Neuroprotection and P450 2C11 Upregulation After Experimental Transient Ischemic Attack

Nabil J. Alkayed, MD, PhD; Toru Goyagi, MD; Hung-Dong Joh, MS; Judith Klaus, RN; David R. Harder, PhD; Richard J. Traystman, PhD; Patricia D. Hurn, PhD

Background and Purpose—Transient ischemic attack (TIA) is a risk factor for stroke. However, recent clinical evidence suggests that TIA may improve stroke outcome. We tested the hypothesis that experimental TIA induces expression of P450 2C11, an arachidonic acid epoxygenase that produces vasodilator epoxyeicosatrienoic acids, leading to increased tissue perfusion and reduced stroke damage.

Methods—Wistar rats underwent three 10-minute middle cerebral artery occlusions (TIA) or sham surgery. Three days later, animals were subjected to 2-hour middle cerebral artery occlusion and 24 hours of reperfusion. Brains were stained with 2,3,5-triphenyltetrazolium chloride for infarct size measurement or processed for quantification of P450 2C11 mRNA and protein with the use of RNase protection assay and Western blotting. Regional cerebral blood flow (CBF) at the end of 2-hour ischemia was measured in separate groups of rats with iodoantipyrine autoradiography.

Results—Cerebral infarct was reduced by >50% in TIA- versus sham-preconditioned animals. 2C11 mRNA and protein were increased in ipsilateral hemisphere by 3 days after TIA but not sham surgery. Induction of 2C11 by TIA was also evident in ipsilateral hemisphere at 24 hours after 2-hour middle cerebral artery occlusion and 24 hours of reperfusion. End-ischemic regional CBF was not different between TIA- and sham-pretreated groups.

Conclusions—We conclude that experimental TIA induces ischemic tolerance by a mechanism temporally linked to upregulation of P450 2C11. Enzyme induction does not attenuate ischemic severity by amplifying end-ischemic CBF. (Stroke. 2002;33:1677-1684.)

Key Words: cerebral ischemia, transient ■ gene expression ■ neuroprotection ■ stroke, experimental ■ rats
(EETs), which exert a variety of biological functions in brain, including cerebral vasodilatation\textsuperscript{17,18} and maintenance of CBF.\textsuperscript{19} We reasoned that if TIA induces 2C11 expression, then TIA-preconditioned brain may have enhanced ability to generate EETs from AA normally released during cerebral ischemia, which may be advantageous during subsequent stroke. In heart, EETs production is increased during myocardial ischemia,\textsuperscript{20} which is protective against ischemia/reperfusion injury,\textsuperscript{21} likely via enhanced endothelium-dependent coronary artery relaxation.\textsuperscript{22} Accordingly, we tested the hypothesis that 2C11 induction in TIA-preconditioned brain enhances intraschismic vasodilator capacity and CBF and consequently reduces tissue damage during subsequent stroke.

### Materials and Methods

The study was conducted in accordance with the National Institutes of Health guidelines for care and use of animals in research, and the protocols were approved by the institutional Animal Care and Use Committee.

#### Experimental Groups

A total of 95 adult male Wistar rats (aged 13 to 15 weeks) were subjected to 1 of 3 prestroke treatments: (1) three 10-minute middle cerebral artery (MCA) occlusions, separated by 45 minutes of reperfusion (TIA procedure); (2) sham operation (sham procedure); or (3) no surgical manipulation or anesthesia (control). Animals were then allowed to survive for 1, 2, or 3 days. On day 3, some animals were additionally subjected to 2 hours of MCA occlusion (stroke procedure) and allowed to survive for 24 hours. The following end points were measured: (1) infarct size was measured by 2,3,5-triphenyltetrazolium chloride (TTC) at 24 hours after stroke procedure in animals pretreated with TIA procedure, sham procedure, or no procedure (n=6 per group); (2) 2C11 mRNA was determined by RNase protection assay (RPA) at days 1, 2, and 3 after TIA or sham procedure (samples were pooled from 3 to 4 animals per group at each time point) and at 24 hours after stroke procedure in sham- (n=3) and TIA-pretreated (n=7) rats; (3) 2C11 protein was determined by Western blot analysis on days 1, 2, and 3 after TIA or sham procedure (n=4 to 6 animals per group at each time point) and at 24 hours after stroke procedure in sham- and TIA-preconditioned groups (n=4 to 5 animals per group); and (4) regional CBF was determined by \([\text{[14C]iodoantipyrine (IAP)}\) autoradiography at the end of 2-hour stroke procedure in TIA- versus sham-pretreated groups (n=4 per group).

#### Experimental TIA

Transient focal cerebral ischemia was achieved by modifying a previously described technique for intraluminal MCA occlusion.\textsuperscript{23,24} Briefly, rats were anesthetized with halothane and instrumented with a femoral artery catheter and a small probe for monitoring laser-Doppler perfusion (LDP) over ipsilateral parietal cortex.\textsuperscript{23} A 4-0 nylon surgical suture was inserted through the internal carotid artery until LDP dropped. The filament was then fixed in place and was withdrawn after 2 hours to reexpose, and an occluding filament was inserted through a small incision into the internal carotid artery until LDP dropped. The filament was then fixed in place and was withdrawn after 2 hours to initiate reperfusion. After 22 hours of reperfusion, the rat was decapitated under deep halothane, and the brain was removed and sliced into seven 2-mm-thick coronal sections. LDP was digitally measured in slices and integrated across the hemisphere to calculate infarct volume.

#### RNase Protection Assay

P450 2C11 mRNA was measured with the use of both indirect and direct RPA. Indirect RPA was performed on RNA samples isolated from whole hemispheres at 22 hours of reperfusion (poststroke expression), as previously described.\textsuperscript{25} Direct RPA was performed on tissue samples microdissected from 8 distinct brain regions (dorsomedial and dorsolateral somatosensory cortex and dorsolateral and ventromedial striatum of ipsilateral and contralateral hemispheres), as previously described.\textsuperscript{26} A rat 2C11 cDNA fragment (296 bp long, between 873 and 1169 bp of 2C11 cDNA sequence)\textsuperscript{14} was amplified by polymerase chain reaction with the use of the following primers: 5'-GAAA GCTT GGTG GCTA CTG-3' (sense) and 5'-CTTG GGAA TGAA GTA-3' (antisense). The T7 phage polymerase promoter sequence (5'-TAAT ACGA CTCA CTAT AGGG-3') was added to the 5' end of the antisense primer to allow for in vitro transcription of 2C11 antisense RNA from the polymerase chain reaction product. Radiolabeled 2C11 antisense riboprobe was transcribed to a specific activity of approximately 10\textsuperscript{6} cpm/\mu g. Total RNA for indirect RPA was prepared separately from whole contralateral hemispheres of individual animals with the use of TRizol (Gibco BRL). Micropunches representing 1 region were pooled from 3 to 4 animals, homogenized in lysis solution for direct RPA (Ambion), and incubated with excess bcl-2 and \(\beta\)-actin probes overnight. Samples were digested with RNase and resolved via electrophoresis on a denaturing polyacrylamide gel. Protected fragments were visualized on a Phosphorimager, and bands were quantified with the use of ImageQuant software (Molecular Dynamics). Optical density of 2C11 and \(\beta\)-actin bands was sampled with the use of a rectangle with predetermined dimensions based on the band with highest intensity. Mean 2C11/\(\beta\)-actin ratio of sham animals was considered 100%, and ratios of all other animals were expressed as a percentage of sham.

#### Western Blotting

Microsomal proteins were prepared from whole hemispheres or from middle coronal slices (at the level of caudate between 0 and −2 mm relative to bregma), as previously described.\textsuperscript{14,15} Brain tissue was homogenized in 250 mM/L sucrose, 10 mM/L KPO\textsubscript{4}, 1 mM/L EDTA, and 0.1 mM/L phenylmethylsulfonyl fluoride (pH 7.7) in the presence of protease inhibitors (2 \mu g/mL aprotonin, 2 \mu g/mL leupeptin, 1 \mu g/mL pepstatin), and the pellet was resuspended in 100 mM/L KPO\textsubscript{4}, 1 mM/L EDTA, 1 mM/L dithiothreitol, and 30% glycerol (pH 7.25). The homogenate was centrifuged at 10 000 \(\times\) g for 15 minutes at 4°C, and the supernatant was further centrifuged at 100 000 \(\times\) g for 60 minutes at 4°C. The protein concentration was measured by BCA protein assay (Pierce). Samples of 100 \mu g protein were then boiled in Laemmli buffer for 5 minutes, electrophoresed on 10% sodium dodecyl sulfate–polyacrylamide gels, and blotted onto polyvinylidene fluoride membrane (Bio-Rad). Blots were blocked with 5% nonfat dry milk in 0.05 mol/L Tris-buffered saline/0.05% Tween for 2 hours at room temperature, then incubated with rabbit anti-rat 2C11 polyclonal antibody,\textsuperscript{14} (1:2500, overnight at 4°C), followed by incubation with biotinylated goat anti-rabbit IgG...
Physiological Variables at Baseline and During Ischemia in Control and Sham- and TIA-Preconditioned Animals

<table>
<thead>
<tr>
<th>Group</th>
<th>MAP, mm Hg</th>
<th>pH</th>
<th>$P_{aO_2}$, mm Hg</th>
<th>$P_{aCO_2}$, mm Hg</th>
<th>Hemoglobin, g/100 mL</th>
<th>Glucose, g/100 mL</th>
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<td>TIA</td>
<td>Baseline</td>
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<td>141±08</td>
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<td></td>
<td>Ischemia</td>
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<td>47±4</td>
<td>133±07</td>
<td>10.5±0.4</td>
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<td>Sham</td>
<td>Baseline</td>
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<td>7.36±0.01</td>
<td>49±4</td>
<td>145±03</td>
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<tr>
<td></td>
<td>Ischemia</td>
<td>87±4</td>
<td>7.37±0.01</td>
<td>48±2</td>
<td>136±08</td>
<td>11.0±0.3</td>
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<tr>
<td>Control</td>
<td>Baseline</td>
<td>86±1</td>
<td>7.37±0.01</td>
<td>46±1</td>
<td>130±05</td>
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<tr>
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<td>Ischemia</td>
<td>87±2</td>
<td>7.37±0.00</td>
<td>48±1</td>
<td>140±05</td>
<td>11.0±0.5</td>
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MAP indicates mean arterial pressure.

Effect of TIA on Infarct Size

Cortical and striatal infarcts were reduced by 50% to 60% in TIA-pretreated compared with sham-pretreated rats (Figure 2; $P<0.05$). There were no significant differences in infarct size between groups. LDP was reduced to 12% of baseline during sham surgery. *Statistically significant difference from preocclusion baseline ($P<0.05$).

Results

Physiological variables measured before and during 2-hour stroke did not differ among groups (Table). Figure 1 traces LDP changes during TIA or sham surgery. LDP was reduced by 70% during TIA but remained within 97±12% of baseline during sham surgery. During 2-hour stroke procedure, LDP was equivalently reduced in sham- and TIA-pretreated rats to 34±5% and 34±4% and returned on reperfusion to 109±6% and 92±10% of baseline, respectively.
was 2-fold in ipsilateral hemisphere of TIA-compared with
eral hemispheres of sham- and TIA-preconditioned brains.

Effect of TIA on P450 2C11 Expression
The time course of 2C11 mRNA induction after TIA in 4egions within the MCA territory is summarized in Figure 3.
Induction of 2C11 mRNA was apparent by day 2 in TIA
versus sham groups in medial cortex and throughout striatum.
By day 3, mRNA levels were 2 to 3 times higher in ipsilateral
cortex and striatum in TIA versus sham groups. Contralateral
expression was unchanged by TIA at all time points (data not
shown). Increased levels of 2C11 mRNA in TIA versus sham
animals persisted after the stroke procedure. Figure 4 dem-
strates representative RPA blots illustrating poststroke
2C11 and β-actin RNA expression in ischemic and contralat-
eral hemispheres of sham- and TIA-preconditioned brains.
The level of 2C11 mRNA at 24 hours after stroke procedure
was 2-fold in ipsilateral hemisphere of TIA- compared with
sham-pretreated animals (244±45% versus 100±24%;
P=0.049). Expression of 2C11 mRNA in contralateral hemi-
sphere did not differ between the 2 groups (197±69% in TIA
versus 100±44% in sham animals; P=0.342).

Figure 5 depicts representative Western blots demonstrat-
ing 2C11 protein expression within ischemic hemisphere
after TIA or sham surgery. P450 2C11 is a 55-kDa protein
that is expressed in male but not female liver.14 However, as
seen in Figure 5A, in brain, 2C11 protein is expressed in both
males and females. The level of 2C11 protein in ipsilateral
hemisphere on days 1 and 2 after TIA was not different than
in sham animals and averaged 82±7% and 119±17% of
sham level, respectively. In agreement with RPA results, the
level of 2C11 protein in ipsilateral hemisphere on day 3 was
2-fold higher in TIA versus sham animals (Figure 5B).
Furthermore, 2C11 protein expression 24 hours after stroke
procedure was 2-fold higher in TIA versus sham groups
(Figure 6). No differences in 2C11 protein expression after
stroke were observed between the 2 groups in the contralat-
eral hemisphere.

Effect of TIA on Regional CBF
End-ischemic regional CBF was not different between TIA-
and sham-pretreated animals. Mean CBF rates at the end
of 2-hour ischemia were 53±13 and 41±9 mL/100 g per minute
in the cerebral cortex and striatum of TIA-pretreated rats
compared with 62±13 and 33±7 mL/100 g per minute in
sham animals. No differences in CBF rates were observed
in the contralateral hemisphere between the 2 groups. Furth-
more, cumulative distribution analysis of CBF rates within
ischemic hemisphere revealed no differences in the amount of
tissue (mm3) perfused with any given flow rate (mL/100 g per
minute) between TIA- and sham-pretreated groups (Figure 7).

Figure 2. Cortical and striatal infarcts at 24 hours after 2-hour
MCA occlusion in rats subjected 3 days earlier to TIA or sham
operation. Cortical and striatal infarcts were reduced by 50% to
60% in TIA vs sham animals. There were no differences in
infarct size between control and sham groups. *Statistically sig-
ificant difference from sham (P<0.05). Ipsilat indicates
ipsilateral.

Figure 3. Regional and temporal expression of P450 2C11
mRNA after experimental TIA. P450 2C11 mRNA was quanti-
ded in dorsomedial and dorsolateral cerebral cortex and ventrome-
dial and dorsolateral striatum on days 1, 2, and 3 after TIA by
direct RNase protection assay. Levels of P450 2C11 were 2 to 3
times higher on day 3 after TIA in ipsilateral cortex and striatum
in TIA vs sham animals. Contralateral expression was
unchanged by TIA at all time points. Each value represents 3 to
4 pooled samples.

Figure 4. Representative poststroke 2C11 and β-actin mRNA
blots in ischemic (I) and contralateral (C) hemispheres of sham-
and TIA-preconditioned brains. 2C11 mRNA was quantified at
24 hours after 2-hour MCA occlusion with the use of indirect
RNase protection assay and normalized to β-actin. 2C11 mRNA
after stroke was 2-fold higher in ischemic hemisphere of TIA- vs
sham-preconditioned animals. No difference in 2C11 mRNA was
observed between the 2 groups in contralateral hemisphere.

Figure 5. Representative 2C11 Western blots after TIA or sham
surgery. A, Antibody specificity. 2C11 protein (molecular weight,
55kDa) is expressed in male (+ctrl) but not female (−ctrl) liver.
In contrast, both male (MB) and female (FB) rat brain express
2C11 at a comparable level. B, 2C11 protein in ipsilateral hemi-
sphere on days 1, 2, and 3 after TIA or sham surgery. The level
of 2C11 protein in ipsilateral hemisphere on day 3 was 2-fold
higher in TIA vs sham group. No difference was observed in
contralateral hemispheres at any time point.
Discussion

We demonstrated that prior exposure to TIA decreases infarct size in experimental stroke. The protection coincided with upregulation of brain P450 2C11 AA epoxygenase, which does not alter severity of the ischemic insult. These findings suggest a protective role for EETs, the products of AA epoxygenase, against cerebral ischemia.

We found that three 10-minute periods of MCA occlusion provide protection against lethal ischemia consisting of 2 hours of MCA occlusion within 3 days, which is consistent with previous reports of preconditioning in focal cerebral ischemia.\(^8\)\(^{-11}\) In our study this preconditioning paradigm does not cause tissue injury as identified by TTC. However, in previous studies,\(^8\) more subtle signs of tissue injury, such as selective neuronal necrosis, were identified at various time points after TIA.

The mechanism of TIA-acquired ischemic tolerance in brain is unknown. Acute preconditioning results from rapid posttranslational modifications of existing proteins, while delayed tolerance involves gene induction and de novo protein synthesis. One highly inducible group of proteins involved in cellular adaptation to stress is that of the cytochrome P450 enzymes. During cerebral ischemia, AA is released from membranes, which can then be metabolized via 3 enzymatic pathways: cyclooxygenase, lipoxygenase, or cytochrome P450 pathways. We previously showed that rat brain astrocytes metabolize AA via the P450 pathway to

Figure 6. P450 2C11 protein expression after stroke. 2C11 protein was quantified in ischemic (ISCHEM) and contralateral (CONTRALAT) hemispheres at 24 hours after 2-hour MCA occlusion by Western blotting. Protein samples were prepared from 2-mm-thick coronal slices at the level of the caudate of brains pretreated 3 days earlier with either TIA or sham surgery. The level of 2C11 protein after stroke was 2-fold higher in ischemic hemisphere in TIA-preconditioned brain compared with sham. No difference was observed in contralateral hemispheres. *Statistically significant difference from sham.

Figure 7. Cumulative distribution of CBF rates within the ischemic hemisphere at end of 2-hour MCA occlusion in TIA- versus sham-pretreated groups. Flow rates were measured by quantitative autoradiography with [\(^{14}\)C]IAP, and analysis was performed at 3 coronal levels represented by the color images. No differences in the amount of tissue (mm\(^3\)) perfused with any given flow rate (ml/100 g per minute) were observed between TIA and sham-pretreated groups.
EETs, which play an important role in coupling neuronal activity to regional blood flow. We have also demonstrated that astrocytes express cytochrome P450 2C11, a P450 isoform with AA epoxygenase activity, which is inducible by glutamate and is an important contributor to the epoxygenase activity of brain. Recent reports suggested that EETs are protective against myocardial ischemia. Thus, we examined whether TIA induces expression of P450 2C11 in brain and whether the temporal profile of 2C11 induction in brain coincides with neuroprotection observed in this model.

We measured 2C11 mRNA and protein on days 1, 2, and 3 after TIA or sham operation and at 24 hours after stroke. We found that the level of 2C11 mRNA in TIA-pretreated animals on day 1 was less than that in sham-operated animals, especially in the striatum. P450 2C11 mRNA expression is known to be suppressed by inflammatory mediators. Thus, the relative attenuation of 2C11 mRNA expression on day 1 may reflect a more severe insult and inflammatory response on day 1 after TIA. By day 2 after TIA, 2C11 mRNA recovers to equal or exceed its level of expression in sham-operated animals. By day 3, 2C11 mRNA levels were 2 to 3 times higher in TIA- versus sham-preconditioned animals. In agreement with the changes observed at the mRNA level, 2C11 protein was 2-fold higher in preconditioned compared with sham animals in ischemic, but not contralateral, hemisphere.

The observed upregulation of 2C11 beginning on day 2 is in agreement with the finding of Chen et al. that a similar preconditioning paradigm confers protection against MCA occlusion on days 2 and 3 but not on day 1. We also measured 2C11 expression 24 hours after stroke onset in TIA-pretreated and sham animals. Measurements performed on total RNA and tissue homogenates demonstrated that the 2-fold increase in 2C11 mRNA and protein expression in ipsilateral hemisphere was maintained during the immediate posts ischemic period critical to infarct maturation. The 2-fold upregulation of 2C11 at the mRNA and protein levels at this time point is important because this is when infarct was measured and protection was observed. These findings demonstrate that 2C11 upregulation precedes and correlates spatially and temporally with tissue protection from stroke but does not prove that 2C11 upregulation and neuroprotection are mechanistically linked.

We then examined whether TIA preconditioning decreases infarct size by attenuating severity of the ischemic insult. Our previous work has identified EETs as endothelium-dependent hyperpolarizing factors and mediators of functional hyperemia in brain. Furthermore, EETs are vasodilators in the cerebral circulation and promote capillary angiogenesis. Therefore, we tested the hypothesis that 2C11 upregulation by TIA is associated with higher tissue perfusion during ischemia, attenuating ischemic severity, and accounting for the observed reduction in infarct size. Measurement of LDP on MCA occlusion on day 3 demonstrated that the relative change from baseline in LDP was similar between the 2 groups. However, because the relationship between LDP and true blood flow is unknown and because LDP measures only surface tissue perfusion, we also quantified absolute blood flow rates within MCA territory using quantitative IAP autoradiography. Measurement of regional CBF by this technique at end ischemia indicated that ischemic severity was equivalent between the sham and TIA groups despite the striking difference in infarct size, suggesting that ischemic tolerance in our model is not mediated via a blood flow–enhancing mechanism acting during vascular occlusion. This is in agreement with most studies demonstrating no difference in CBF within protected regions before or during stroke between preconditioned and sham-operated animals. In contrast, Matsushima and Hakim observed higher CBF during stroke in preconditioned versus sham animals; however, the increase in CBF in their study was not associated with a reduction in infarct size. In summary, evidence from our study with the use of LDP and autoradiography corroborates studies by others to support the conclusion that TIA preconditioning does not alter the severity of ischemia. Our study did not address the question of whether TIA preconditioning improves regional CBF during the reperfusion period.

In addition to vasodilation, EETs have other properties that can potentially mediate their neuroprotective effect. These include the antioxidant, anti-inflammatory, and antipyretic properties of EETs. Furthermore, EETs regulate intracellular calcium concentration, inhibit platelet aggregation, and prevent leukocyte adhesion to vascular wall. A related P450 metabolite, 16(R)-hydroxyecosatetraenoic acid [16(R)-HETE], suppresses human leukocyte activation and reduces intracranial pressure in a rabbit model of thromboembolic stroke. These mechanisms may be important factors in how upregulation of 2C11 reduces stroke damage.

Several other mechanisms have been proposed to mediate the protection associated with late-phase preconditioning in brain. Tolerance to ischemia is associated with higher expression of neuroprotective genes such as heat shock proteins, superoxide dismutase, and bel-2. Ischemic tolerance has also been induced by the cytokines interleukin-1 and tumor necrosis factor–α (TNF-α) and has been linked to greater ability to buffer cytosolic Ca2+ loads, enhanced ceramide synthesis, and higher capacity to neutralize reactive oxygen species. Several signaling pathways have been shown to be activated by ischemic preconditioning, including the N-methyl-D-aspartate/nitric oxide/p21(ras)/extracellular regulated kinase, the adenosine A1 receptor/protein kinase C(AMP-dependent potassium channel, and the Janus tyrosine kinase(signal transducers and activators of transcription signaling pathways. It is possible that EETs may exert their protective effect via some of these mechanisms. For example, EETs may induce bel-2 or other cAMP-responsive genes with neuroprotective properties via their action on cAMP. EETs may also exert neurotrophic effects via their mitogenic actions, such as their role in mediating epidermal growth factor signaling. Furthermore, EETs may mediate the preconditioning effects by regulating intracellular calcium concentrations. EETs also antagonize TNF-α effects, and both TNF-α and interleukin-1β downregulate 2C11 expression, likely via the sphingomyelin/ceramide pathway. Finally, EETs activate ATP-sensitive K+ channels in rat cardiac ventricular myocytes.

The clinical implication of our findings is limited by the fact that these studies were conducted in young healthy...
animals and that they measured only short-term outcome. Because stroke and TIA mostly affect the aging population, it is important to determine whether experimental TIA remains effective in eliciting protection in aged animals with underlying vascular disease and whether the protection is long lasting. In heart, some studies demonstrated loss of preconditioning effect with aging, while other studies demonstrated that preconditioning is well preserved in senescent myocardium.

One explanation for the discrepancy is that studies that showed loss of preconditioning effect with aging have mostly been conducted on isolated buffer-perfused rat heart. Furthermore, these studies demonstrated loss of early-phase preconditioning, which is not surprising given age-related changes in the vasculature. It is yet to be determined whether late-phase preconditioning in heart or brain is lost with advanced age. In brain, global ischemic preconditioning was more effective in aged versus young gerbils. In elderly humans, antecedent (preinfarction) angina, which presumably is a preconditioning stimulus, has been reported to exert a beneficial effect on outcomes from myocardial infarction in some but not other studies. Finally, diabetes mellitus in humans and hyperglycemia in animals prevented ischemic preconditioning in heart. Interestingly, diabetes also suppresses 2C11 expression, but it is not clear yet whether 2C11 suppression by chronic hyperglycemia alters the preconditioning effect.

In summary, we described a model of experimental TIA and demonstrated its effect to reduce subsequent stroke damage. The protection against stroke was temporally and spatially linked to upregulation of P450 2C11 epoxygenase without affecting ischemic stroke severity. These results suggest that EETs may serve as endogenous neuroprotectants in brain, which are induced by TIA and can be targeted as a novel strategy for ischemic brain injury prevention and treatment. EET levels in brain can be enhanced by increasing their synthesis or by decreasing their breakdown. EETs are inactivated by hydration into dihydroeicosatrienoic acids by the epoxide hydrolase.

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### References

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