Changes of pH and Energy State in Subacute Human Ischemia Assessed by Multinuclear Magnetic Resonance Spectroscopy

Johann Philipp Zöllner; Elke Hattingen, MD; Oliver C. Singer, MD*; Ulrich Pilatus, PhD*

Background and Purpose—In vivo changes in tissue pH and energy metabolism are key to understanding stroke pathophysiology. Our goal was to study pH changes in subacute ischemic stroke and their relation to energy metabolism, which, unlike acidosis in acute stroke, are not yet well understood.

Methods—We measured tissue pH and phospholipid as well as cell energy markers, including creatine, phosphocreatine, and N-acetyl-aspartate in subacute stroke with combined 1H and 31P magnetic resonance spectroscopy. We included 19 patients with first-ever ischemic stroke (mean time after stroke, 6 days). We then compared metabolite concentrations in the ischemic tissue to contralateral (healthy) tissue using multivariate ANOVA to assess significant differences in metabolite levels between both tissue compartments.

Results—In subacute stroke, a tissue fraction with significantly increased tissue pH was observed as compared with healthy contralateral tissue (pH, 7.09 versus 7.03; P=0.002) concurrent with splitting of the pH signal with 1 peak being more alkalotic. Furthermore, only a moderate decrease of energy-rich metabolites (phosphocreatine reduced by 17%, ATP reduced by 19%) was present, whereas total creatine was reduced by 51%.

Conclusions—The finding of an alkalotic pH split in subacute ischemia is unprecedented. The pH split and only incomplete energy loss in subacute stroke suggest 2 differently viable cellular moieties, best explained by active compensatory mechanisms after acute cerebral ischemia. (Stroke. 2015;46:441-446. DOI: 10.1161/STROKEAHA.114.007896.)

Key Words: energy metabolism ■ lactate ■ magnetic resonance spectroscopy ■ pH

Ischemic stroke is characterized by several key metabolic changes. Among them are changes in cell energy status, membrane lipid metabolism, and tissue pH. These changes reflect processes of cell degradation, excitotoxicity, and inflammation. Commonly, an acidotic milieu is assumed in acute ischemic stroke because of a switch from aerobic to anaerobic glycolysis. This results in a reduction of tissue pH by accumulation of lactate. Several magnetic resonance spectroscopy (MRS) studies have shown acidotic intracellular pH values in acute ischemia.1,2 However, a reversal from acute acidosis to subacute alkalosis has been described in longitudinal studies using MRS.3,4

Acute ischemia is expected to show a near complete depletion of energy substrates in the center of the infarct as reflected in drastically reduced levels of phosphocreatine, ATP, and increased levels of inorganic phosphate (Pi).1 Phosphocreatine is an important energy source to regenerate ATP from ADP by creatine kinase reaction. Owing to the breakdown of energy metabolism, the vast majority of total creatine (tCr) should, therefore, be unphosphorylated creatine (uCr). N-acetylaspartate (NAA), an amino acid exclusively produced in neuronal mitochondria, should be greatly reduced, as severe neuronal damage is supposed to appear in infarcted tissue. These changes in cell energy status can be assumed to persist well into the subacute phase of stroke.

Combined 1H and 31P magnetic resonance spectroscopic imaging (MRSI) at 3 Tesla is able to measure important metabolites of the above-mentioned processes in vivo. We, therefore, aimed to measure the pathophysiological fate of neuronal and glial tissue in 19 first-ever stroke patients focusing on alterations of tissue pH and energy metabolism, especially concerning the as of yet incompletely described fate of ischemic tissue in the subacute stage.

Materials and Methods

Subjects

We studied 19 adult patients suffering a first-ever ischemic stroke. The diagnosis of ischemic stroke was established clinically and by standard computed tomography or MRI. Patients were studied in the subacute phase of stroke (3 days to 2 weeks after stroke onset). Patients who had previously suffered (clinically evident or silent) strokes were excluded, as were those with a positive medical history of other brain pathologies. Further exclusion criteria were cerebral microangiopathy or hemorrhagic transformation of infarcted tissue. Patients with contraindications for MRI examination were excluded.
as well. The study was approved by the ethics committee of the medical faculty of the Goethe University, Frankfurt. Each patient signed an informed consent form before the examination.

**MRS Imaging**

Magnetic resonance examinations were performed on a 3.0 T whole-body scanner (Siemens TRIO, Siemens AG, Erlangen, Germany). A $^{1}$H/$^{31}$P double-tuned head-coil (Rapid Biomedical, Würzburg, Germany) was used to obtain anatomic and spectroscopic data with the same equipment (ie, no repositioning required during scanning). We acquired axially oriented T2w anatomic images (slice thickness, 5 mm) for stroke localization aligned at the anterior–posterior commissure-axis to facilitate congruence between images. At the end of the magnetic resonance examination, a 3D T1-sequence (three-dimensional magnetization-prepared rapid gradient-echo imaging) was recorded for tissue segmentation into white matter, gray matter, and cerebrospinal fluid.

For in vivo MRS, MRSI sequences were used. The $^{1}$H data were obtained using an axial 2D-MRSI sequence with a slice thickness of 12 mm. The volume of interest was chosen to include infarcted tissue, as well as the respective contralateral tissue, which served as control region. We used point-resolved selective spectroscopy combined with outer volume suppression, applying an acquisition-weighted circular phase-encoding scheme on a 24×24 matrix, FOV of 240×240 mm, nominal voxel size of 10×10×12 mm, repetition time=1500 ms, echo time=144 ms, and 2 acquisitions. For $^{31}$P MRS, a three-dimensional MRSI sequence with WALTZ4 proton decoupling was used. The signal was acquired as free induction decay applying a 60° flip angle (repetition time=2000 ms; echo time=2.3 ms; 10 acquisitions). An axial slab of 100 mm aligned to the axial $^{1}$H MRSI slice was recorded, using elliptic phase encoding with a weighted acquisition scheme on a 10×10×8 matrix, FOV of 300×300×200 mm, and nominal voxel size of 30×30×25 mm. For all modalities, slice (slab) angularizations were identical. The total examination time was ≈45 minutes.

**Data Processing**

Spectroscopic data were evaluated offline on a Linux workstation, using MATLAB (The MathWorks, Inc., Natick, MA) scripts. The MRSI matrix was extrapolated before Fourier transformation to 48×48 voxels for $^{1}$H data (resulting in a 5×5 mm in-plane grid size) and to 20×20×16 voxels for $^{31}$P data (resulting in a series of axial slices with a thickness of 12.5 mm and 15×15 mm in-plane grid size). Voxel selection was based on the extrapolated grid.

For the $^{1}$H data evaluation, several voxels of interest were placed in the infarcted area. Voxel selection was guided by T2w images, avoiding any partial volume from unaffected tissue. Control voxels were placed in the corresponding contralateral area, mirrored at the midline. The total amount of voxels per patient (mean value of 12 voxels per patient, range: 2–40) was depended on infarct size and position of the volume of interest. Spectra from selected voxels were analyzed with the software LCModel (V6.1-4E, S. Provencher, http://lspm-proverchen.com), which performs a nonlinear analysis of the data in the frequency domain using a linear combination of model spectra. Our set of model spectra was restricted to the metabolites NAA, lactate, creatine (tCr), total choline (tCho), myo-inositol, and alanine (Ala), but only NAA, tCr, and tCho were detected with Cramer-Rao lower bounds (SD values given by the tool) of <20%. Voxels were averaged for further processing whenever >1 voxel had been selected. Lactate was evaluated separately by examining individual voxels to distinguish a possible lipid signal overlap. If a clear negative doublet at 1.3 ppm was discernible and SD was <25%, the voxel was assumed to contain lactate.

For the $^{31}$P data evaluation, the grid was automatically shifted in the axial direction to place the $^{1}$H slice at the center of a $^{31}$P slice. To minimize partial volume effects from less affected tissue, an in-plane grid-shifting was applied manually positioning at least one voxel in T2w-positive infarcted tissue. Only 1 voxel in the center was included in the analysis, which was performed with the software jMRUI (version 4.0; http://www.mrui.uab.es/mrui). We used the AMARES algorithm, which applies a nonlinear-least-square fit of the metabolites in the frequency domain. The algorithm can handle constraints based on previous knowledge. In our case, the fit was restricted to the metabolites ATP, phosphocholine, phosphoethanolamine, phosphocreatine, glycero-phosphocholine, glycerophosphoethanolamine, and Pi.

Figure 1 shows example of $^{31}$P and $^{1}$H magnetic resonance spectra in 1 patient with left middle cerebral artery stroke.

Tissue pH calculation was performed by 2 different approaches. After the procedure described by Levine et al, the frequency area for signals from inorganic phosphate at 3.6 to 5.6 ppm relative to phosphocreatine at 0 ppm was analyzed by a single Lorentzian line with variable line width, frequency, and intensity (single-peak approach). The position of this peak was used to establish a general pH value. In all but 1 case (with the smallest infarct size) the improved spectral resolution at 3 T (compared with the study at 1.5 T from Levine et al.) allowed a clear distinction of non-Lorentzian signal broadening in the area between 4 and 6 ppm as it is shown in Figure 2. Assuming that the broadening is caused by an additional Pi component reflecting infarcted tissue, we analyzed the area separately (double-peak approach). For this, we used 2 signals, Pi(1) and Pi(2). Pi(1) was fixed at the position of inorganic phosphate from healthy tissue (4.82 ppm), whereas the position of Pi(2) was variable between 4.85 and 5.5 ppm. Both signals had a fixed line width of 12 Hz. Resulting signal intensities and estimated pH values for Pi(1) and Pi(2) are plotted in Figure 3 against the stroke volume. For both approaches, the tissue pH was calculated from the shift of the relative Pi peaks using the formula by Petroff et al., which is implemented in jMRUI.

Stroke lesion size was calculated on the basis of T2w images using the software ImageJ (http://rsbweb.nih.gov/ij/). Every slice was evaluated for visible ischemic lesions. All T2w positive areas were quantified using the polygon selection tool and the total area was then multiplied with the slice thickness.

Segmented images of white matter, gray matter, and cerebrospinal fluid were obtained using the segmentation algorithm implemented in the Statistical Parametric Mapping (SPM)-8 (Wellcome Department of Imaging Neuroscience, University College of London, United Kingdom; http://www.fil.ion.ucl.ac.uk/spm) tool as described previously. The relative amount of gray matter, white matter, and cerebrospinal fluid contributing to the individual voxel was calculated using MATLAB scripts.
Calibration of MRSI spectra was performed as described by Hattingen et al. Concentrations for 1H detectable metabolites were calculated by referring to an independent measurement of a spherical phantom containing an aqueous solution of 100 mmol/L acetate as a calibration standard. A repetition time of 10 s was applied to avoid T1 saturation. For 31P data, a spherical phantom with 20 mmol/L phosphate was used (repetition time, 60 s). Calibrated signal intensities were divided by the sum of gray matter and white matter fractions in a given voxel to correct for cerebrospinal fluid. This method does not take into account B1 inhomogeneities, but additional measurements performed by our group showed that the inhomogeneities are small compared with differences between healthy and affected tissue. Therefore, we are confident that the obtained values provide a reasonable estimate for absolute concentrations in mmol/L.

uCr concentrations were calculated by the formula $\text{[uCr]} = \text{[creatinine]} - \text{[phosphocreatine]}$.

### Statistical Analysis

Results were statistically evaluated using the software STATISTICA (version 7.1, StatSoft, Tulsa, OK). Multivariate ANOVA was applied to analyze significant differences between stroke and control area considering data from different regions as repetitive measurements. The different metabolite groups ($^{31}$P and pH, $^1$H, combined data) were analyzed separately. Significance for each metabolite was tested using contrast analysis in ANOVA. For correlation analysis, the Pearson correlation coefficient was used.

### Patients

Clinical data of the 19 patients are summarized in Table 1. Mean age at stroke onset was 56 years (SD, 16; range, 27–82 years), 26% were female patients. Seventeen patients had a stroke within the territory of the middle cerebral artery, 2 within the posterior cerebral artery territory. Mean infarct volume was 53.9 cm$^3$ (SD, 54.3; range, 1.3–217.8 cm$^3$). The mean time elapsed between stroke onset and MRSI was 6 days (SD, 2.8; range, 3–12 days).

### Tissue pH and Lactate Concentrations

For the pH value obtained from the signal position using the single-peak approach, we found a significantly increased pH value in the infarct core relative to healthy tissue (7.09±0.07 versus 7.03±0.02; $P=0.002$; Table 2). Using this single-peak approach, a strong correlation between lesion volume and pH increase ($r=0.91$; $P<0.0001$) was observed, suggesting that larger strokes

### Table 1. Patient Characteristics

<table>
<thead>
<tr>
<th>Patient</th>
<th>Age, y</th>
<th>Sex</th>
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Mean 56 n.a. n.a. n.a. 6 53.9
SD 16 n.a. n.a. n.a. 2.8 54.3
Range 27–82 n.a. n.a. n.a. 3–12 1.3–217.8

F indicates female; M, male; MCA, middle cerebral artery; n.a., not applicable; and PCA, posterior cerebral artery.
Table 2. Quantitative Measurements of pH and Markers of Energy- and Phospholipid-Metabolism Within Infarcted and Healthy Tissue

<table>
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<th>Stroke Mean±SD</th>
<th>Control Mean±SD</th>
<th>P Value</th>
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<td>7.03±0.02</td>
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<td>Linewidth Pi (Hz)</td>
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<td>12.6±7.0</td>
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<tr>
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<td>7.6±3.1</td>
<td>0.60</td>
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<tr>
<td>pH (double peak approach)</td>
<td>7.37±0.28*</td>
<td>Not applicable†</td>
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Markers of energy metabolism

- NAA: 3.65±2.85 versus 10.47±1.75; <0.001
- tCr: 4.00±2.1 versus 8.24±1.84; <0.001
- Phosphocreatine: 3.07±0.60 versus 3.70±0.62; <0.001
- Unphosphorylated choline: 0.99±1.79 versus 4.66±1.77; <0.001
- ATP: 1.40±0.46 versus 1.73±0.31; <0.001
- Pi: 1.59±0.55 versus 1.31±0.42; 0.06
- Lactate: 2.22±2.09 versus 0.97±0.77; 0.04

Markers of phospholipid metabolism

- tCho: 1.42±0.63 versus 2.15±0.40; 0.007
- Glycero-phosphocholine: 1.26±0.27 versus 1.51±0.28; 0.002
- Glycero-phosphoethanolamine: 1.08±0.42 versus 1.31±0.38; 0.03
- Phosphoethanolamine: 1.01±0.23 versus 1.13±0.30; 0.03
- Phosphocholine: 0.29±0.11 versus 0.30±0.12; 0.67

Values are given as mmol/L. NAA indicates N-acetyl-aspartate; Pi, inorganic phosphate; tCho, total choline; and tCr, total creatine.

*Value refers to the Pi(2) peak.
†No Pi(2) peak is observable in healthy tissue. For details, please refer to the Methods and Discussion sections.

By combining metabolite concentrations for tCr and phosphocreatine obtained from 1H and 31P examinations, we calculated the concentration of uCr. A significantly reduced amount of uCr was apparent in ischemic tissue (0.99±1.79 versus 4.66±1.77 mmol/L; P<0.001).

There was a significant negative correlation between stroke lesion volume and NAA in ischemic tissue (r=-0.56; P=0.03, indicating that larger infarcts go along with greater NAA reductions. Stroke lesion volume was also negatively correlated with creatine (r=-0.58; P=0.02) and phosphocreatine (r=-0.63; P=0.004).

Markers of Phospholipid Metabolism

We found a significant reduction in tCho in infarcted tissue (1.42±0.63 versus 2.15±0.40 mmol/L; P=0.007; Table 2). Glycerophosphocholine, being the major component of the tCho signal, was significantly lower in infarcted tissue as well (1.26±0.27 versus 1.51±0.28 mmol/L; P=0.002). Glycerophosphoethanolamine (1.08±0.42 versus 1.31±0.38 mmol/L) and phosphoethanolamine (1.01±0.23 versus 1.13±0.30 mmol/L) were also significantly reduced (P=0.03). No change of phosphocholine concentrations was observed in infarcted versus healthy tissue (0.29±0.11 versus 0.30±0.12 mmol/L; P=0.67).

We found a significant correlation between stroke lesion volume and reduction of glycerophosphocholine (r=-0.57; P=0.01). No significant correlation was found for all other lipid metabolites and lesion volume or time to first MRSI.

Discussion

This MRSI study of patients with ischemic stroke investigated tissue pH in subacute stroke together with intermediates of the cell energy and membrane lipid cycles, as well as neuronal and glial metabolites. Its major findings are an alkalotic milieu and an only incomplete depletion of energy-rich metabolites within ischemic tissue.

Tissue pH

We found a markedly increased tissue pH in the subacute phase of stroke. Further, we could not consistently find lactate within the ischemic tissue under investigation. Both the results seem counterintuitive but may be explained the following way: we suggest that the alkalotic pH and the frequent absence of lactate is attributable to the time point of our measurements, performed on average 6 days after stroke onset ranging from day 3 to 12. Commonly, acidosis is found in acute ischemic tissue subsequent from anaerobic glycolysis and accumulation of lactate. Several studies have shown an intracellular acidic pH and lactate in acute experimental stroke. Animal and human studies, however, have shown a switch from acidosis to alkalosis during the first days after ischemia. A previous human positron–emission tomography study also supports the hypothesis of alkalosis in subacute infarcted tissue, as it shows increased intracellular pH together with reduced oxygen extraction fraction, that is, a transient luxury perfusion. This alkalosis has been explained as a result of microglial invasion, of alternated buffering mechanisms especially by the sodium–hydrogen antiporter and as a result of cerebral edema. Our first (and widely used) approach for pH assessment (single-peak
approach) suggests an increasing pH with increasing stroke size. However, the more sophisticated data evaluation using 2 distinct Pi signals gives room for additional explanations: One is that Pi(2) is actually present in all infarcts but is only visible in larger strokes because of the physical limitations of MRSI. The real pH in subacute ischemia would thus be closer to the pH derived from Pi(2) in large strokes, namely ≈7.37. The lower pH values in older single-peak studies being closer to the 7.09 value of our single-peak approach would thus be the result of averaging Pi(1) and Pi(2), with Pi(1) also resulting from partial volume effects and this method may underestimate changes in tissue pH. Therefore, we suggest that the more sensitive double-peak approach used here should be incorporated in further studies evaluating changes of the tissue pH in the time course after stroke.

We further suggest that the 2 distinct Pi signals are a key finding in interpreting the paradoxical switch of pH from acute to subacute stroke. Splitting of the pH suggests an active process in a subgroup of cells because passive adjustments would be on a continuum from acidic to pH neutral. We propose that this switch could be because of upregulated countering mechanisms in ischemic tissue. The hypoxia-inducible factor 1 (HIF-1) has been shown to act on important mediators of intracellular pH, such as the sodium–hydrogen antiporter and the carbonic anhydrase enzyme. In brain tumors, these mechanisms have been shown to increase the pH, despite a hypoxic milieu. The more alkalotic milieu may be because of upregulated buffering processes within still viable cells. As an example, the sodium–hydrogen antiporter may be activated by inflammatory cytokines to be continuously active above a physiological threshold of tissue pH ≈7.05 in brain tissue.15 This hypothesis presupposes that there is still a viable amount of cells in the subacute stage of stroke that can counteract acidosis from hypoxia and the 2 Pi peaks may represent 2 different cellular entities, either neuronal and glial cells or neuronal cell populations being differentially affected by ischemia.

Our results suggest that further attention should be directed toward the role of tissue pH in the poststroke microenvironment, especially with regard to the effect on tissue transformation through active enzymatic cascades. Results by Zhao et al16 demonstrated that the inhibition of the matrix metalloproteinase 9 yields drastically different results depending on the time of modification, being beneficial in the acute and deleterious in the subacute phase of stroke. Given the fact that enzymes of the metalloproteinase group are pH-dependent and have been shown to be activated in tissue acidosis17 it is of importance to study the effect of temporal changes of tissue pH on their enzymatic activity. As a consequence, further animal studies should combine histological and in vivo pH imaging methods to clarify these interactions.

We found neither a significant correlation between the existence of lactate and elapsed time to first measurement nor with the infarct size and suggest that accumulated lactate had been removed from the ischemic tissue at the time of our measurements by diffusion or partial reperfusion.

Energy Metabolism
The hypothesis of still viable cells within the infarcted tissue is further reflected in our results concerning cell energy metabolism. After breakdown of aerobic glycolysis, a pronounced reduction of energy carriers in ischemia may be expected because anaerobic glycolysis produces only 2 ATP molecules compared with the additional 36 ATP molecules resulting from oxidative phosphorylation. Surprisingly, the levels of phosphocreatine and ATP were only reduced by about one fifth (and not completely depleted). One possible explanation is in line with the apparent pH split, suggesting that a still functioning moiety of cells would contribute to the phosphocreatine and ATP signals. It is well known that the vulnerability of cerebral tissue is different between cell types and even between neurons themselves.18 We found a significant reduction in NAA, tCr, and tCho in subacute stroke; the NAA reduction to 35% of healthy levels was more prominent than that of tCr (to 49%). The predominant reduction in NAA hints at a higher susceptibility of neurons to ischemia because NAA synthesis is almost only performed in neuronal mitochondria. Creatine, however, being synthesized in neurons and especially in glial cells which are more resistant to ischemia,19 was less reduced. We found that the amount of uCr was significantly higher in healthy compared with infarcted tissue supporting the concept of a reduced energy demand within ischemic tissue. In other words, most of the remaining creatine in infarcted tissue was in fact energy-rich phosphocreatine. To conclude, different vulnerability to hypoxia, different perfusion states in the subacute phase of stroke and decreased energy demand may explain the relative maintenance of high-energy substrates within ischemic tissue.

An important limitation of our study is the relatively heterogeneous patient collective with some patients having smaller infarctions than others. Some metabolic disturbances may thus be underestimated because of partial volume effects. A major limitation is the lack of longitudinal data examining the transition from the acute to the subacute stroke phase. Perfusion-weighted imaging was not acquired at the time of scanning, prohibiting the assessment of interactions between reperfusion status and brain pH. Strengths of our study are the combined use of 1H and 31P spectroscopic and anatomic T2w imaging ensuring an excellent overlap among modalities. The combined use of 3 Tesla MRSI allowed a sophisticated analysis of tissue pH and to calculate further metabolites, such as uCr.

In conclusion, we could show that tissue pH in subacute human cerebral ischemia is characterized by a shift toward a more alkalotic milieu. This is best explained by active compensatory mechanisms because of still viable cell populations within the ischemic tissue. Further studies are necessary to clarify the effect of tissue pH on tissue regeneration after stroke.

Disclosures
None.

References


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http://stroke.ahajournals.org/content/46/2/441

Data Supplement (unedited) at:
http://stroke.ahajournals.org/content/suppl/2016/04/06/STROKEAHA.114.007896.DC1
多核磁気共鳴分光法により評価したヒトの亜急性期脳虚血における pH およびエネルギー状態の変化

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背景および目的：脳のpHおよびエネルギー代謝のin vivoにおける変化は、脳卒中の病態生理を理解する上で重要である。本研究は、亜急性期脳虚血中におけるpHの変化およびエネルギー代謝との関連性を検討することを目的とした。脳卒中におけるアンドローシスとは異なり、これらについてはまだ十分に解明されていない。

方法：1H MRSおよび31P MRSを併用し、亜急性期脳卒中における脳組織のpH、リン脂質、さらにクレアチニン、クレアチニン酸、N-アセチルアスパラギン酸を含む細胞のエネルギー指標を測定した。本研究では初発虚血性脳卒中患者19例を登録した（脳卒中発症後期間の平均値は6日）。多変量ANOVAを用いて虚血組織と対側（健常）組織における代謝物濃度を比較し、それらの組織コンパートメント間の代謝物濃度の有意差を評価した。

結果：亜急性期脳卒中では、1つのピークがアルカリ側に移行したpHシグナルの分裂を同時に、健常な対側組織と比較してpHが有意に上昇した組織群が認められた（pH = 7.09対7.03, p = 0.002）。さらに、高エネルギー代謝物は中程度の減少にとどまったが（クレアチニン酸は17%の減少、ATPは19%の減少）、クレアチニンは51%減少した。

結論：これまで亜急性期脳虚血においてアルカリ側に移行したpHの分裂は示されていなかった。亜急性期脳虚血においてpHの分裂、およびエネルギーの不完全な喪失がみられたことから2種類の生存可能な細胞が存在することが示唆され、これは急性脳虚血後の代謝メカニズムの活性化によって最も良く説明される。

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左中大脳動脈領域の脳卒中患者の31P MRSおよび1H MRS。1H MRSでは1Hの主要代謝物のシグナルが減少しているが、乳酸は可視される。虚血領域におけるpHピークの拡大は対側（健常）組織の狭いpHピークの違いから明確に区別できる。クレアチニン酸（PCr）は減少しているが、広範囲のMCA脳卒中の中程度で検出可能である。GPC：グリセロリン酸コリン,GPE:グリセロホスホエタノールアミン,NAA:N-アセチルアスパラギン酸,PCr:ホスフォリチルシン,PEth:ホスフォエタノールアミン, Pi:無機リン酸,tCho:総コリン, tCr:総クレアチニン。