Endothelial Nitric Oxide Synthase Mediates Endogenous Protection Against Subarachnoid Hemorrhage-Induced Cerebral Vasospasm

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Background and Purpose—Vasospasm-induced delayed cerebral ischemia remains a major source of morbidity in patients with aneurysmal subarachnoid hemorrhage (SAH). We hypothesized that activating innate neurovascular protective mechanisms by preconditioning (PC) may represent a novel therapeutic approach against SAH-induced vasospasm and neurological deficits and, secondarily, that the neurovascular protection it provides is mediated by endothelial nitric oxide synthase (eNOS).

Methods—Wild-type mice were subjected to hypoxic PC or normoxia followed 24 hours later by SAH. Neurological function was analyzed daily; vasospasm was assessed on post-surgery Day 2. Nitric oxide availability, eNOS expression, and eNOS activity were also assessed. In a separate experiment, wild-type and eNOS-null mice were subjected to hypoxic PC or normoxia followed by SAH and assessed for vasospasm and neurological deficits.

Results—PC nearly completely prevented SAH-induced vasospasm and neurological deficits. It also prevented SAH-induced reduction in nitric oxide availability and increased eNOS activity in mice with and without SAH. PC-induced protection against vasospasm and neurological deficits was lost in wild-type mice treated with the nitric oxide synthase inhibitor $\text{N}^\text{G}$-nitro-L-arginine methyl ester and in eNOS-null mice.

Conclusions—Endogenous protective mechanisms against vasospasm exist, are powerful, and can be induced by PC. eNOS-derived nitric oxide is a critical mediator of PC-induced neurovascular protection. These data provide strong “proof-of-principle” evidence that PC represents a promising new strategy to reduce vasospasm and delayed cerebral ischemia after SAH. (Stroke. 2011;42:776-782.)

Key Words: cerebral vasospasm ■ hypoxia ■ nitric oxide synthase ■ preconditioning ■ subarachnoid hemorrhage

Preconditioning (PC) refers to a strategy whereby exposure to a sublethal quantity of a normally injurious stimulus can be used to augment the brain’s inherent evolutionarily conserved protective mechanisms to attenuate subsequent injury. Although the contribution of neurons to the observed protection is well documented, the role of the cerebrovasculature (vascular PC) has only recently been appreciated (for review, see Gidday1). Most studies examining PC have used physiological stimuli such as hypoxia, ischemia, or lipopolysaccharide administration in the setting of brain ischemia.1,2 Although these studies have provided critical insight into the underlying mechanisms of endogenous brain protection, it appears that clinical application of a PC strategy may be limited to select patient populations with especially high risk for ischemic events,2 including those undergoing carotid endarterectomy, cerebral aneurysm surgery, and coronary artery bypass grafting.3 Another potential population is aneurysmal subarachnoid hemorrhage (SAH), although preclinical studies examining this possibility are currently lacking.

SAH is a disease associated with high morbidity and mortality,4 a significant portion of which is due to the development of delayed cerebral ischemia. This condition may be ideal for a PC strategy because (1) there is a wide therapeutic window between SAH and delayed cerebral ischemia (typically 4 to 12 days)5; (2) vasospasm is a key factor responsible for delayed cerebral ischemia,4 and PC stimuli have known vasoprotective effects6–11; and (3) endothelial nitric oxide synthase (eNOS)-derived nitric oxide (NO), a known mediator of vascular PC,2,7,12 is strongly linked to SAH-induced vasospasm.13 Therefore, in this “proof-of-principle” study, we sought to determine whether PC results in protection against SAH-induced vasospasm and neurological dysfunction and...
whether eNOS-derived NO contributes to the mechanism underlying this protective effect.

## Materials and Methods

### Animals
All experimental protocols were approved by the Animal Studies Committee at Washington University in St Louis. Three to four months old male wild-type mice (C57BL/6J) and homozygous eNOS<sup>−/−</sup> mice (B6.129P2-Nos3<sup>tm1Unc</sup>/J with a C57BL/6 genetic background) from Jackson Laboratories (Bar Harbor, ME) were used.

### Hypoxic PC
Mice were placed in a hypoxic chamber and exposed to air containing 8% O<sub>2</sub>/92% N<sub>2</sub> for 4 hours with access to food and water ad libitum<sup>11</sup>; mice were then returned to their cages. Normoxic controls were placed in chambers containing room air.

### Pharmacological Inhibition of NO Synthase
To examine the effect of NO synthase inhibition on PC-induced neurovascular protection in SAH, wild-type mice were administered the pan-NO synthase inhibitor L-NG-nitroarginine methyl ester (L-NAME; 20 mg/kg intraperitoneally) or vehicle (normal saline) starting 1 hour before PC and continued once daily thereafter.

### Experimental Protocols and Groups
The first set of experiments was performed to assess whether hypoxic PC attenuates SAH-induced vasospasm and neurological deficits. Three groups of mice were used: (1) normoxia followed 24 hours later by sham surgery (Norm:Sham), n=11; (2) normoxia followed 24 hours later by SAH (Norm:SAH), n=16; and (3) PC followed 24 hours later by SAH (PC:SAH), n=13. The second set of experiments was designed to explore the molecular pathways affected by PC and SAH. Four groups of mice were used: (1) Norm:Sham, n=6; (2) PC followed 24 hours later by sham surgery (PC:Sham), n=5; (3) Norm:SAH, n=10; and (4) PC:SAH, n=8. The third set of experiments was designed to examine whether pharmacological inhibition NO synthase blocks the neurovascular protective effects of PC in SAH. Six groups were used: (1) Vehicle:Norm:Sham, n=3; (2) Vehicle:Norm:SAH, n=5; (3) Vehicle:PC:SAH, n=5; (4) L-NAME:Norm:Sham, n=5; (5) L-NAME:Norm:SAH, n=11; and (6) L-NAME:PC:SAH, n=11. For statistical power, data from the vehicle-treated groups were combined with data from Experiment 1 once it was determined that the corresponding groups were statistically similar (data not shown). The fourth set of experiments was designed to directly assess the contribution of eNOS in PC-induced neurovascular protection. Six groups of mice were used: (1) eNOS<sup>−/−</sup>:Norm:Sham, n=14; (2) eNOS<sup>+/−</sup>:Norm:Sham, n=12; (3) eNOS<sup>−/−</sup>:PC:SAH, n=17; (4) eNOS<sup>−/−</sup>:Norm:SAH, n=19; and (6) eNOS<sup>−/−</sup>:PC:SAH, n=20. See Figure 1 for detailed experimental protocols. All assessments were performed by investigators blinded to the surgical procedure and genotypes.

### Experimental SAH
Endovascular perforation SAH was performed as described<sup>15</sup>. Briefly, mice were anesthetized with isoflurane (4% induction, 1.5% maintenance), and a 5-0 nylon suture was introduced into the external carotid artery and advanced through the internal carotid artery until the internal carotid artery bifurcation. The suture was then advanced to induce SAH, then removed and the external carotid artery ligated. Mice in the sham surgery groups underwent these procedures except for suture perforation.

### Neurobehavioral Tests
Neurobehavioral outcome was examined daily using Neuroscore and Rotarod tests as described<sup>15,18</sup>. Briefly, neurological function was graded based on a motor score (0 to 12) that evaluated spontaneous activity, symmetry of limb movements, climbing, balance, and coordination and a sensory score (4 to 12) that evaluated body proprioception and vibrissae, visual, and tactile responses. Balance and coordination were assessed by performance on the Rotarod (Rotamex-5; Columbus Instruments, Columbus, OH). Mice were pretrained on the Rotarod 1 day before surgery. Latency on 3 trials of 180 seconds was averaged daily.

### Vasospasm Assessment
Vasospasm assessment was performed on post-surgery Day 2 through cerebrovascular casting, as described<sup>15</sup>. Briefly, mice were anesthetized and transcardially perfused with phosphate-buffered saline, 10% formalin, and 3% gelatin–India ink solution. Brains were removed, SAH was graded as described<sup>15</sup>, and blood vessels imaged under a microscope using a charge-coupled device camera. The narrowest diameter within the first 1000 μm of the middle cerebral artery was measured to assess vasospasm.

### NO Availability Assay
NO availability was determined using DAF-2DA (fluorophore 4,5-diaminofluorescein-2-diacetate; EMD Chemicals, Gibbstown, NJ) as described<sup>17</sup>. Briefly, brain lysates were incubated with 50 μmol/L DAF-2DA for 30 minutes at 22°C in the dark. The reaction mixture was excited at 495 nm and emission read at 515 nm in a spectrophotometer (Biotek, Winooski, VT).

### Western Blot
Western blots were performed as described<sup>18</sup>. Briefly, brains were harvested 48 hours after sham or SAH surgery. Tissue from the hemisphere ipsilateral to endovascular perforation was lysed in a buffer containing 10 mmol/L 4-(2-hydroxyethyl)-1-piperazine-ethanesulfonic acid, 5 mmol/L MgCl<sub>2</sub>, 1 mmol/L dithiothreitol, 2 mmol/L ethylenediaminetetra-acetic acid, 2 mmol/L ethylene glycol bis(β-aminoethyl ether)-N,N,N′,N′-tetra-acetic acid, 1 mmol/L phenylmethylsulfonyl fluoride, 1% Triton X-100, 0.5 mmol/L sodium orthovanadate, 0.1 μmol/L okadaic acid, 25 mmol/L β-glycerophosphate, and protease inhibitor cocktail.

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**Figure 1.** Experimental protocol. Arrows indicate time points for PC, surgical procedures, and functional, morphological, and biochemical analyses.
The following primary antibodies were used: mouse anti-eNOS (BD Biosciences), rabbit anti-inducible nitric oxide (iNOS; Santa Cruz Biotechnology), mouse anti-neuronal nitric oxide synthase (nNOS; Santa Cruz Biotechnology), and mouse anti-α-tubulin (Sigma). Blots were subsequently incubated with anti-mouse or anti-rabbit horseradish peroxidase-conjugated IgG and visualized using an enhanced chemiluminescence kit (BioRad).

### eNOS Activity Assay

NO synthase activity assay was performed as described. Briefly, brain tissue ipsilateral to endovascular perforation was lysed in a buffer containing 20 mmol/L 4-(2-hydroxyethyl)-1-piperazine-ethanesulfonic acid, pH 7.4, 320 mmol/L sucrose, 1 mmol/L dithiothreitol, and protease inhibitor cocktail. Brain lysates were incubated for 30 minutes in an assay buffer containing 20 mmol/L 4-(2-hydroxyethyl)-1-piperazine-ethanesulfonic acid, pH 7.4, 60 μmol/L \[^{[3]H}\]-l-arginine 1 μCi/mL, 1 mmol/L ethylenediaminetetra-acetic acid, 1.2 mmol/L CaCl\(_2\), and 2.5 mmol/L β-reduced nicotinamide adenine dinucleotide phosphate. Samples were also incubated in this mixture along with 10 μmol/L L-NG-monomethyl arginine (competitive inhibitor of all NO synthase isoforms) or 10 μmol/L 1-(2-trifluoromethylphenyl) imidazole (selective inhibitor of nNOS and iNOS). Protein was removed by centrifugation and supernatant was passed through Dowex50WX-8 columns (Na\(^+\) form) and eluted. The radioactivity of \[^{[3]H}\]-citrulline was measured within eluates by scintillation spectrometry (Packard Instrument Company, Meriden, CT). The activity in the mixture containing L-NG-monomethyl arginine was considered background and subtracted from the activity of the mixture containing 1-(2-trifluoromethylphenyl) imidazole to obtain activity specific to eNOS.

### Statistical Analyses

Data are presented as the mean±SEM and were analyzed by analysis of variance followed by Newman-Keuls multiple comparison method unless otherwise indicated. \(P<0.05\) was considered statistically significant.

### Results

#### PC Attenuates SAH-Induced Vasospasm

Significant vasospasm (27%±6% vessel constriction, \(P<0.05\)) was noted 2 days after SAH in nonpreconditioned mice (Figure 2). In contrast, preconditioned mice did not develop significant vasospasm (7%±6% vessel constriction, \(P>0.05\); Figure 2). Hemorrhage grade (nonpreconditioned=3.9, preconditioned=3.7) and mortality (nonpreconditioned=5.8%, preconditioned=7.1%) were not significantly different between SAH groups.

#### PC Improves Neurological Outcome After SAH

SAH led to significant neurobehavioral deficits by Neuroscore and Rotarod latency test (\(P<0.05\)) in nonpreconditioned mice (Figure 3). These neurobehavioral deficits were significantly attenuated in preconditioned mice (\(P<0.05\); Figure 3).

#### PC Attenuates SAH-Induced Reduction in NO Availability

NO availability was assessed 72 hours after PC (48 hours after sham or SAH surgery). SAH reduced NO availability by 42%±20% (\(P<0.05\)) in nonpreconditioned mice (Figure 4). However, in preconditioned mice, NO availability was unchanged after SAH (ie, SAH-induced reduction in NO availability was prevented; Figure 4). PC also increased NO availability in sham-operated mice by 69%±34% (\(P<0.05\)).

### NO Synthase Inhibition Blocks PC-Induced Neurovascular Protection in SAH

NO synthase inhibition with L-NAME was found to (1) reduce baseline middle cerebral artery diameter by 12%±4% (\(P=0.03\); (2) have no impact on the degree of vasospasm after SAH (29%±5% vessel constriction in vehicle-treated versus 32%±8% vessel constriction in L-NAME-treated nonpreconditioned mice, \(P=0.7\)); and (3) completely block PC-induced protection against vasospasm (66%±15% vasospasm protection in vehicle-treated versus −25%±23% vasospasm protection in L-NAME-
sham or SAH surgery). PC increased eNOS activity by 1.4-fold (\(P<0.05\)) in SAH-operated mice (Figure 5). SAH alone did not alter eNOS activity (Figure 5).

### eNOS Contributes to PC-Induced Protection Against Vasospasm and Neurological Deficits After SAH

To directly assess the role of eNOS in PC-induced neurovascular protection, a separate group of wild-type (eNOS\(^{+/+}\)) and eNOS-null (eNOS\(^{-/-}\)) mice was examined. SAH caused significant (\(P<0.05\)) vasospasm in both eNOS\(^{+/+}\) (21\%\(\pm\)5\% constriction) and eNOS\(^{-/-}\) (24\%\(\pm\)5\% constriction) mice (Figure 6A). As expected, SAH-induced vasospasm was nearly completely prevented (94\%\(\pm\)19\% vasospasm reduction; \(P<0.05\)) in preconditioned eNOS\(^{+/+}\) mice (Figure 6A–B). However, vasospasm protection by preconditioning was largely absent (24\%\(\pm\)20\% vasospasm reduction) in eNOS\(^{-/-}\) mice (Figure 6A–B). A commensurate loss in PC-induced neurobehavioral protection (by Neuroscore) was also found in eNOS\(^{-/-}\) mice (Figure 6C). Rotarod testing was not performed because naïve eNOS\(^{-/-}\) mice had substantial Rotarod deficits at baseline (data not shown). Hemorrhage grade (eNOS\(^{+/+}\) nonconditioned mice=3.2; eNOS\(^{-/-}\) preconditioned mice=3.1; and eNOS\(^{-/-}\) preconditioned mice=3.7) and mortality (eNOS\(^{+/+}\) nonconditioned mice=6.7%; eNOS\(^{-/-}\) preconditioned mice=10.3%; eNOS\(^{-/-}\) nonpreconditioned mice=5.0%; and eNOS\(^{-/-}\) preconditioned mice=4.7%) were not significantly different between SAH groups.

**Figure 4.** NO synthase-derived NO mediates PC-induced protection against neurovascular dysfunction after SAH. Mice underwent hypoxic PC or normoxia (Norm) followed 24 hours later by SAH or sham surgery. NO availability was quantified by fluorometric detection of DAF-2DA on brain homogenates obtained on post-surgery Day 2 (A). Data indicate mean\(\pm\)SEM. *\(P<0.05\) vs Norm:Sham, \#\(P<0.05\) vs Norm:SAH. In a separate experiment, mice were administered the pan-NOS inhibitor L-NAME and subjected 1 hour later to hypoxic PC or normoxia (Norm) followed 24 hours later by SAH or sham surgery. Vasospasm was assessed in the middle cerebral artery on post-surgery Day 2 by the gelatin–India ink casting method (B). *\(P<0.05\), n.s. \(P>0.05\) by analysis of variance and Newman-Keuls multiple comparison test; neurobehavioral assessment was performed on Days 0 to 2 by Neuroscore (C) and Rotarod latency (D) tests. *\(P<0.05\) vs Norm:Sham. Data indicate mean\(\pm\)SEM.

**Figure 5.** PC increases eNOS expression and activity. Mice underwent hypoxic PC or normoxia (Norm) followed 24 hours later by SAH or sham surgery. eNOS, nNOS, and iNOS expression was assessed by Western blot and eNOS activity was assessed by [\(^{3}\)H]L-arginine to [\(^{3}\)H]L-citrulline conversion, both performed on brain homogenates obtained on post-surgery Day 2. (A) Representative immunoblot images (N indicates normoxia); (B) quantification data of eNOS, nNOS, and iNOS expression normalized to \(\alpha\)-tubulin; and (C) quantitative data of eNOS activity. Data indicate mean\(\pm\)SEM (B) *\(P<0.05\), n.s. \(P>0.05\) by Student’s t test; (C) *\(P<0.05\) vs Norm:Sham, \#\(P<0.05\) vs Norm:SAH by analysis of variance and Newman-Keuls multiple comparison test.

**Figure 6A–B.** Hemorrhage grade (eNOS\(^{+/+}\) nonconditioned mice=3.2; eNOS\(^{-/-}\) preconditioned mice=3.1; and eNOS\(^{-/-}\) preconditioned mice=3.7) and mortality (eNOS\(^{+/+}\) nonconditioned mice=6.7%; eNOS\(^{-/-}\) preconditioned mice=10.3%; eNOS\(^{-/-}\) nonpreconditioned mice=5.0%; and eNOS\(^{-/-}\) preconditioned mice=4.7%) were not significantly different between SAH groups.
ies,1,2,13 although the latter has not been previously reported.

...operation mice. Both results are consistent with past stud-
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...onding that PC increased NO levels in both sham- and SAH-operated mice. Both results are consistent with past stud-
...y, provides support to the evolving concept of vascular PC
...
...is due, in part, to decreased vasospasm, a well-recognized risk

Figure 6. PC-induced protection against vasospasm and neuro-

Discussion

In the present study, we established the existence of endog-

neurological protection afforded by PC in the setting of SAH.

With respect to the molecular mechanism underlying

PC-induced vasospasm protection, we examined NO given its

result is similar to that reported in studies examining PC in

the setting of cerebral ischemia7,12,21 and neonatal hypoxia–

ischemia.9,20 Collectively, these data strongly suggest that NO

secondary to a persistent upregulation in eNOS. Our findings in eNOS-null mice provided further support

for this hypothesis. That PC-induced abrogation of vaso-

spasm and neurobehavioral deficits in wild-type mice was

lost in eNOS-null mice (although the degree of vasospasm

and neurological deficits after SAH was similar between the

2 groups when not preconditioned) indicates that the innate

protective mechanisms by which PC reduces SAH-induced

neurovascular deficits are dependent, to a significant extent,

eNOS. More broadly for the field of PC, our results lend

additional support to the notion that eNOS is a key contri-

butor to PC-induced neuroprotection. Other studies of PC-

induced vasculoprotection in focal ischemic stroke have

advanced similar conclusions regarding the participation of
eNOS.7,22

The effect of SAH on eNOS expression and/or activity is

controversial. We found no effect of SAH on either metric,

but previous studies report no change,23–25 a reduction,26 or an

increase17 in eNOS expression after SAH. Similarly, SAH

was found to either have no effect24,27 on phosphorylated

eNOS (at Ser1177, one of many active forms of eNOS77) or an

increase.17,25,26 In addition, our experiments identified

what could be considered a discrepancy between eNOS

activity (no change) and NO availability (decreased) in

response to SAH. Several possibilities could explain this

observation. First, it is possible that a SAH-induced decrease

in eNOS activity actually occurred in vivo but was undetected

by our ex vivo assay. This could occur if availability of one

or more substrates (eg, L-arginine) or cofactors (eg, reduced

nicotinamide-adenine dinucleotide phosphate), which were

artificially provided in our ex vivo assay, became limited; or

if the endogenous NO synthase inhibitor asymmetrical di-

methyl L-arginine was upregulated by SAH.13 Second, it is

possible that NO availability decreased after SAH despite

unchanged eNOS activity. For example, this could occur

through NO scavenging by oxyhemoglobin28 and/or

leukocytes29 present in the subarachnoid space. In either case,

PC-induced changes appear significant enough to overwhelm

these factors and restore NO production to pre-SAH levels.
Some limitations of our study should be noted. First, PC was administered before SAH. This was by design, because our study was conceived as a “proof-of-concept” one to establish whether endogenous protective mechanisms against vasospasm are present and thus permit their exploration. Studies examining the efficacy of post-SAH conditioning paradigms have already been initiated in our laboratory. Second, although the clinical applicability of hypoxia as a preconditioning stimulus may be difficult to envision, several pharmacological agents with known PC effects are available for future testing. Third, because no animal model fully recapitulates human SAH, it is essential to validate our findings in complementary SAH models. Fourth, although vasospasm is a major contributor to delayed cerebral ischemia, other factors may also contribute (for review, see MacDonald et al11); whether PC also positively affects these factors will require future study. Fifth, our study does not exclude a contribution from nNOS or iNOS; however, it is very unlikely that either plays a major role given that neither was upregulated after PC in the setting of SAH. Finally, we recognize that the neurological deficits documented in our SAH model are likely multifactorial and that the observed neuroprotection could be explained in part by beneficial effects of PC on reducing neuronal apoptosis,32 inflammatory pathways that prevent vasospasm, and to examine the therapeutic potential of postconditioning strategies for SAH, are warranted.

In conclusion, results from this “proof-of-principle” study indicate that powerful endogenous protective mechanisms against vasospasm exist, that they can be activated by a preconditioning stimulus, and that their underlying mechanism is at least partly dependent on eNOS-derived NO. Further studies to more fully elucidate the innate molecular pathways that prevent vasospasm, and to examine the therapeutic potential of postconditioning strategies for SAH, are warranted.

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Disclosures
None.

References
6. Dirnagl U, Becker K, Meisel A. Preconditioning and tolerance against vasospasm exist, that they can be activated by a preconditioning stimulus may be difficult to envision, several pharmacological agents with known PC effects are available for future testing. Third, because no animal model fully recapitulates human SAH, it is essential to validate our findings in complementary SAH models. Fourth, although vasospasm is a major contributor to delayed cerebral ischemia, other factors may also contribute (for review, see MacDonald et al11); whether PC also positively affects these factors will require future study. Fifth, our study does not exclude a contribution from nNOS or iNOS; however, it is very unlikely that either plays a major role given that neither was upregulated after PC in the setting of SAH. Finally, we recognize that the neurological deficits documented in our SAH model are likely multifactorial and that the observed neuroprotection could be explained in part by beneficial effects of PC on reducing neuronal apoptosis,32 inflammation,33 and/or blood–brain barrier breakdown.34 In fact, the robustness of the PC-induced protection noted in our study suggests a multifaceted underlying mechanism; therapeutic strategies for SAH that target several deleterious cascades would likely have great potential for clinical efficacy.

In conclusion, results from this “proof-of-principle” study indicate that powerful endogenous protective mechanisms against vasospasm exist, that they can be activated by a preconditioning stimulus, and that their underlying mechanism is at least partly dependent on eNOS-derived NO. Further studies to more fully elucidate the innate molecular pathways that prevent vasospasm, and to examine the therapeutic potential of postconditioning strategies for SAH, are warranted.


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