Cyclooxygenase-2 Inhibitor NS-398 Protects Neuronal Cultures From Lipopolysaccharide-Induced Neurotoxicity

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Background and Purpose—The prostanoid-synthesizing enzyme cyclooxygenase (COX)-2 is markedly upregulated after cerebral ischemia and may participate in the mechanisms by which postischemic inflammation contributes to the late stages of ischemic brain injury. In the present study, we sought to provide additional evidence for a role of COX-2 in the mechanisms of neurotoxicity associated with inflammation.

Methods—Nine-day-old neuronal-glial cultures, prepared from the cerebral cortex of newborn C57BL/6J mice, were exposed to lipopolysaccharide (LPS), a potent proinflammatory agent. The contribution of COX-2 was investigated by using the COX-2 inhibitor NS-398.

Results—LPS produced a dose-dependent (0.001 to 10 μg/mL) and selective neuronal death that was well developed 72 hours after treatment. The effect was associated with a marked increase in the concentration of the COX reaction product prostaglandin E₂ (PGE₂) and of the cytokine tumor necrosis factor-α (TNF-α). NS-398 (10 μmol/L) blocked the PGE₂ increase, attenuated the TNF-α increase, and prevented the neuronal death produced by LPS. TNF-α–blocking antibodies attenuated LPS-induced neuronal death, but the protection was less pronounced than that afforded by NS-398. LPS failed to elevate PGE₂ or to produce cell death in neuron-enriched cultures, suggesting that glial cells are required for these effects.

Conclusions—COX-2, in part through TNF-α–related mechanisms, contributes to LPS-induced neuronal death. The data support the hypothesis that COX-2, in addition to its role in glutamate excitotoxicity, participates in the cytotoxicity associated with inflammation. (Stroke. 2001;32:2370-2375.)

Key Words: cerebral ischemia ■ cytokines ■ inflammation ■ lipopolysaccharides ■ neurons ■ mice

Prostanoids are potent biological mediators that play an important role in a wide variety of physiological and pathological processes (see review). Cyclooxygenase (COX) is a rate-limiting enzyme for the synthesis of prostanoids (see review). Two COX isoforms have been described: COX-1 and COX-2. COX-1 is ubiquitously expressed and produces prostanoids that are involved in normal cellular functions. COX-2 expression can be induced in several cell types by cytokines, mitogens, bacterial endotoxins, and growth factors (see review). In the normal brain, COX-2 is present in selected neurons, and its expression is regulated by synaptic activity.

There is increasing evidence that COX-2 is involved in the mechanisms of ischemic brain injury. In ischemic stroke, COX-2 is upregulated in the late stages of the injury and is expressed in neurons, glia, inflammatory cells, and blood vessels. Treatment with the COX-2 inhibitor NS-398, starting 6 hours after the induction of ischemia, attenuates cerebral ischemic injury in rodent models of ischemic stroke. In addition, COX-2–null mice have reduced brain injury after focal cerebral ischemia. These data raise the possibility that COX-2 is one of the factors through which the inflammatory reaction that involves the postischemic brain contributes to the delayed progression of ischemic brain injury.

In the present study, we sought to provide further evidence that COX-2 contributes to the mechanisms by which inflammation kills neurons. Using neocortical neuronal-glial cultures, we demonstrate that the endotoxin lipopolysaccharide (LPS) elevates the COX-2 reaction product prostaglandin E₂ (PGE₂) and produces neuronal death. The COX-2 inhibitor NS-398 blocks PGE₂ production and markedly ameliorates LPS-induced neuronal damage. Furthermore, NS-398 attenuates the increase in tumor necrosis factor-α (TNF-α), a cytokine involved in LPS-induced neuronal damage. The data provide new evidence that COX-2, in part through TNF-α, plays a critical role in the brain damage produced by inflammation. Therefore, COX-2 could also be involved in the mechanisms by which inflammation exerts its deleterious effects on the postischemic brain.
Materials and Methods

Materials

Hibernate-A medium, Neurobasal-A medium, B-27 supplement, N-2 supplement, DMEM, diazylated horse serum, heat-inactivated horse serum, Glutamax, and penicillin-streptomycin were obtained from Life Technologies. Papain and DNase1 were purchased from Worthington Biochemical. LPS (Escherichia coli 011:B4) and BSA were from Sigma Chemical Co. NS-398 was obtained from Cayman Chemical. Anti-mouse TNF-α antibodies, neutralizing antibodies to mouse TNF-α, were purchased from R&D Systems. Anti-microtubule-associated protein (MAP)-2 antibodies were purchased from Upstate Biotechnology. Mac-1 antibodies were obtained from Chemicon. Anti–glial fibrillary acidic protein (GFAP) antibodies were from Boehringer-Mannheim. Prediluted goat anti-mouse IgG antibodies were from Biocare. Rabbit anti-rat IgG antibodies were purchased from Vector Laboratories.

Cell Culture

Primary neuron-glial cultures were prepared from postnatal day 0 from C57BL/6J mice by the method of Brewer,14 with some modifications. Briefly, the whole cerebral cortices of 10 mixed in Hibernate-A medium with B-27 supplement (Hibernate-A/B-27). Individual cells were first treated with papain (25 U/mL) and DNase1 (25 U/mL) in Hibernate-A/B-27 for 30 minutes at 30°C. After they were rinsed, the cells were dissociated mechanically in Hibernate-A/B-27). Individual cells were first treated with papain (25 U/mL) and DNase1 (25 U/mL) in Hibernate-A/B-27 for 30 minutes at 30°C. After they were rinsed, the cells were dissociated mechanically in Hibernate-A/B-27 by using a Pasteur pipette. The cell suspension was laid on Hanks’ balanced salt solution containing 4% BSA and centrifuged at 1000 rpm for 5 minutes. The pellet was resuspended in Neurobasal-A medium with B-27 supplement, 0.25 mM glutamate, 1.0 mM nicotinamide, 40 μg/mL streptomycin (Neurobasal-A/B-27), and 10% dialyzed horse serum. Cells were plated on polyornithine-coated 96-well plates or 16-well chamber slides at 5×10⁴ cells per well. Cultures were maintained at 37°C in a humidified atmosphere containing 5% CO₂. Half of the medium was replaced with Neurobasal-A/B-27 and 5% dialyzed horse serum twice a week. After 9 days of plating, the composition of major cell types was estimated by counting cells immunostained with antibodies against cell-type–specific markers. Neurons were stained with an antibody against MAP-2. Astrocytes were stained with an antibody against GFAP. Microglia were stained by using Mac-1 antibodies. Neuron-enriched cultures were prepared by growing dissociated cells in Neurobasal-A/B-27 without serum. The density of plated cells in neuron-enriched cultures was not different from that of the mixed cultures. For glia-enriched cultures, dissociated cells were grown in DMEM containing 10% heat-inactivated horse serum. After 8 days, neurons were killed by exposure to 1 mM/L glutamate, and the medium was changed. Gliarial enrichment was verified by immunostain for GFAP and MAP-2.

Treatment of Cell Cultures

Toxicity experiments were performed 9 days after plating. In all the cultures studied (neuron-enriched, glia-enriched, and mixed cultures), cells were exposed to LPS, alone or in combination with other agents, for 72 hours in Neurobasal-A with N-2 supplement and 1% dialyzed horse serum, unless otherwise stated. NS-398 was dissolved in dimethyl sulfoxide and then diluted in the culture medium. The final concentration of dimethyl sulfoxide was 0.016%. NS-398 (10 μmol/L) was used at the concentration of 3 μmol/L, unless otherwise stated. NS-398 was dissolved in dimethyl sulfoxide and then diluted in the culture medium. The final concentration of dimethyl sulfoxide was 0.016%. NS-398 (10 μmol/L) was added in the medium at times 0 and 48 hours. In some experiments, anti-mouse TNF-α antibodies (10 μg/mL) were added once in the medium at time 0. These concentrations of NS-398 and of TNF-α antibodies were determined in preliminary experiments to provide the greatest effects.

Assessment of Neuronal Death

Neuronal death was examined by the lactate dehydrogenase (LDH) efflux assay.13 Cell culture supernatants were tested for LDH activity by using the CytoTox96 nonradioactive assay (Promega). This assay results in the production of a red formazan product after the conversion of lactate and NAD⁺ to pyruvate and NADH. The color intensity is directly proportional to the number of lysed cells and can be quantified by measuring the absorbance (490 nm). Data are expressed as optical density. The ability of this method to detect neuronal death was validated in our system by comparing the LDH data with neuronal counts in cultures immunostained with the neuronal marker MAP-2 (see below). Neuronal counts were made in 4 random fields (0.25 mm²) at ×200 magnification.

Immunocytochemistry

The cultures were fixed with 4% paraformaldehyde for 30 minutes at room temperature. For Mac-1 staining, cells were treated with 1% Triton X in Tris-buffered saline for 15 minutes. Cells were incubated overnight at 4°C with a primary antibody in antibody dilution buffer (anti–MAP-2 at 1:2000, anti–GFAP at 1:100, and Mac-1 at 1:100; Dako). The cells were then incubated sequentially with a prediluted goat anti-mouse antibody (for anti–MAP-2 and anti–GFAP) and streptavidin-linked horseradish peroxidase (Biocare). Antibody-antibody complexes were visualized by using diaminobenzidine as the chromogen. For Mac-1 staining, cells were incubated with a rabbit anti-rat antibody (1:200) and visualized by using the Vectastain ABC kit (Vector) with diaminobenzidine.

PGE₂,TNF-α, and Nitrite Assays

The concentration of PGE₂ in the culture medium was measured by using an ELISA kit (Cayman Chemical). Levels of TNF-α in the culture medium were determined with an ELISA kit (Biosource), according to the manufacturer’s instructions. Nitrite was measured in the culture medium by using the Griess reaction system (Promega). Absolute concentrations were derived by comparison with a standard curve.

Statistical Analysis

The data are expressed as mean±SE. Multiple comparisons were evaluated by ANOVA followed by the Tukey or Dunnnett test, as indicated. Two-group comparisons were analyzed by the 2-tailed Student t test. For all analyses, a value of P<0.05 was considered significant.

Results

We used 9-day-old neocortical cultures for LPS treatment. At this time, the mixed cultures contained ~48% neurons, 51% astrocytes, and 1% microglia, as estimated by immunostaining with the neuronal marker MAP-2, the astroglial marker GFAP, and the microglial marker Mac-1. Treatment of the cultures with LPS produced a time-dependent degeneration of neurons and an increase in LDH concentration in the supernatant. The effect was most evident 72 hours after LPS when degenerating neurons were most abundant, and LDH levels were significantly higher than those observed in vehicle-treated cultures (Figure 1A). The effect of LPS on LDH concentration was dose dependent (0.001 to 10 μg/mL, Figure 1B).

Treatment of mixed cultures with NS-398 attenuated the LPS-induced LDH efflux (Figure 2A). NS-398 also attenuated the release of PGE₂ produced by LPS treatment (Figure 3A). To confirm that the reduction in LDH reflected a reduction in neuronal death, cultures were immunostained with the neuronal marker MAP-2. In LPS-treated cultures, there was a loss of neuritic network and neuronal perikarya (Figure 4). Although NS-398 itself had no effect in control cultures, it prevented the loss of MAP-2–positive neurons in LPS-treated cultures (Figure 4). Counting of MAP-2–positive neurons showed that NS-398 prevented the loss of neurons in...
LPS-treated cultures (Figure 2B), a finding consistent with the results obtained with the LDH assay.

To examine the cell type responsible for the toxicity induced by LPS, the effects of LPS were studied in neuron-enriched cultures. In these cultures, LPS did not increase PGE2 (Figure 3A) and failed to produce neurotoxicity, as assessed morphologically or by LDH efflux (Figure 3B). We also studied the effect of LPS in glia-enriched cultures. In such cultures, LPS did not produce glial degeneration, as assessed by morphological criteria, although a small increase in LDH was observed (optical density at 72 hours, 0.05 ± 0.01 for control and 0.08 ± 0.01 for LPS; P < 0.05; n = 6 per group). The lack of nitrite production by LPS stimulation may reflect the fact that LPS alone is not sufficient to induce expression of inducible NO synthase in mouse neuronal-glial cultures (eg, see Jeohn et al17).

There is evidence that NO enhances COX-2 activity and that such activation contributes to COX-2–induced neurotoxicity.12,16 To determine whether NO participates in the mechanisms of the neurotoxicity associated with inflammation,18 therefore, we sought to determine whether LPS treatment increases TNF-α and, if so, whether NS-398 attenuates such TNF-α elevation. In control culture, TNF-α concentration was below the detection limit of the assay (19.5 pg/mL). LPS produced a marked increase in TNF-α concentration that was attenuated by NS-398 (see Figure 4 for MAP-2 immunocytochemistry). Comparison with LDH data in panel A indicates that a ~3-fold LDH increase corresponds to ~50% neuronal death. *P < 0.05.

Figure 1. Effect of LPS (10 μg/mL) or vehicle on LDH release in the supernatant of mixed neuronal-glial cultures. A, Time course of LDH release in cultures treated with vehicle or LPS. OD490 indicates optical density at 490 nm. *P < 0.05 (ANOVA and Tukey test). B, Relationship between LPS concentration and LDH release 72 hours after LPS treatment. LDH data in this and other figures are expressed as mean ± SE of 4 wells from 3 independent experiments. *P < 0.05 compared with vehicle-treated cultures (ANOVA and Dunnett test).

Figure 2. Effect of NS-398 on LPS-induced LDH release and neuronal counts. A, Cultures were incubated for 72 hours with medium alone (vehicle), vehicle plus NS-398, LPS alone, or LPS plus NS-398. *P < 0.05 (ANOVA and Tukey test). The LPS+NS-398 group is not statistically different from the vehicle and vehicle+NS-398 groups (P > 0.05). B, Quantification of the effect of NS-398 on MAP-2–immunostained cultures exposed to LPS is shown (see Figure 4 for MAP-2 immunocytochemistry). Comparison with LDH data in panel A indicates that a ~3-fold LDH increase corresponds to ~50% neuronal death. *P < 0.05.
by treatment with NS-398 (Figure 5A). Treatment with anti–TNF-α antibodies attenuated the LPS-induced increase in LDH (Figure 5B). However, the attenuation in LDH produced by anti–TNF-α antibodies was less pronounced than that observed with NS-398 (Figure 5B).

**Discussion**

There is increasing evidence that COX-2 is involved in the pathogenesis of several neurological disorders associated with inflammation, including ischemic brain injury. Cerebral ischemia is associated with a marked inflammatory reaction that contributes to the progression of the damage. In models of cerebral ischemia, as well as in human stroke, COX-2 is upregulated in neurons, inflammatory cells, glial cells, and the vasculature. Inhibition of COX-2 with NS-398 or genetic deletion of COX-2 in null mice attenuates cerebral ischemic damage, suggesting that COX-2 reaction products participate in ischemic injury. In systemic models of inflammation, COX-2 is well known to contribute to the associated tissue damage. However, the role of COX-2 in the neuronal damage produced by inflammation in the brain has not been extensively investigated.

In the present study, we sought to examine further the role of COX-2 in the mechanisms of neurotoxicity produced by inflammatory stimuli. To achieve this goal, we treated neocortical neuronal-glial cultures with the endotoxin LPS. LPS produced neuronal death that was well developed 2 or 3 days after LPS exposure and was associated with a marked elevation of the COX reaction product PGE2 but not of the NO metabolite nitrite. Treatment of the cultures with the COX-2 inhibitor NS-398 blocked the increase in PGE2 and prevented neuronal death. These data suggest that COX-2 is involved in the mechanisms of the delayed cell death produced by LPS.

NS-398 was used as a relatively selective COX-2 inhibitor. Although NS-398 can activate peroxisome proliferator–activated receptors, which are involved in inflammation, such an effect is observed at concentrations of 100 to 300 μmol/L. Therefore, at the relatively low concentration used in the present study (10 μmol/L), it is unlikely that NS-398 exerts its protective effect through the activation of peroxisome proliferator–activated receptors. Furthermore, NS-398 does not protect against NMDA-induced cytotoxicity in COX-2–null mice, demonstrating that COX-2–independent mechanisms do not participate in its neuroprotective effects. Experiments using brain cultures from COX-2–null mice would provide further evidence for a role of COX-2 in LPS-induced cell death.

To gain insight into the cellular sources of COX-2, cultures enriched with neurons were studied. In such cultures, LPS failed to produce PGE2 elevation or elicit neuronal death. This observation is consistent with the hypothesis that glial cells are the source of PGE2 and, presumably, COX-2 in our...
exacerbate neurotoxicity by facilitating astrocytic glutamate release. However, it remains to be established whether glutamate plays a role in the neuronal death induced by LPS. Furthermore, other studies have reported that prostanoids protect against glutamate neurotoxicity. Therefore, it remains unclear whether COX-2-derived prostanoids are involved in LPS toxicity through glutamate-dependent mechanisms. Experiments in which LPS-stimulated cultures are treated with glutamate receptor inhibitors would be required to address this issue. Furthermore, the role of arachidonic acid metabolites produced by the 5-lipoxygenase pathway in the mechanisms of LPS-induced cell death remains to be defined.

It is unlikely that the protective effect of COX-2 inhibition is entirely mediated through TNF-α signaling, because the protection exerted by anti–TNF-α antibodies was less than that observed with NS-398. Thus, the beneficial effects of NS-398 are also mediated by mechanisms independent of TNF-α. Furthermore, the fact that TNF-α–blocking antibodies did not completely abolish LPS-induced neurotoxicity suggests that in our model, as in other cell culture systems, TNF-α is not the sole mediator of neurotoxicity. As for the cellular origin of TNF-α, glial cells are a likely source of this cytokine, although a neuronal source cannot be ruled out. Recent data suggest that COX-2 is also involved in pathogenic events occurring in the early stages of cerebral ischemia. COX-2 inhibition by NS-398 protects neuronal cultures from glutamate neurotoxicity and reduces the brain damage produced by direct injection of N-methyl-D-aspartate into the mouse cerebral cortex. Furthermore, the damage produced by N-methyl-D-aspartate microinjection is attenuated in COX-2-null mice. These observations suggest that COX-2 reaction products participate in the mechanisms of glutamate neurotoxicity, a process that occurs in the early stages of cerebral ischemia. It is of interest that in glutamate excitotoxicity, at variance with LPS-induced cytotoxicity, the cellular source of COX-2 is neuronal. COX-2 inhibition is an attractive therapeutic strategy for ischemic stroke because it can counteract pathogenic mechanisms taking place in both the early and late stages of cerebral ischemia.

In conclusion, we have demonstrated that the COX-2 inhibitor NS-398 attenuates the neurotoxicity produced by the proinflammatory agent LPS. The protective effect is associated with a reduction in the COX-2 reaction product PGE₂ and the cytokine TNF-α. The data indicate that COX-2 is involved in the mechanisms of neurotoxicity associated with inflammation, and the data are consistent with the hypothesis that COX-2 contributes to the neuronal damage resulting from the inflammatory reaction triggered by cerebral ischemia.

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References


33. Araki et al. COX-2 and Neuronal Damage 2375
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