoshinari Isaka, MD; Satoshi Furukawa, MD; Hideki Etani, MD; Etsuko Nakanishi, MD; Yosuke Ooe, MD; Masatoshi Imaizumi, MD

Background and Purpose—The arterial and venous blood concentration of technetium 99m–labeled hexamethylpropyleneamine oxime (99mTc-HMPAO) reaches an equilibration more rapidly than other CBF tracers. We hypothesized that 99mTc radioactivity of a venous sample at equilibrium, which is similar to that of an arterial sample, would allow estimation of the integrated input function for the clinical measurement of CBF by use of single-photon emission CT.

Methods—in 53 patients with stable cerebrovascular disease, the radioactivity of a venous sample 5 minutes after injection of 99mTc-HMPAO was correlated with 5-minute arterial blood radioactivity and the first 5 minutes of the integrated arterial curves of the lipophilic tracer. The measured CBF values were compared with those of xenon 133.

Results—the radioactivity of 5-minute venous blood was almost equivalent to that of 5-minute arterial blood (r² = 0.987; y = 0.993x + 1.63; P < 0.0001). The correlation between the venous blood radioactivity and the integrated arterial lipophilic fraction was excellent (r² = 0.935, P < 0.0001). A strong correlation was obtained between 99mTc-HMPAO and 133Xe CBF values (r² = 0.825, P < 0.0001). CBF values were reproducible (coefficient of variation, 8.6%).

Conclusions—This approach is fast, simple, and an alternative to continuous blood sampling in clinical quantitative 99mTc-HMPAO CBF studies. (Stroke. 2000;31:2203-2207.)

Key Words: cerebral blood flow ■ cerebrovascular disorders ■ tomography, emission computed
the brain, R is the retained fraction of lipophilic radioactivity in the brain, and CaL is the lipophilic tracer radioactivity in the arterial blood.

If E and R are known and steady-state radioactivity of the venous blood has close associations with CaL and $f_i$, CaL(t), Equation 1 can be solved simply in terms of the tracer concentrations in the venous blood and in the brain as follows:

$$\text{CBF (mL/100 g/min)} = \frac{E \cdot k \cdot C_v(5)}{R} \cdot \frac{\text{SPECT } [\muCi/100 g]}{\text{Arterial blood [CI/min/mL]}}$$

where $E = 0.72, R = 0.54, C_v(5)$ is the radioactivity of the venous blood at steady state (5 minutes), and k is the slope of the regression line of

$$\int_0^5 \text{CaL(t)} \, dt$$

with $C_v(5) = 4.62$.

**Patients**

CBF was measured in 53 patients with stable cerebrovascular disease (CVD) (28 men and 25 women; mean age 70.6 [range 47 to 87] years). No patients had a renal insufficiency. Diagnosis was based on criteria from the National Institute of Neurological Disorders and Stroke. Six patients had had a transient ischemic attack, 33 a lacunar infarction, 1 an atherothrombotic infarction, 2 a cerebral hemorrhage, and 11 a poststroke dementia. All patients gave informed consent for participation in the study.

**CBF Studies**

$_{99m}$Tc-HMPAO was formed by reconstituting a commercial vial of HMPAO (Cerebrotec, Amersham Health Care) with 5 mL of 15 to 30 mCi (555 to 1110 MBq) fresh $_{99m}$Tc pertechnetate. Arterial blood samples were obtained from a small catheter placed in the brachial artery. The sampling was performed every 15 seconds for the first 2 minutes and every 30 seconds for the next 3 minutes after intravenous injection of 10 mCi $_{99m}$Tc-HMPAO. Arterial blood was collected in vials containing 1 mL of octanol, and the arterial concentration of lipophilic tracer was measured by the rapid octanol extraction technique. Venous blood also was sampled 5 minutes after injection.

SPECT scanning was then started with a single-head rotating camera (GCA-901A, Toshiba) with a resolution of 17 mm full-width half-maximum, using a low-energy, high-resolution collimator. Sixty views, 20-second frames collected over 360°, were recorded into a 128 × 128 matrix. Transaxial sections at 2.7-mm intervals were used to reconstruct computed images 10.8 mm thick in planes parallel to the orbitomeatal line.

In all patients, CBF was measured within 2 weeks of SPECT examination with the intravenous $^{133}$Xe method and a helmet-type parallel 32-detector system (BF 1400, Valmet). Sixteen detectors were symmetrically placed in each hemisphere. Approximately 20 mCi of $^{133}$Xe-labeled saline was injected into the antecubital vein. The clearance of the head curve was recorded over 15 minutes from each head detector as well as from a separate detector that monitored the radioactivity in expired air. The clearance curve was fitted by a 2-compartment deconvolution, with end-tidal $^{133}$Xe counts as an input function.

Integrated lipophilic activity was determined by summing the area under the measured concentration curve CaL(t) between 0 and 5 minutes and was calibrated and converted to units of microcuries per minute per millilitre. The time course of the ratio of lipophilic to nonlipophilic radioactivity was expressed as a percentage of the total radioactivity in each arterial sample. Venous blood radioactivity 5 minutes after $_{99m}$Tc-HMPAO injection was compared with that of arterial blood and the integrated lipophilic tracer activity up to 5 minutes after injection. Eight subjects were scanned twice, 1 week apart, to determine the reproducibility of CBF values. An ROI in the whole brain was defined by incorporating all pixels that were >30% of the maximum counts per pixel on a single SPECT section containing the basal ganglia. Tracer concentration measured within the whole brain was expressed as $\mu$Ci (37 kBq)/100 g, assuming a brain weight of 1270 g. A mean whole-brain $^{133}$Xe CBF value was calculated for each subject from fast flow (f1), slow flow (f2), and the obtained weight ratio between gray and white matter (w1/w2).

Results were analyzed by using the Pearson equation and linear regression. Data were presented as mean±SD. Statistical significance was set at P<0.05.

**Results**

As shown in Figure 1, the 5-minute tracer counts of venous blood sample were almost equivalent to those of the arterial blood sample: 364.4 ± 137.1 × 10⁴ counts per minute per milliliter (cpm/mL) versus 567 ± 137.2 × 10⁴ cpm/mL ($r^2 = 0.987, P<0.0001; y = 0.993x + 1.63$). The table shows the time course of the concentration of lipophilic tracer in the arterial blood. The percentage of the lipophilic radioactivity was highest after 15 seconds, then decreased rapidly within

<table>
<thead>
<tr>
<th>Time, s</th>
<th>Lipophilic%</th>
<th>Nonlipophilic%</th>
</tr>
</thead>
<tbody>
<tr>
<td>15</td>
<td>85.9</td>
<td>14.1</td>
</tr>
<tr>
<td>30</td>
<td>73.6</td>
<td>26.4</td>
</tr>
<tr>
<td>45</td>
<td>42.9</td>
<td>57.1</td>
</tr>
<tr>
<td>60</td>
<td>19.4</td>
<td>80.6</td>
</tr>
<tr>
<td>75</td>
<td>15.3</td>
<td>84.7</td>
</tr>
<tr>
<td>90</td>
<td>5.5</td>
<td>94.5</td>
</tr>
<tr>
<td>105</td>
<td>7.8</td>
<td>92.2</td>
</tr>
<tr>
<td>120</td>
<td>2.7</td>
<td>97.3</td>
</tr>
<tr>
<td>150</td>
<td>4</td>
<td>96</td>
</tr>
<tr>
<td>180</td>
<td>6.5</td>
<td>93.5</td>
</tr>
<tr>
<td>210</td>
<td>1.8</td>
<td>98.2</td>
</tr>
<tr>
<td>240</td>
<td>1.3</td>
<td>98.7</td>
</tr>
<tr>
<td>270</td>
<td>0.1</td>
<td>99.9</td>
</tr>
<tr>
<td>300</td>
<td>0</td>
<td>100</td>
</tr>
</tbody>
</table>

Lipophilic% indicates the percentage of total radioactivity extracted by octanol; nonlipophilic%, the percentage of radioactivity extracted, primarily in the hydrophilic form.
the subsequent 1 minute, and reached nearly zero after 4 minutes. There was a strong correlation between the 5-minute radioactivity of venous blood and the integrated arterial lipophilic tracer activity up to 5 minutes after injection ($r^2 = 0.935$, $P < 0.0001$; $y = 4.62x - 0.14$) (Figure 2). The $y$-intercept of this regression line was sufficiently small compared with the magnitude of integrated arterial lipophilic tracer activity.

Average values of $^{99m}$Tc-HMPAO CBF and $^{133}$Xe CBF in the whole brain were 35.6 ± 7.3 and 37.0 ± 8.3 mL·100 g$^{-1}$·min$^{-1}$, respectively. A close correlation was observed for $^{99m}$Tc-HMPAO CBF versus $^{133}$Xe CBF ($r^2 = 0.825$, $P < 0.0001$; $y = 0.804x + 5.84$) (Figure 3).

Mean whole-brain CBF values obtained in the second measurement (33.5 ± 7.8 mL·100 g$^{-1}$·min$^{-1}$) did not differ significantly from those obtained in the first (34.2 ± 4.7 mL·100 g$^{-1}$·min$^{-1}$). The reproducibility of CBF values was good ($r^2 = 0.622$, $P < 0.005$; $y = 0.987x + 0.9$). The coefficient of variation for CBF was 8.6%.

**Discussion**

We found that the radioactivity of the 5-minute venous blood sample was almost identical to that of the 5-minute arterial blood sample and was proportional to the integrated arterial lipophilic radioactivity of $^{99m}$Tc-HMPAO. At 5 minutes, equilibration between brain and blood is fully achieved, because the ratio of radioactivity in the arterial blood versus that in the venous blood reaches a plateau. This rapid decline of lipophilic $^{99m}$Tc-HMPAO from the blood can be caused by rapid conversion to hydrophilic metabolites and binding to some blood component. In rat biodistribution data, a high proportion of the radioactivity remaining in the blood appears to be trapped within red blood cells. It is possible that the mechanism for entrapment within these cells is similar to that of brain retention. This may explain why radioactivity of the 5-minute blood sample is strongly related to the integrated lipophilic radioactivity from the plasma input curve. Pupi et al. found that fractional brain uptake derived from the injected dose was not a reliable indicator of $^{99m}$Tc-bicisate CBF, suggesting that intracellular and extracellular radioactivities may vary from individual to individual. Considering the intersubject variability of kinetic parameters in the blood and in the brain, the need of blood sampling and counting for CBF determination was indicated.

We were able to measure CBF with a shorter examination period than those in the previous methods that used continuous arterial blood sampling and/or kinetic analysis of the tracer with a high-performance gamma camera system. This was made possible through the substitution of 1-point venous blood sampling for arterial blood sampling to obtain an input and no need for dynamic SPECT data acquisition. SPECT counts/integrated lipophilic activity reflects essentially the steady-state influx constant of Patlak and Blasberg, which can be measured with a single SPECT scanning. In other words, net SPECT counts/integrated lipophilic activity is expected to be a quantitative index of CBF. Data acquisition time can be reduced further by using a multi-ring SPECT camera. CBF measurement can be completed within 25 minutes, and thus the use of the present method as a test for brain function would not delay acute stroke therapy.

For radiolabeled microspheres, the extraction fraction is $\approx 100\%$ in the brain tissue of humans, and tracer uptake versus CBF have a linear relationship. However, this method is not suitable for human use because of its invasiveness. Most of the limitations of CBF tracers, including $^{133}$Xe-, $^{99m}$Tc-, or $^{123}$I-labeled CBF tracers and $H_2^{15}$O arise from the nonlinear relationship between true CBF and measured radionuclide concentration. Over a CBF range of 20 to 120 mL·100 g$^{-1}$·min$^{-1}$, $^{133}$Xe CBF correlates linearly with true CBF. At a CBF level that corresponds to normal regional CBF for human cortex, 50 mL·100 g$^{-1}$·min$^{-1}$, $^{99m}$Tc-HMPAO has a first-pass extraction of approximately $\geq 70\%$. The underestimation of CBF in the present method appeared to be less at whole-brain CBF levels of up to 50 mL·100 g$^{-1}$·min$^{-1}$. When CBF exceeded 50 mL·100 g$^{-1}$·min$^{-1}$, CBF was underestimated because of the limitation of brain permeability to $^{99m}$Tc-HMPAO; an estimated $^{99m}$Tc-HMPAO CBF was 86.2 mL·100 g$^{-1}$·min$^{-1}$. We measured CBF by using fixed values of E and R that were obtained from the whole brain after an intracarotid bolus injection of the tracer, at a mean CBF level of 59 mL·100 g$^{-1}$·min$^{-1}$. Calculating the permeability surface area product of brain capillaries or the regression between E and $^{99m}$Tc-HMPAO CBF, we can correct low extraction of $^{99m}$Tc-HMPAO CBF SPECT.
rational approach to linearize $^{99m}$Tc-HMPAO CBF is to correct flow-dependent backdiffusion of the tracer by an equation described by Andersen et al.\textsuperscript{11} and Lassen et al.\textsuperscript{9} The HMPAO conversion/clearance ratio ($\alpha$) is different in each individual case, and this assumption is not true in specific diseased regions of the brain. The clinical relevance of the use of correction equations for $E$ and $R$ remains to be clarified. Further studies are necessary to verify the accuracy of the assumptions and to determine the optimal correction method to linearize brain uptake of CBF tracers versus blood-flow relationship.

The accuracy of the estimation of CBF is influenced by multiple sources of variation, such as the difference in $E$, variation in $R$, shape and height of the input function, and errors in the measurement of brain and blood radioactivities. The possibility of unreliable CBF estimates arises from propagation of errors. The whole-brain $^{99m}$Tc-HMPAO CBF value of 35.6±5.3 ml·100 g$^{-1}$·min$^{-1}$ in patients with chronic CVD is in agreement with the values previously reported in the literature.\textsuperscript{21–23} As for the reproducibility of the measurement of CBF, coefficients of variation of the $^{133}$Xe clearance method\textsuperscript{24} and the $^{15}$O inhalation method\textsuperscript{25} are 6.5% and 5%, respectively. Our coefficient of variation of 8.6% is thought to be acceptable for the measurement method of CBF.

Early assessment of patient characteristics that predict outcome after acute ischemic stroke is essential in therapeutic trials and clinical practice.\textsuperscript{26,27} At present, CBF SPECT analysis of the effects of acute stroke therapy with tissue plasminogen activator is under study.\textsuperscript{28} The clinical significance of knowing severity, size, and location of ischemia in CVD has not yet been fully determined.\textsuperscript{29} Some question remains about whether the theory for CBF quantification is true in a particular tissue of the brain, because many brain regions contain a variety of disease and pathological states. Functional tissue heterogeneity (ie, inclusion of tissues with different rates of $E$, $R$, flow, and metabolism within a single ROI) is an unavoidable problem with functional imaging modality. Focal alteration in $E$ and $R$ in pathological tissue may contribute to the error in calculated CBF, but presently it is impossible to separate these effects from global estimates of $E$ and $R$. Calculation of true CBF is essentially difficult in most of CBF tracers as long as $E$ is not complete and backdiffusion exits. Furthermore, the hyperfixation of HMPAO in infarct reperfusion may limit the estimation accuracy.\textsuperscript{30} In patients with CVD, not only the degree of neurological impairment but also age, gender, risk factors, and severity of carotid atherosclerosis can influence CBF.\textsuperscript{31} We have focused on the design and methodology of a simpler, noninvasive method for $^{99m}$Tc-HMPAO CBF quantification. Our method is noninvasive, computationally fast, and effective for measuring CBF in patients with CVD. Future studies are needed to determine whether the use of $^{99m}$Tc-HMPAO-SPECT in the evaluation of CVD promises better differentiation between areas of potentially viable and irreversibly injured tissue than that possible by conventional neuroimaging methods alone.

Acknowledgments

We gratefully acknowledge the help of the staff from the Division of Nuclear Medicine of Osaka National Hospital in acquiring the data. This work was supported by the Research Grant for Cardiovascular Diseases (11–10) from the Ministry of Health and Welfare.

References

Noninvasive Measurement of Cerebral Blood Flow With $^{99m}$Tc-Hexamethylpropyleneamine Oxime Single-Photon Emission Computed Tomography and 1-Point Venous Blood Sampling

Yoshinari Isaka, Satoshi Furukawa, Hideki Etani, Etsuko Nakanishi, Yosuke Ooe and Masatoshi Imaizumi

*Stroke*. 2000;31:2203-2207
doi: 10.1161/01.STR.31.9.2203

*Stroke* is published by the American Heart Association, 7272 Greenville Avenue, Dallas, TX 75231
Copyright © 2000 American Heart Association, Inc. All rights reserved.
Print ISSN: 0039-2499. Online ISSN: 1524-4628

The online version of this article, along with updated information and services, is located on the World Wide Web at:

http://stroke.ahajournals.org/content/31/9/2203

Permissions: Requests for permissions to reproduce figures, tables, or portions of articles originally published in *Stroke* can be obtained via RightsLink, a service of the Copyright Clearance Center, not the Editorial Office. Once the online version of the published article for which permission is being requested is located, click Request Permissions in the middle column of the Web page under Services. Further information about this process is available in the Permissions and Rights Question and Answer document.

Reprints: Information about reprints can be found online at:
http://www.lww.com/reprints

Subscriptions: Information about subscribing to *Stroke* is online at:
http://stroke.ahajournals.org/subscriptions/