Doxycycline Suppresses Cerebral Matrix Metalloproteinase-9 and Angiogenesis Induced by Focal Hyperstimulation of Vascular Endothelial Growth Factor in a Mouse Model

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Background and Purpose—A number of central nervous system (CNS) disorders are associated with abnormalities in or activation of angiogenesis, including vascular malformations. To test the hypothesis that the nonspecific matrix metalloproteinase (MMP) inhibitor, doxycycline, suppresses vascular endothelial growth factor (VEGF)-induced cerebral angiogenesis through inhibition of MMPs, we used a mouse model with enhanced cerebral angiogenesis induced by focal hyperstimulation of VEGF from adenovirus DNA (AdVEGF) transduction.

Methods—The time course study of MMP activity was performed at 7 and 14 days after AdVEGF transduction. MMP activity and expression were examined by zymography and immunohistochemistry, respectively. As an index of cerebral angiogenesis, microvessel counting was performed in the brains of 3 groups of mice (n = 6): (1) control; (2) AdVEGF only; and (3) AdVEGF plus doxycycline (30 mg/kg per day).

Results—Brain MMP-9 activities increased 4-fold (883 ± 137 versus 179 ± 179; 1-sided P < 0.001) at 7 days after AdVEGF transduction. VEGF transduction increased vessel counts by 19% (255 ± 27 versus 215 ± 15, 1-sided P < 0.01). Doxycycline treatment decreased MMP-9 activity (89 ± 72 versus 883 ± 137; 1-sided P < 0.001) and cerebral microvessel number (231 ± 17 versus 255 ± 27; 1-sided P < 0.05).

Conclusions—Doxycycline is effective in decreasing stimulated cerebral MMP-9 activity and parenchymal angiogenesis. The decrease in MMP-9 activity is associated with decreased microvessel counts. Brain pathophysiological processes that involve abnormally enhanced angiogenesis may be amenable to manipulation by MMP inhibitors, including tetracycline derivatives. (Stroke. 2004;35:1715-1719.)

Key Words: drug therapy • metalloproteinases • angiogenesis
There is growing evidence that MMP inhibition may be useful in the management of vascular diseases. Tetracycline derivatives, including doxycycline, have nonspecific MMP inhibitory effects that are distinct from their antimicrobial action. To study the effect of MMP inhibition on states of enhanced angiogenesis, we used a murine model newly developed in our laboratory. The model consists of focal hyperstimulation by VEGF165 in the brain using adenovirus-mediated DNA transduction, resulting in enhanced formation of cerebral microvessels. Our model is not a model of BAVM, but it is characterized by some of the phenotypic features of BAVM lesional tissue. Therefore, use of this model is a first step to mechanistically examine key angiogenic pathways and their response to pharmacological manipulation.

We hypothesized that the nonspecific MMP inhibitor, doxycycline, can suppress AdVEGF-induced cerebral angiogenesis and the suppression is mediated through inhibition of MMP-9 expression and activity. To test this hypothesis, we examined the effects of doxycycline on MMP activities and microvessel formation in the mouse brain after adenovirus-mediated VEGF transduction.

**Materials and Methods**

**Animals and Treatment**
Animal use was approved by the University of California San Francisco Committee of Animal Research. Male CD-1 mice weighing 30 to 35 g were purchased from Charles River Laboratory (Wilmington, Mass). The mice were allowed free access to food and water with a 12-hour alternating light–dark cycle.

Our previous data have shown that VEGF expression increases at day 5 in the mouse brain after AdVEGF transduction compared with the control group, with the peak of microvessel counts occurring later, at ~3 weeks. This finding suggests that there is significant lag time between the activation of angiogenic factors and the actual formation of new vessels. At day 3, mild inflammatory responses around the needle track was detected in both AdlacZ and AdVEGF groups to a similar degree. A time course study at 7 and 14 days after AdVEGF transduction was performed to determine the appropriate time point for assessing MMP expression change induced by VEGF overexpression.

To study the effect of doxycycline treatment on cerebral angiogenesis induced by AdVEGF transfection, the mice were divided into 3 groups: control, AdVEGF, and AdVEGF with doxycycline treatment. The control and AdVEGF groups received AdlacZ and AdVEGF injection, respectively. In the treatment group, doxycycline (Sigma) was administered starting on the day of AdVEGF injection, at 30 mg/kg per day via drinking water, a dose shown to inhibit growth of aortic aneurysm in rodents.

**Adenoviral-Mediated VEGF Gene Transfer in the Brain**

After induction of anesthesia with ketamine and xylazine (intraperitoneally), mice were placed in a stereotactic frame (David Kopf Instruments). A Hamilton syringe was inserted through a burr hole intraperitoneally, mice were placed in a stereotactic frame (David Kopf Instruments). A Hamilton syringe was inserted through a burr hole and the control group, with the peak of microvessel counts occurring later, at ~3 weeks. This finding suggests that there is significant lag time between the activation of angiogenic factors and the actual formation of new vessels. At day 3, mild inflammatory responses around the needle track was detected in both AdlacZ and AdVEGF groups to a similar degree. A time course study at 7 and 14 days after AdVEGF transduction was performed to determine the appropriate time point for assessing MMP expression change induced by VEGF overexpression.

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**Tissue Collection**

Coronal sections of brain tissues including 1 mm anterior and posterior to the injection site were quickly frozen in liquid nitrogen, stored at −80°C, and used for zymography. For microvessel counting and immunostaining, the whole brain was snap-frozen in isoprene with dry ice and stored at −80°C. The tissue was sectioned with a cryostat at 16-μm intervals.

**Microvessel Counting**

Mice were sacrificed for brain microvessel counting at 3 weeks after adenoviral-mediated gene transfer. The decision of timing was based on our previous data showing that the number of newly formed brain microvessels peaks at 3 weeks after AdVEGF transduction. Frozen sections were fixed with 100% ETOH at −20°C, then incubated with fluorescein-lycosperiscin esculentum lectin (Vector Laboratories) 2 μg/ml at 4°C overnight. Three areas of microvessels, left, right, and bottom to the needle track, respectively, were chosen in 2 separate brain coronal sections. Microvessel numbers were counted in images captured from these areas by using National Institutes of Health Image J 1.29x. The number of microvessels was calculated as the mean of the numbers obtained from the 6 pictures. Two investigators blinded to the animal treatment condition confirmed the vessel counts manually.

**Gelatin Zymography**

Equal amounts of sample proteins were separated by electrophoresis on 10% zymogram gels (Invitrogen). The gels were subsequently stained with colloidal blue stain (Invitrogen). Proteolytic bands in zymography were quantified by scanning densitometry using KODAK image analysis software (Eastman Kodak).

**Immunohistochemistry**

Tissue sections were fixed in 4% paraformaldehyde for 30 minutes. After blocking endogenous peroxidase with 1% hydrogen peroxide in 100% methanol followed by preincubation with 0.5% bovine serum albumin, anti-mouse MMP-9 antibody (R&D Systems) was applied at 4 μg/ml for overnight at 4°C. The sections were then incubated with biotinylated rabbit anti-goat IgG (Vector Laboratories) for 1 hour at room temperature, followed by incubation with streptavidin-HRP (BioCare). Chromogenic staining was developed using DAB kit (Zymed) and followed by counter staining.

**Statistical Analysis**

Data are expressed as mean±standard deviation. Parameters between different groups in the MMP expression time course study and doxycycline treatment study were analyzed using 2-way ANOVA and Student t test. Because theory and previous results distinctly predict an increase of MMP activity and microvessel formation with VEGF administration and a reduction with the added administration of doxycycline, 1-sided P values were used for those comparisons; P<0.05 is considered statistically significant.

**Results**

To determine the appropriate time point to examine MMP expression induced by VEGF, the time course of MMP expression after AdVEGF transduction was studied (Figure 1). At 7 days, acute inflammatory response was minimal in all of the animals by H&E staining (data not shown). At day 7 after gene transfer, MMP-9 activities were increased near 4-fold in the AdVEGF transducted mouse brain compared with the ones with AdlacZ (883±137 versus 179±179, arbitrary units [AU] 1-sided P<0.001). At day 14, there was little MMP-9 detected in either of the groups (73±18 versus 165±94 AU, 1-sided P>0.05), and the differences in MMP-9 levels were greater at 7 days than at 14 days (2-way factorial test for interaction, P<0.01). Therefore, day 7 after gene transfer appeared to be an appropriate time for the MMP study. Unlike MMP-9, there was no significant change in MMP-2 expression in response to VEGF either at day 7 (3363±1204 versus 2997±998 AU, 1-sided P>0.2) or at day...
14 (2688±615 versus 3329±661 AU, 1-sided \(P<0.2\)) (see zymograph in Figure 1A; bar graph not shown).

Immunohistochemistry staining was performed to compare the distribution of MMP-9 expression at day 7 after gene transfer (Figure 2). Diffuse positive staining of MMP-9 was distributed in surrounding areas of the needle track after AdVEGF transduction, as illustrated in Figure 2B, but not in the Ad\(\lambda\)cZ mouse brain (Figure 2A).

Doxycycline suppressed MMP-9 expression at 7 days after AdVEGF transduction in mouse brain (Figure 3). MMP-9 activities were much lower in doxycycline-treated mice than those that did not receive the drug (89±72 versus 883±137 AU, 1-sided \(P<0.001\)). In contrast, the Ad\(\lambda\)cZ group was not statistically significantly different from the AdVEGF plus doxycycline group (179±179 versus 89±72 AU, 2-sided \(P>0.4\)). In other words, the VEGF-induced MMP-9 activities were completely diminished by doxycycline treatment. Again, unlike MMP-9, there was no change of MMP-2 activity in response to doxycycline treatment (2997±998 versus 2684±821 AU, 1-sided \(P>0.3\)) (Figure 3A).

To determine the effect of doxycycline on cerebral angiogenesis, 3 groups of mice (\(n=6\) in each group) at 3 weeks after adenoviral DNA transduction were used for comparison. Mice with Ad\(\lambda\)cZ transfer in the brain were used as the control group. Our results showed that doxycycline decreased microvessel counts induced by focal VEGF hyperstimulation in the mouse brain (Figure 4). VEGF transduction increased microvessel formation by 19% in the mouse brain in comparison with the \(\lambda\)cZ group (255±27 versus 215±15, number of microvessels; 1-sided \(P=0.01\)). The number of microvessels was lower in the doxycycline group than in the VEGF group (231±17 versus 255±27, number of microvessels; 1-sided \(P<0.05\)).

**Discussion**

Our results have demonstrated for the first time to our knowledge that doxycycline suppresses VEGF-induced cerebral MMP-9 activity in vivo. The doxycycline-induced changes in MMP-9 activity were associated with decreased regional angiogenesis, as evidenced by decreased microvessel counts.

Previous reports have shown that tetracycline derivatives, including doxycycline, influence many aspects of the angiogenesis process.\(^{18}\) Many of those data, however, were obtained from in vitro systems or from large blood vessels in animals. First, there have been conflicting results of the effect of doxycycline...
on cell growth between in vitro and in vivo studies, suggesting the possibility of different mechanisms. For example, in vivo studies on smooth muscle cell proliferation showed that effects of doxycycline differ from known MMP inhibitors, including GM6001 and BB94. However, in vitro studies using tissue culture showed contradictory results. One reported that doxycycline inhibited angiogenesis whereas GM6001 did not. The other one reported that the antiangiogenesis effects of tetracycline derivatives were associated with inhibition of MMP activities. Secondly, vascular endothelium in the central nervous system may be functionally distinct from the endothelium of other organ systems. Doxycycline has been shown to inhibit MMP-9 in aorta homogenate cultures and in human and animal aortic aneurysm studies. However, there are significant differences in baseline endothelial MMP activities between the brain microvessel and the aorta. For example, MMP-9 levels increased in the brain microvessel endothelial cells in response to stimulation by inflammatory cytokines, whereas no change was observed in the aortic endothelial cells. Our finding that doxycycline inhibits MMP-9 activity and the formation of capillaries in the mouse brain provides evidence that doxycycline can influence cerebral angiogenesis.

VEGF and MMPs are considered as potent regulators of angiogenesis. Our findings have shown that focal VEGF hyper-stimulation is associated with increased MMP-9 expression in the mouse brain. Similarly, Lamoreaux et al reported that VEGF increases the release of another gelatinase, MMP-2, and decreases the release of tissue inhibitors of metalloproteinases by microvascular endothelial cells in vitro. Further, MMP-9 can facilitate the availability of tissue-bound VEGF, which in turn may potentiate angiogenic activities.

There are somewhat contradictory evidence of MMP-9 and MMP-2 in response to stimulatory or inhibitory factors. In our study, there was no change in MMP-2 levels with VEGF stimulation or after doxycycline treatment, in contrast to changes in microvessels and MMP-9 levels. In the brain, in addition to endothelial cells and smooth muscle cells, astrocytes constitutively produce MMP-2. MMP-2 activities that we detected from all groups of mice regardless of the treatment could be a reflection of constitutive expression of MMP-2, which, in this case, does not respond to either the stimulation from VEGF or the inhibition from doxycycline.

There are several limitations to our study. As pointed out, this model is not a specific model for any particular disease but can
allow mechanistic investigations of various potential interventions in the angiogenic process. Ideally, we would have had performed a dose–response study, but the dose used appears to be sufficient to provide proof-of-concept for the hypothesized effects of doxycycline, taken together with other information in the literature. An inflammatory tissue response from adenoaviral transduction may confound the direct effects of VEGF overexpression, but our previous studies have demonstrated a minimal degree of acute inflammation with this model.\(^1\)\(^2\) Finally, we have only demonstrated an association between decreased MMP-9 activity and diminished ability of VEGF to induce capillary angiogenesis; further studies can better characterize the causal relationship of the 2 observations.

In conclusion, the present study has demonstrated that doxycycline can reduce MMP-9 activity and angiogenesis induced by focal VEGF hyperstimulation in the mouse brain. The ability to manipulate angiogenesis may have importance in the study of various CNS disorders, and in particular may be of interest in developing models to study the pathogenesis of brain vascular malformations. The mechanism of the effect of doxycycline on brain angiogenesis in relation to its anti-MMP activity remains to be further clarified.

Acknowledgments
Supported in part by NIH T32 GM08840 (C.Z.L.) and NIH R01 NS27713 and K24 NS02091 (W.L.Y.). The authors wish to thank Broderick Belenson, Achal Achrol, Manju Chopra, Gaurab Basu, and members of the UCSF BAVM study project for their assistance in data collection and manuscript preparation.

References
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Stroke. 2004;35:1715-1719; originally published online May 27, 2004; doi: 10.1161/01.STR.0000129334.05181.b6

Stroke is published by the American Heart Association, 7272 Greenville Avenue, Dallas, TX 75231
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Print ISSN: 0039-2499. Online ISSN: 1524-4628

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http://stroke.ahajournals.org/content/35/7/1715

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