HMG-CoA Reductase Inhibitor, Atorvastatin, Promotes Sensorimotor Recovery, Suppressing Acute Inflammatory Reaction After Experimental Intracerebral Hemorrhage

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Background and Purpose—Statins have neuroprotective effects on ischemic stroke. They modify the endothelial function, increase blood flow, and inhibit thrombus formation, which are independent of lipid-lowering effects. However, whether statins have a protective effect toward hemorrhagic stroke is yet unknown. To test this possibility, we attempted to determine the effect of atorvastatin on experimental intracerebral hemorrhage (ICH).

Methods—ICH was induced using stereotaxic infusion of collagenase into the left basal ganglia in adult rats. Atorvastatin (1 mg/kg or 10 mg/kg) or phosphate-buffered saline was administered for 2 weeks. To monitor the sensorimotor deficits, limb placing and Rotorod tests were performed. Hematoma volume, brain water content, and hemispheric atrophy were analyzed. Immunohistochemical staining for myeloperoxidase (MPO), microglia (OX42), inducible nitric oxide synthase (iNOS), or endothelial nitric oxide synthase (eNOS) was performed. Perihematomal cell death was determined by TUNEL staining.

Results—The atorvastatin-treated ICH group showed better performance on Rotorod and limb placing tests when compared with the vehicle-treated group (P<0.01). The hematoma volumes between groups were not different, but the brain water content and hemispheric atrophy were reduced in the atorvastatin-treated ICH group. Atorvastatin reduced TUNEL-positive cells, iNOS expression, and MPO-positive or OX42-positive cells in the perihematomal regions in a dose-dependent manner, whereas it increased eNOS expression.

Conclusion—The present study shows that atorvastatin reduces the perihematomal cell death via antiinflammation, which is associated with sensorimotor recovery after experimental ICH. (Stroke. 2004;35:1744-1749.)

Key Words: HMG-CoA reductase inhibitors ■ intracerebral hemorrhage ■ brain edema ■ apoptosis ■ inflammation

Intracerebral hemorrhage (ICH) represents 15% of all strokes in Western populations and a higher proportion (20% to 30%) in Asian and black populations.1 The mechanisms by which cellular injury occurs after ICH include mechanical destruction or displacement caused by blood accumulation.2 Inflammation and impairment of cerebral blood flow (CBF) around the blood clot contribute to delayed neuronal death.3,4 The suppression of the inflammatory response has been reported to reduce brain edema and tissue damage and improve functional outcome in experimental ICH.5–8

Statins, 3-hydroxy-3-methyl-glutaryl-coenzyme A (HMG-CoA) reductase inhibitors, have been shown to have many pleiotropic properties on the nervous and vascular systems, independent of cholesterol-lowering effect, and provide neuroprotection with beneficial effects on endothelial function, immunomodulation, and impeding excitotoxic cell injury via antiinflammatory actions and angiogenesis/neurogenesis.9–22 The putative neuroprotective actions of statins may lead to functional restoration after ICH. However, the effects of statins in the ICH paradigm are not known at present. In the present study, we investigate whether atorvastatin, when administered after ICH, can reduce edema formation, inflammation, and cell death, and subsequently promote neurological recovery in a rat model of ICH.

Materials and Methods

Induction of Intracerebral Hemorrhage and Atorvastatin Administration

Male Sprague-Dawley rats (12 weeks old; Genomics, Seoul, ROK), weighing 200 to 220 grams, were used in this experiment and our protocols were institutionally approved in accordance with NIH...
Experimental groups consisted of rats subjected to ICH (n=99) and normal control rats (n=27). Experimental ICH was induced by the stereotaxic intrastriatal administration of bacterial collagenase type IV (Sigma), as previously described.23,24 After an intraperitoneal injection of 1% ketamine (30 mg/kg; Sigma) and xylene hydrochloride (4 mg/kg; Sigma), rats were placed in a stereotaxic frame (David Kopf Instruments). A burr hole was made, and a 30-gauge Hamilton syringe needle was inserted through a burr hole into the striatum (location: 3.0 mm left lateral to the midline, 0.2 mm posterior to bregma, 6 mm in depth below the skull). ICH was induced by the administration of 1 µl containing 0.23 U of collagenase over 5 minutes. After infusion, craniotomy holes were sealed with bone wax and wounds were sutured. Rectal temperature was maintained at 37±0.5°C using a thermistor-controlled heating blanket.

Six hours after ICH induction (considering the recovery time from anesthesia), rats were fed atorvastatin (prescription formulation; Pfizer Inc), using 20-mm feeding needles (Popper and Sons Inc), dissolved in phosphate-buffered saline, 1 mg/kg (n=21) or 10 mg/kg (n=39) daily for 3 days or 14 days (to the rats assigned for the behavioral testing). ICH rats fed vehicle alone served as ICH control group (n=39), and normal rats fed vehicle alone (n=18) or 10 mg/kg atorvastatin (n=9) served as normal control group.

Behavioral Testing
Behavioral testing was performed weekly for atorvastatin-treated (10 mg/kg) or vehicle-treated ICH group (n=9 per group) using the modified limb placing (MLPT) and Rotorod tests,23 which were monitored by blinded investigators for group allocation.

Morphometric Measurement of Hematoma Volume and Hemispheric Atrophy
At 3 days after ICH, brains from atorvastatin-treated (10 mg/kg) or vehicle-treated ICH rats (n=9 per group) were cut coronally through the needle entry site (identifiable on the brain surface), and then serial slices (1-mm thickness) both anterior and posterior to the needle entry site were obtained. Digital photography of the serial slices was taken and hematoma volume was measured using image analyzer program (Image Pro-Plus; Media Cybernetics). The total hematoma volume (mm³) was calculated by summing the clot area in each section and multiplying by the distance between sections.24 At 42 days after ICH, the rats used for behavior test were euthanized to analyze hemispheric atrophy. Three sections through the needle entry site and sites 1.0 mm anterior and 1.0 mm posterior to plane were Nissl-stained. The morphometric analyses involved the area of the striatum, cerebral cortex, and ventricle, as well as the whole hemisphere. The total hemispheric areas of each section were traced and measured using image analyzer. The hemispheric atrophy was expressed as a percentage of contralateral hemispheric area.25

Analysis of Brain Water Content, Histology, and Immunohistochemistry
We determined the endpoint of 3 days for experiments on water content, histology, and immunohistochemistry, because inflammatory reaction has been shown to be maximal at 48 to 72 hours after ICH.2 Atorvastatin-treated (1 mg/kg or 10 mg/kg) or vehicle-treated ICH rats and atorvastatin-treated (10 mg/kg) or vehicle-treated normal rats (n=9 per group) were euthanized to measure brain water content. The brains were divided into 2 hemispheres along the midline, and the cerebellum and the brain stem were removed. The brain samples were immediately weighed on an electronic analytical balance to obtain the wet weight. The samples were then dried in a gravity oven at 100°C for 24 hours to obtain the dry weight. Water content was expressed as a percentage of wet weight: the formula for calculation was (water weight–dry weight)/(wet weight) × 100.24 At the same time, atorvastatin-treated (1 mg/kg or 10 mg/kg) or vehicle-treated ICH rats (n=9 per group) and vehicle-treated normal rats (n=6) were euthanized for histological analysis. We removed the brains and cryopreserved tissue blocks for cryostat sectioning (30 µm thickness). Euthanization or immunohistochemistry was processed as described previously.23,24 Rabbit anti-human myeloperoxidase (MPO) (1:100; DAKO Corporation) and mouse anti-rat Ox42 primary antibodies (1:500; Chemicon) were used as markers for neutrophil and microglia, respectively. MPO-positive or Ox42-positive cells were identified and counted. The iNOS and eNOS immunohistochemistries were performed with anti-iNOS antibody (1:300, mouse monoclonal; Transduction Laboratories) and anti-eNOS antibody (1:500, rabbit polyclonal; Transduction Laboratories), respectively. FITC-conjugated anti-mouse IgG (1:100; Jackson ImmunoResearch) and Cy3-conjugated anti-rabbit IgG antibodies (1:500; Jackson ImmunoResearch) were used for the secondary antibodies.

Western Blot
Western blotting was performed on perihematomal tissues dissected out from a 2-mm-thick coronal brain slice cut at the level of the needle insertion. A similar segment was isolated in normal control rats. Western blotting of eNOS (n=3 per group) was performed with a rabbit polyclonal eNOS antibody (1:500; Transduction Laboratories) as previously described.23 Using the same membrane, we also performed immunoblotting of β-actin with a mouse anti-β-actin monoclonal antibody (1:2000; Sigma) to standardize the expression of eNOS. Equal amounts of protein (20 µg) for each group were assayed and immunoreactivity was visualized by enhanced chemiluminescence.

TUNEL Assay
Atorvastatin-treated (1 mg/kg or 10 mg/kg) or vehicle-treated ICH rats (n=9 per group) and vehicle-treated normal rats (n=6) were euthanized with sodium pentobarbital at 3 days after ICH for TUNEL staining. The TUNEL procedure for in situ detection of DNA fragmentation was performed as previously described.23 Only TUNEL-stained cells that showed deep punctate staining with perinuclear chromatin condensation were considered apoptotic and were included for quantitation. Cell density counts were performed on sections counterstained with toluidine blue.

Cell Quantification
Quantitative analysis of the positively stained cell number was performed in the perihematomal regions of 3 axial sections (1-mm width) staining through the center of the hemorrhagic lesion by 2 independent investigators who were blinded to the group allocation. Total counts in the measured sections were converted into cell densities for comparison between the ICH groups.

Statistical Analysis
All data in this study are presented as the mean±SD. Data were analyzed by unpaired Student t test if they were normally distributed (Kolmogorov-Smirnov test, P>0.05). Otherwise, we used Mann–Whitney U test and specified the test used. Two-tailed value of P<0.05 was considered significant.

Results
Physiological Parameters
All animals survived the surgery. The physiological parameters, including mean arterial blood pressure, blood gases, serum glucose, and body temperatures, were not significantly different in any experimental groups before, during, or after ICH.

Atorvastatin Improved the Sensorimotor Deficits After Experimental Rats
The atorvastatin-treated (10 mg/kg) ICH group showed a better performance on the MLPT after 21 days and the Rotorod after 14 days compared with the vehicle-treated group, and the effects persisted up to 28 days. At day 28, the Rotorod performance showed 80% improvement of pre-
trained level, whereas the vehicle-treated ICH group showed 40% (Figure 1; n=9 for each group, P<0.05, Mann–Whitney U test). The initial body weights and those over the course of 4 weeks were similar.

Atorvastatin Reduced Brain Water Content and Long-Term Hemispheric Atrophy

The hematoma volume was 19.16±1.99 mm³ in the vehicle-treated ICH group and 18.83±1.58 mm³ in the atorvastatin-treated group by 3 days (Figure 2A; n=9 for each group, P=0.66, Student t test). Brain water contents of the lesioned hemisphere were reduced in atorvastatin-treated ICH group with dose-dependency (Figure 2B; n=9 for each group, P<0.05, Mann–Whitney U test). Brain water contents of the nonlesioned hemisphere were not different. Hemispheric area analysis showed an atrophy of lesioned hemisphere in both vehicle-treated and atorvastatin-treated ICH groups (Figure 2C; n=9 for each group). However, the atorvastatin-treated ICH group showed lesser hemispheric and striatal atrophy than the vehicle-treated group (15.00±2.16% versus 24.52±4.79% and 15.50±2.38% versus 27.00±3.91%, respectively; P<0.05, Mann–Whitney U test), whereas cortical atrophy was not different between both groups (5.75±2.06% versus 6.75±2.50%; P=0.46).

Atorvastatin Decreased TUNEL-Positive Cells in the Perihematomal Regions

After 3 days, a high density of TUNEL-positive stained cells was present within the hemorrhage as well as in the surrounding periphery (Figure 3). Counterstaining with toluidine blue revealed that the TUNEL-positive cells were not leukocytes. TUNEL-positive cells in atorvastatin-treated groups were significantly decreased by 42% (10 mg/kg) and 24% (1 mg/kg) compared with vehicle-treated group (n=9 for each group, P<0.05, Student t test).
Atorvastatin Decreased iNOS Expression and MPO-Positive or OX42-Positive Cells

iNOS immunoreactivity was increased in the perihematomal regions, but it was attenuated by \( \approx 50\% \) in the atorvastatin-treated (10 mg/kg) ICH group (Figure 4A and 4B). A significant infiltration of MPO-positive or OX42-positive cells adjacent to the hematoma was also detected 3 days after ICH (Figure 4C to 4F). Quantitative analysis (Figure 4G and 4H) showed 36% reduction in MPO\(^+\) and 42% in OX42\(^+\) cells in the atorvastatin-treated (10 mg/kg) ICH group compared with the vehicle-treated group, whereas it showed a minimal reduction in the atorvastatin-treated (1 mg/kg) ICH group.

Atorvastatin Increased eNOS Expression in the Perihematomal Regions

The eNOS expression in the perihematomal regions was increased in the atorvastatin-treated (10 mg/kg) ICH group compared with the vehicle-treated group (Figure 5). In addition, a 2-fold increase in eNOS protein was detected by Western blotting after 3 days of 10 mg/kg treatment, whereas the increase was minimal after 1 mg/kg treatment (Figure 5C).

Discussion

In the present study, atorvastatin, when administered to collagenase-induced ICH model, reduced hemispheric water content and perihematomal cell death, decreased iNOS...
Toxic blood components can induce an inflammatory response in and around the hematoma. Infiltrating inflammatory cells or activated microglia enhance the production of proinflammatory cytokines, cyclooxygenase-2, and iNOS, presumably inducing the signal pathway to mediate the apoptosis. Inhibition of these inflammatory substances has been reported to reduce a brain edema and tissue damage and improve functional outcome in experimental ICH. Statins cause a dose-dependent inhibition of multiple steps of leukocyte recruitment and migration into the central nervous system, either directly by suppressing cell surface receptors and adhesion molecules on nervous or vascular tissues or indirectly by binding to leukocyte function-associated antigen. Statins can also suppress the proinflammatory mediators such as TNF-α, IFN-γ, and iNOS. Recent data have demonstrated that statins act beneficially in numerous inflammatory conditions such as cardiac transplantation (in humans) and experimental autoimmune encephalomyelitis (in mice). In our experiment, attenuation of inflammatory response by atorvastatin might mediate decreasing brain edema and cell death.

Statins have also been shown to enhance CBF in experimental stroke. Statins upregulate eNOS in ischemic stroke and experimental subarachnoid hemorrhage, indicating that enhanced eNOS activity by statins is the predominant mechanism by which these agents protect against cerebral injury. In our experiment, eNOS was upregulated in the perihematomal regions. Thus, atorvastatin might attenuate the ischemic neuronal injury in the perihematomal ischemic penumbra by enhancing CBF. To clarify other pleiotropic effects of statin including neurogenesis and angiogenesis, further investigations are needed.

The patients with ischemic stroke often undergo hemorrhagic stroke. Up to 30% of all ischemic strokes develop hemorrhagic transformations, and thrombolytic therapy of ischemic stroke can lead to serious complications of ICH. Therefore, in practice, it is important to identify agents that are safe and efficacious in treating both ischemic and hemorrhagic stroke. Taken together with the pleiotropic effects of statin in acute ischemic stroke, statin might be a useful agent in these complex conditions.

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References


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