Background and Purpose—Inflammation is thought to be a major contributor to carotid artery disease. Lipopolysaccharide (LPS) activates inflammatory mechanisms thought to contribute to endothelial dysfunction by mechanisms that are not well defined. The goal of this study was to determine whether overexpression of CuZn-SOD protects against LPS-induced increases in superoxide and endothelial dysfunction.

Methods—Carotid arteries from CuZn-SOD transgenic (SOD-Tg) and nontransgenic (non-Tg) littermates were examined in vitro. Superoxide levels were measured using lucigenin-enhanced chemiluminescence.

Results—In non-Tg mice, LPS (0.5 μg/mL for 22 hours) produced marked impairment of vasorelaxation in response to the endothelium-dependent dilator acetylcholine (ACh). For example, 100 μmol/L ACh relaxed carotid arteries from non-Tg mice by 86±6% and 38±8% after treatment with vehicle and LPS, respectively. In contrast, LPS did not significantly impair responses of carotid artery to ACh in SOD-Tg mice, and LPS had no effect on relaxation responses to the endothelium-independent dilator nitroprusside in carotid artery from non-Tg or SOD-Tg mice. LPS-induced increases in superoxide, as measured using lucigenin-enhanced chemiluminescence, were higher in vessels from non-Tg mice than from SOD-Tg mice.

Conclusions—These results indicate that LPS increases superoxide and impairs endothelium-dependent relaxation. Overexpression of the CuZn isoform of SOD effectively prevents LPS-induced oxidative stress and endothelial dysfunction in the carotid artery. (Stroke. 2004;35:1963-1967.)

Key Words: animal models  ■  carotid arteries  ■  endothelium  ■  inflammation  ■  reactive oxygen species  ■  superoxide dismutase
stroke.22 Using CuZn-SOD transgenic (SOD-Tg) and non-transgenic (non-Tg) littermates, we found that LPS increases superoxide and impairs endothelium-dependent relaxation, and that these effects are prevented by overexpression of CuZn-SOD.

Methods

Animal Preparation
The animal protocol used in these experiments was reviewed and approved by the University of Iowa Animal Care and Use Committee. Mice for this study were derived from breeding of male hemizygous CuZn-SOD (human)-transgenic (C57BL/6-TgN(SOD-1)110je) mice with female C57BL/6j mice to generate SOD-Tg and non-Tg mice within the same litter (Jackson Labs, Bar Harbor, Maine). This approach allowed us to use non-Tg littermates as controls. Mice were fed regular chow and water, and available ad libitum. The ages of mice in the different groups were similar (non-Tg, 10±1 month; SOD-Tg, 11±1 month), and both male and female mice were studied. Genotyping of mice was performed using PCR of DNA from tail biopsies.

General Preparation
The method used to measure responses of carotid arteries in mice has been described in detail.12,23,24 Briefly, mice were anesthetized with pentobarbital (100 mg/kg, intraperitoneally) followed by removal of both carotid arteries and the thoracic aorta. Arteries were placed in Krebs buffer, loose connective tissue was removed, and vessels were cut into rings (3 to 4 mm in length). Each segment of carotid artery and aorta was placed in individual wells using 48-well cell culture dishes containing 0.5 mL DMEM containing 5 mmol/L glucose, 120 U/mL penicillin, 120 μg/mL streptomycin, and 50 μg/mL polymyxin B. Vessels were then incubated with either vehicle (ddH2O) or LPS (0.5 μg/mL Escherichia coli, serotype 026:B6; Sigma-Aldrich, St. Louis, Missouri) for 22 hours at 37°C. Following incubation, vascular rings were connected to a force transducer to measure isometric tension in an organ bath containing Krebs solution maintained at 37°C. Resting tension was increased stepwise to reach a final tension of 0.25 g, and the rings were allowed to equilibrate for at least 45 minutes. This amount of resting tension is optimal for contraction of murine carotid arteries.

Protocol
Relaxation of carotid arteries in response to acetylcholine (Sigma) and nitroprusside (Sigma) was measured following submaximal precontraction using the thromboxane analog, U46619 (9,11-dideoxy-11a,9a-epoxy-methanoprostaglandin F2α; Biomol Research Laboratories Inc). Using pharmacological approaches and gene-targeted mice, we and others have shown previously that responses of the carotid artery to acetylcholine are mediated by endothelial nitric oxide synthase (eNOS).25 At the end of each experiment, we obtained full dose responses of each carotid artery to U46619 (0.03 to 3 μg/mL) to determine maximal contractile responses.

Measurement of SOD Protein and Activity
Total SOD activity of aortic homogenates from non-Tg and SOD-Tg mice was determined as described.24 CuZn-SOD protein expression in the aorta of both groups of mice was examined by Western blotting as described.24

Measurement of Superoxide
Superoxide levels were measured using aorta and 5 μmol/L lucigenin-enhanced (Sigma) chemiluminescence as described in detail.24,25 Basal (control) levels of superoxide are reported as the value of tissue plus lucigenin-containing buffer minus background. Superoxide levels were normalized per dry weight. We have shown previously that the signal obtained using this approach is markedly inhibited by scavengers of superoxide (PEG-SOD or Tiron).24,25

![Western blot for CuZn-SOD protein expression in aorta from non-Tg and SOD-Tg mice (left). This Western blot is representative of 4 separate experiments. Total SOD activity (U/mg) (right) expressed as U/mg total protein in aortic homogenates from non-Tg (n=5) and SOD-Tg mice (n=5). Values are mean±SE. *P<0.05 vs non-Tg.](image)

Drugs
U46619 was obtained from Biomol Research Laboratories Inc and dissolved in 100% ethanol. Acetylcholine, nitroprusside, and LPS (E. coli, serotype 026:B6) were obtained from Sigma Chemical Co and dissolved in distilled water. Lucigenin was obtained from Sigma Chemical Co and dissolved in PBS.

Statistical Analysis
Contractions are presented as grams of tension developed and are presented as mean±SEM. Relaxation in response to acetylcholine and nitroprusside are presented as % change in tension from the level of precontraction. When multiple vessel rings were studied from 1 mouse, responses were averaged and n represents the number of mice per group. Comparisons were made using a 1-way ANOVA with repeated measures followed by the Student–Newman–Keuls test to detect individual differences. A P<0.05 was defined as significant.

Results

SOD Activity and CuZn-SOD Protein Expression
Western blotting confirmed that CuZn-SOD protein was present in higher levels in vascular tissue in SOD-Tg mice (Figure 1). Two distinct bands were observed in aorta from SOD-Tg mice. The lower band corresponds to endogenous mouse CuZn-SOD, and the slightly higher band corresponds to human CuZn-SOD (human CuZn-SOD is known to be slightly larger than mouse CuZn-SOD).26 As expected, total SOD activity was higher in aorta from SOD-Tg mice compared with non-Tg mice (Figure 1).

Increases in Vascular Superoxide in Response to LPS Are Prevented in SOD-Tg Mice
Superoxide levels, as measured using lucigenin-enhanced chemiluminescence, were higher in vessels treated with LPS than in those treated with vehicle in non-Tg control mice (Figure 2). In contrast, increases in superoxide in response to LPS were greatly reduced in SOD-Tg mice (Figure 2). These findings indicate that overexpression of CuZn-SOD is effective in preventing large LPS-mediated increases in vascular superoxide.

Vascular Responses in Non-Tg and SOD-Tg Mice
U46619 produced concentration-dependent contraction of carotid arteries. For relaxation studies, arteries were contracted to 60% to 70% of maximum using 0.3 μg/mL U46619. The response to this concentration of U46619 was similar in non-Tg and SOD-Tg mice and was not affected by LPS (P>0.05). Carotid arteries contracted by
0.23±0.02 and 0.20±0.02 g in non-Tg mice treated with vehicle and LPS, respectively. In SOD-Tg mice, carotid arteries contracted by 0.18±0.03 and 0.17±0.04 g following treatment with vehicle and LPS, respectively. Thus, at the concentration used in these studies, vasoconstriction to U46619 was not affected by LPS.

Responses of the carotid artery to acetylcholine under normal conditions were similar in non-Tg mice and SOD-Tg mice (Figure 3). Thus, overexpression of CuZn-SOD per se does not alter endothelial function. In arteries from non-Tg mice, acetylcholine-induced relaxation was markedly inhibited by LPS (Figure 3). For example, in non-Tg mice, relaxation in response to 100 μmol/L acetylcholine was 86±6% and 38±8% in carotid arteries treated with vehicle and LPS, respectively (Figure 3). In contrast to arteries from non-Tg mice, LPS did not significantly affect responses to acetylcholine in vessels from SOD-Tg mice (Figure 4).

In arteries from non-Tg and SOD-Tg mice, nitroprusside produced similar relaxation in vessels treated with vehicle or LPS (Figure 4). Thus, LPS did not inhibit vasorelaxation in response to nitroprusside in either non-Tg or SOD-Tg animals. These findings indicate that impaired responses to acetylcholine following LPS in non-Tg mice were selective for the endothelium-dependent agonist and were not a result of generalized nonspecific alteration in vascular function.

**Discussion**

There are several major findings in this study. First, superoxide levels were increased in vessels from non-Tg control mice following local treatment with LPS. Second, LPS produced marked impairment of vasorelaxation in response to acetylcholine in non-Tg mice. This effect was selective as vascular responses to nitroprusside were not affected by LPS. Third, in contrast to non-Tg animals, increases in vascular superoxide were greatly attenuated and responses to acetylcholine were essentially normal in carotid arteries from SOD-Tg mice following treatment with LPS. These results indicate that LPS increases superoxide and impairs endothelium-dependent relaxation. Overexpression of the CuZn isoform of SOD effectively prevents LPS-induced oxidative stress and endothelial dysfunction.

Vascular inflammation is present, or components of the inflammatory response are activated, within blood vessels with aging and in many cardiovascular diseases including atherosclerosis (carotid artery disease), diabetes, hypertension, and hyperhomocysteinemia. This vascular inflammation is thought to contribute significantly to vascular dysfunction, including endothelial dysfunction, in these disease states.

Many studies have used LPS to activate inflammatory mechanisms, including nuclear factor κB and production of proinflammatory cytokines, in blood vessels. In this experimental model, impairment of endothelium-dependent relaxation is commonly observed. Nitric oxide, which is the major mediator of endothelium-dependent relaxation, reacts with superoxide at a rate 3 times faster than dismutation of superoxide by SOD. LPS and proinflammatory cytokines increase levels of superoxide and peroxynitrite within intact blood vessels and in vascular cells in culture, but the functional importance of these changes has been very difficult to define. For example, treatment with exogenous scavengers of superoxide have had no effect or have worsened endothelial dysfunction in these models. In contrast, high concentrations of vitamin C improved impaired vascular responses to acetylcholine following LPS. An explanation for these divergent results is not clear. There are potential limitations with the use of pharmacological scavengers or exogenous SOD, including cytotoxicity and uncertainties in subcellular access and the degree of scavenging of superoxide. In addition, scavengers of superoxide may have nonspecific effects. Even when an antioxidant has positive effects, as in the case of vitamin C in 1 study, the use of this strictly pharmacological approach provides no insight into subcellu-

**Figure 2.** Superoxide levels (as detected by lucigenin-enhanced chemiluminescence) in aorta from non-Tg (n=13) and SOD-Tg mice (n=6) after treatment with vehicle (control) or LPS. Values are mean±SE. *P<0.05 vs control; †P<0.05 vs SOD-Tg with LPS.

**Figure 3.** Relaxation of carotid arteries from non-Tg mice (n=10, left) and SOD-Tg mice (n=7, right) in response to acetylcholine following treatment with vehicle (control) or LPS. Values are mean±SE. *P<0.05 vs control.

**Figure 4.** Relaxation of carotid arteries from non-Tg mice (n=10, left) and SOD-Tg mice (n=7, right) in response to nitroprusside following treatment with vehicle or LPS. Values are mean±SE.
lar localization of oxidative stress and site of action of superoxide.

The protection by overexpression of CuZn-SOD observed in this study suggests that increases in superoxide in intracellular or cytosolic compartments following LPS are functionally important. This conclusion is consistent with recent work, which suggests that superoxide contributes to intracellular protein tyrosine nitration (via its reaction with nitric oxide and production of peroxynitrite) in vascular muscle during inflammation.\(^{21}\) Potential sources of superoxide in inflammation, such as NAD(P)H oxidase, are known to increase superoxide intracellularly.\(^{18-20}\) Because superoxide is charged and may not easily diffuse across cell membranes, we hypothesized that overexpression of the isoform of SOD that is expressed in close proximity to these intracellular increases in superoxide would be protective in inflammation. Our results are consistent with this hypothesis in that overexpression of CuZn-SOD was very effective in protecting against LPS-induced increases in superoxide and endothelial dysfunction.

In addition to CuZn-SOD, 2 other isoforms of SOD are expressed within the vessel wall: mitochondrial or Mn-SOD (SOD-2), and extracellular or EC-SOD (SOD-3). Within blood vessels, the predominant isoform of SOD is CuZn-SOD, accounting for approximately 50% to 80% of total SOD activity.\(^{24,31,32}\) Constitutive levels of these SODs were not sufficient to prevent increases in superoxide and endothelial dysfunction in response to a high concentration of LPS in vessels from non-Tg (normal) animals (ie, LPS increased superoxide and produced endothelial dysfunction in this and in previous studies). In SOD-Tg mice, levels of CuZn-SOD protein and SOD activity are increased several-fold (present study).\(^{33}\) This level of overexpression of CuZn-SOD was sufficient to prevent LPS-induced vascular dysfunction.

It is worth noting that overexpression of any SOD will not always be beneficial or protective, because higher levels of hydrogen peroxide result from increased expression of SOD and hydrogen peroxide can have deleterious effects in some systems. Thus, although it was possible that vascular dysfunction would not be improved by overexpression of CuZn-SOD, we did not find this to be the case. Endothelial function was not altered by overexpression of CuZn-SOD per se (in the absence of LPS) and endothelium was protected from LPS-induced dysfunction.

In summary, overexpression of CuZn-SOD protects against LPS-induced increases in superoxide and vascular dysfunction. These results complement previous work demonstrating that overexpression of this isoform of SOD attenuates/protects against vasospasm after subarachnoid hemorrhage\(^{34}\) as well as endothelial dysfunction produced by ceramide\(^{35}\) and overexpression of amyloid precursor protein.\(^{36}\) SOD-Tg mice are also protected from increases in vascular superoxide, peroxynitrite, and blood pressure in response to angiotensin II.\(^{37}\) Overall, these studies suggest that overexpression of CuZn-SOD is very effective in attenuating oxidative stress and in attenuating vascular dysfunction in several disease models.

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References


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