Mild Postischemic Hypothermia Prolongs the Time Window for Gene Therapy by Inhibiting Cytochrome c Release

Heng Zhao, PhD; Midori A. Yenari, MD; Robert M. Sapolsky, PhD; Gary K. Steinberg, MD, PhD

Background and Purpose—We showed previously that Bcl-2 overexpression with the use of herpes simplex viral (HSV) vectors improved striatal neuron survival when delivered 1.5 hours after stroke but not when delivered 5 hours after stroke onset. Here we determine whether hypothermia prolongs the therapeutic window for gene therapy.

Methods—Rats were subjected to focal ischemia for 1 hour. Hypothermia (33°C) was induced 2 hours after insult and maintained for 3 hours. Five hours after ischemia onset, HSV vectors expressing Bcl-2 plus β-gal or β-gal alone were injected into each striatum. Rats were killed 2 days later.

Results—Striatal neuron survival of Bcl-2–treated, hypothermic animals was improved 2- to 3-fold over control-treated, hypothermic animals and Bcl-2–treated, normothermic animals. Neuron survival among normothermic, Bcl-2–treated animals was not different from control normothermics or control hypothermics. Double immunostaining of cytochrome c and β-gal demonstrated that Bcl-2 plus hypothermia significantly reduced cytochrome c release.

Conclusions—Postischemic mild hypothermia extended the time window for gene therapy neuroprotection using Bcl-2 and reduced cytochrome c release. (Stroke. 2004;35:572-577.)

Key Words: cerebral ischemia • gene therapy • hypothermia • ischemia, focal • proto-oncogene proteins c-bcl-2

Materials and Methods

Generation of Plasmids
HSV vectors were generated as previously described. The amplicon plasmid pα22bβgal4bcl2-1 contained the human bcl-2 gene and the Escherichia coli lacZ gene under the control of the HSV α4 and α22 promoters, respectively. A control vector, designated α4b, which contains lacZ gene alone, was also generated. Amounts of infectious vector particles in each injection were as follows: Bcl-2 vector, 4.6 to 6.3×10⁶; control vector, 5.8×10⁵. Vector:helper virus ratios ranged from 1:4 to 1:5 for Bcl-2 and 1:5 for control.

Surgery
The Stanford University Administrative Panel on Laboratory Animal Care approved all animal procedures. We based the design of the current experiment on our previous time course study in which Bcl-2 overexpression improved neuronal survival when delivered 1.5 hours but not 5 hours after stroke. Figure 1 presents a diagram of the surgical procedure. Sprague-Dawley rats (weight, 290 to 350 g) were anesthetized with 5% isoflurane, then isoflurane was decreased to 1.5% to 3.0% for the remainder of the procedures. Rats were placed on a heating/cooling blanket to maintain rectal temperature between 37°C and 38°C. There were 4 groups: (1) normothermic rats given control vector (n=11); (2) normothermic rats given Bcl-2 vector (n=11); (3) hypothermic rats given control vector (n=11); and (4) hypothermic rats given Bcl-2 vector (n=10). The left middle cerebral artery (MCA) was occluded by inserting an intraluminal 3-0 nylon monofilament suture through the common carotid artery to the branch point of the MCA. After 1 hour of ischemia, the suture was withdrawn. Normothermic rats were allowed to recover for 3.5 hours and then were reanesthetized. Hypothermic animals were allowed to

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PaCO₂ and PaO₂ were measured (by methods described previously) in 2 groups of animals (n=3 per group) that were treated exactly the same as the rats in the normothermic and hypothermic groups, except that no vector was delivered. Additionally, to verify a close correlation between brain and rectal temperature in these animals, a small burr hole was drilled to permit insertion of a 33-gauge thermocouple temperature probe to measure brain temperature in the nonischemic hemisphere. The probe was inserted 4.5 mm into the striatum.

Bcl-2 or control vectors were injected bilaterally into the striata 5 hours after ischemia onset. Coordinates for the 2.5-μL injections were as follows (from bregma): anteroposterior=0, mediolateral=3.5 mm, with 2 injection sites at dorsoventral=5 and 4 mm. Animals were killed 48 hours later with an overdose of halothane, and brain temperature was equal to rectal temperature during hypothermia. There were no significant differences in other physiological parameters (arterial pH, PaCO₂, and PaO₂) between groups before, during, or after ischemia (Table). Brain temperature was equal to rectal temperature during hypothermia and was 0.4°C to 0.6°C below rectal temperature during normothermia. There were no significant differences in other physiological parameters (arterial pH, PaCO₂, PaO₂, MAPB) between groups before, during, or after ischemia (Table).

**Statistical Analysis**

One-way ANOVA followed by Student-Newman-Keuls post hoc tests were used. Data are presented as mean±SEM.

### Results

#### Physiological Variables

Rectal and striatal temperatures differed as planned between normothermic and hypothermic rats (n=3 per group; Table). Rectal temperature was equal to rectal temperature during hypothermia and was 0.4°C to 0.6°C below rectal temperature during normothermia. There were no significant differences in other physiological parameters (arterial pH, PaCO₂, PaO₂) between groups before, during, or after ischemia (Table).

#### Overexpression of Bcl-2 Plus Hypothermia Protects Against Neuron Loss After Ischemia

Occlusion of the MCA for 1 hour caused a focal infarct within the medial and lateral striatum. Striatal infarct sizes did not differ across groups. Infarct size was not altered by gene transfer because only a limited number of neurons were transfected. Delivery of vectors 5 hours after ischemia onset resulted in local expression in the striatum (Figure 2). Since the number of transfected neurons in nonischemic hemispheres was similar in hypothermic and normothermic rats, we have shown that the bipromoter vector coexpresses transgene and reporter gene similarly. Thus, we compared the percentage of surviving transfected X-gal-positive neurons (relative to the number in the contralateral striatum) in Bcl-2–treated and control animals. Individual X-gal–positive neurons were counted over successive sections in each hemisphere using criteria similar to those in our previous studies, and the number in the ischemic hemisphere was expressed as a percentage of the number in the nonischemic hemisphere.

**Cell Counts**

β-gal–positive neurons and double-labeled (β-gal with cytochrome c) neurons were counted using morphological criteria similar to those used for the X-gal–stained neurons. Since there were no cytochrome-c–positive neurons in the nonischemic striatum, only those in the ischemic striatum were counted. The number of double-labeled neurons was determined from consecutive sections and expressed as a percentage of the total number of β-gal–positive neurons.

### Table

<table>
<thead>
<tr>
<th>Groups (n=3/Group)</th>
<th>Before Ischemia</th>
<th>During Ischemia</th>
<th>5 Hours After Ischemia</th>
<th>Before Ischemia</th>
<th>During Ischemia</th>
<th>2.5 Hours After Ischemia</th>
<th>5 Hours After Ischemia</th>
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<tr>
<td>MABP</td>
<td>93.6±2.4</td>
<td>84.3±9.7</td>
<td>98.0±1.2</td>
<td>96.1±2.8</td>
<td>95.4±3.2</td>
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<td>pH</td>
<td>7.41±0.02</td>
<td>7.39±0.01</td>
<td>7.40±0.02</td>
<td>7.44±0.02</td>
<td>7.39±0.01</td>
<td>7.40±0.01</td>
<td>7.41±0.00</td>
</tr>
<tr>
<td>PaCO₂</td>
<td>43.90±0.93</td>
<td>38.76±1.76</td>
<td>36.40±2.19</td>
<td>42.69±0.75</td>
<td>39.68±1.94</td>
<td>38.48±1.82</td>
<td>40.3±1.22</td>
</tr>
<tr>
<td>PaO₂</td>
<td>134.9±4.96</td>
<td>127.8±3.23</td>
<td>125.8±5.59</td>
<td>115.1±12.70</td>
<td>120.0±4.22</td>
<td>122.5±3.59</td>
<td>119.16±2.39</td>
</tr>
<tr>
<td>Rectal temperature</td>
<td>37.1±0.06</td>
<td>37.1±0.05</td>
<td>37.2±0.06</td>
<td>37.2±0.06</td>
<td>37.1±0.05</td>
<td>33.1±0.04</td>
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</tr>
<tr>
<td>Brain temperature</td>
<td>36.6±0.05</td>
<td>36.7±0.05</td>
<td>36.6±0.05</td>
<td>36.7±0.07</td>
<td>36.6±0.03</td>
<td>33.1±0.04</td>
<td>33.1±0.05</td>
</tr>
</tbody>
</table>

*During hypothermia.*

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temperature did not alter transfection efficiency. Neuron survival in normothermic/control rats did not differ from that in hypothermic/control or normothermic/Bcl-2 rats (Figure 3). This suggests that neither hypothermia nor Bcl-2 alone improved neuron survival 2 days after stroke. In contrast, survival of striatal neurons infected with Bcl-2 in hypothermic rats was improved 2- to 3-fold relative to hypothermic/control rats (P < 0.01, ANOVA) and normothermic/Bcl-2 rats (P < 0.01, ANOVA; Figure 3). This suggests a synergistic effect of Bcl-2 and hypothermia.

Overexpression of Bcl-2 Plus Hypothermia Inhibits Cytochrome c Release

There was no cytochrome c immunoreactivity in the contralateral striatum (Figure 4) or in control tissue in which primary antibody was eliminated (data not shown). Many neurons labeled with cytochrome c and β-gal were present in all control vector–treated animals and in normothermic/Bcl-2 animals. In contrast, the percentage of β-gal–positive neurons that were positive for cytosolic cytochrome c was significantly lower among hypothermic/Bcl-2 rats than in other groups (Figure 5). In the contralateral cortex, β-gal–stained neurons did not colocalize with cytochrome c.

Discussion

This is the first demonstration that postischemic hypothermia prolongs the time window for neuroprotection by gene therapy. In previous studies Bcl-2 overexpression improved neuron survival when delivered 1.5 but not 5 hours after stroke. We now show that hypothermia extends the window for Bcl-2 gene therapy to 5 hours after ischemia onset.

Although intraischemic hypothermia provides long-term protection against cerebral ischemia, postischemic hypothermia may only transiently delay ischemic cell death unless protracted periods of hypothermia are used. Neuroprotective agents may be coupled with short-term hypothermia to enhance its protective effects and to avoid the adverse effects of long-term hypothermia. For example, 3 hours of immediate postischemic hypothermia (30°C) combined with MK-801 (a noncompetitive N-methyl-D-aspartate antagonist) increased CA1 neuron survival over either treatment alone. Similar findings were reported when postischemic hypothermia was combined with either an anti-inflammatory cytokine or a free radical scavenger.

Intraischemic hypothermia protects against cell damage by lowering metabolism and energy demand, inhibiting glutamate release, and preventing dysfunction of the blood-brain barrier. Recently, intraischemic hypothermia has been shown to decrease caspase-3 expression and cytochrome c release after focal ischemia, although one study showed no change in Bcl-2, Bax, or caspase-3 expression. In contrast, mild hypothermia increased Bcl-2 expression after global ischemia. Therefore, the mechanism of hypothermic protection may depend on the nature and severity of the insult.

Unfortunately, few studies have clarified the protective mechanisms of postischemic hypothermia. One recent study

![Figure 2](http://stroke.ahajournals.org/)

Figure 2. Representative sections from striata of 3 ischemic animals transfected with Bcl-2 or control (α4s) vector. Left panels (A, C, E) show X-gal–positive neurons in the contralateral (C) nonischemic hemisphere, and right panels (B, D, F) show X-gal–positive neurons in the ipsilateral (I) ischemic hemisphere. Compared with ischemic striata of hypothermic/Bcl-2 vector rats (Bcl-2 33C/I), fewer intact X-gal–positive neurons remained in normothermic/control vector (α4s 37C/I) and normothermic/Bcl-2 vector rats (Bcl-2 37C/I). Similarly, few intact X-gal–positive neurons remained in the ischemic striata of hypothermic/ control vector rats (data not shown).

![Figure 3](http://stroke.ahajournals.org/)

Figure 3. Survival of Bcl-2– and control vector–transfected neurons after MCA occlusion, represented as the number of X-gal–positive neurons remaining in the ischemic hemisphere relative to the number in the nonischemic hemisphere. In the normothermia groups, X-gal–positive neurons in control animals totaled 122 ± 23 in ischemic and 339 ± 44 in nonischemic striatum; in Bcl-2–treated animals there were 135 ± 42 X-gal–positive neurons in ischemic and 345 ± 68 in nonischemic striatum. In the hypothermia groups, X-gal–positive neurons in control animals totaled 132 ± 39 in ischemic and 309 ± 27 in nonischemic striatum. Striatal neuron survival of hypothermic/Bcl-2 rats (33C/Bcl-2) was improved 2- to 3-fold compared with hypothermic/control rats (33C-control) (P < 0.01), normothermic/Bcl-2 rats (37C-Bcl-2) (P < 0.01), and normothermic/control rats (37C-control) (P > 0.01).
showed that postischemic hypothermia only delayed neutrophil accumulation and microglial activation, which may account for the lack of persistent protection. However, our laboratory demonstrated that hypothermia inhibits leukocyte infiltration as late as 7 days and inflammatory cell generation of inducible nitric oxide synthase, nitric oxide, and peroxynitrite (ONOO⁻). Delayed cooling attenuates neuronal nitric oxide synthase expression to a greater extent than intraischemic hypothermia. It is still unclear why we found synergistic effects of Bcl-2 and hypothermia. Transgene expression from HSV vectors requires several hours, with peak expression occurring approximately 12 hours after delivery to brain. Bcl-2, originally characterized as an antiapoptotic protein, can also block necrosis. Bcl-2 has various roles within cells, including increasing mitochondrial calcium uptake, blocking Bax translocation, and inhibiting cytochrome c release. We have recently shown that overexpression of Bcl-2 decreased cytochrome c release when delivered before focal ischemia. In the current study Bcl-2 overexpression did not significantly inhibit cytochrome c release in normothermic rats, perhaps because the vector was delivered 5 hours after ischemia onset. In contrast, Bcl-2 plus hypothermia significantly reduced cytochrome c release. Our previous study demonstrated that intraischemic hypothermia reduced cytochrome c release at 5 but not 24 hours after stroke. Similarly, in this study, although hypothermia itself did not inhibit cytochrome c release 48 hours after stroke, it may have delayed the onset of cytochrome c release. This delay could allow the late expression of Bcl-2 to block cytochrome c release and protect against neuronal death. Hypothermia in the current study may also have inhibited or delayed some other aspects of cell death, such as intracellular calcium accumulation, generation of free radicals, or caspase activation. This inhibition or delay could allow Bcl-2 time to block these detrimental events and protect against neuronal death.

Hypothermic rats experienced postischemic isoflurane anesthesia for approximately 3 hours longer than normothermic rats, which may have influenced the outcome beyond the
effects of hyperthermia. However, isoflurane did not influence any of the hemodynamic or physiological variables, nor did it alter brain temperature in this study (Table). Additionally, whether isoflurane itself protects against cerebral ischemia is not clear. Many reports disagree, and some argue that it provides little protection or even worsens ischemic damage.34–37 In reports that demonstrated that isoflurane reduced the infarct caused by focal ischemia, isoflurane was applied during rather than after ischemia.38,39 It is unknown whether postischemic isoflurane provides protection. The effect of differences in postischemic isoflurane exposure would likely be small relative to the protective effect of hyperthermia. Although we cannot completely exclude an influence of isoflurane, we conclude that hyperthermia prolongs the therapeutic time window for gene therapy.

Conclusion

Brief, mild postischemic hyperthermia prolonged the temporal therapeutic window for Bcl-2 gene therapy from 1.5 to 5 hours, and Bcl-2 plus hyperthermia blocked cytochrome c release 48 hours after ischemia onset. These data demonstrate a synergistic effect of hyperthermia and Bcl-2 overexpression, suggesting a potential clinical application of combined hypo- 

thermia and gene therapy.

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References


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