Deep-Tissue Oxygen Monitoring in the Brain of Rabbits for Stroke Research

Nadeem Khan, PhD; Huagang Hou, MD; Clifford J. Eskey, MD; Karen Moodie, MSc; Sangeeta Gohain, MSc; Gaixin Du, MSc; Sassan Hodge, PhD; William C. Culp, MD; Periannan Kuppusamy, PhD; Harold M. Swartz, MD, PhD

The primary event in the ischemic stroke is a rapid decline in the oxygen levels after the loss of blood flow in specific areas of the brain. Subsequent pathological processes result in a central core area of severely ischemic tissue surrounded by a region of moderate ischemic tissue (penumbra) with a preserved cellular metabolism. The outcome of an ischemic stroke depends on the size of the infarct core and the potential to salvage the cells in the penumbra, which is hypoperfused, and therefore, at risk of infarction but still viable. Such viable penumbral tissue can be rescued by quick interventions that can increase oxygen levels or slow metabolism in the ischemic area to minimize oxidative injury on reperfusion.

Several strategies have been investigated to rescue ischemic tissue using experimental models, especially rodents, but largely failed in subsequent clinical trials. The rabbit model of ischemic stroke using embolic clot is a promising model for developing effective strategies. This model first led to the prediction of the clinical response of recombinant tissue-type plasminogen activator to restore blood flow in patients. The drug is currently recommended for administration within 3 to 4.5 hours for best outcomes and has also shown modest benefit when administered within 4.5 to 6 hours of clinical onset. The rabbit model of embolic clot is now considered as a per -
loop (12–16 mm diameter) at one end and a transmission line with sensor loops (0.4–0.5 mm inner diameter) at the other end (Figure 1A). The sensor loops (or tips) are loaded with 30 to 50 μg of lithium phthalocyanine (LiPc, oximetry probe) crystals. The length of the transmission lines defines the depth and can be varied as needed for the experiment. The number of sensor loops and the distance between them can also be varied to measure \( pO_2 \) at \( \geq 1 \) sites in the brain of rabbits. The entire resonator is coated with a gas permeable and biocompatible Teflon AF2400. The mean area of the oximetry probe at the surface of each sensor loop is estimated to be 1.3 to 1.6 mm²; EPR oximetry, therefore, samples a region that includes many capillary segments and provides average \( pO_2 \) at the site of sensor loop. Histological examination of the cerebral tissue in the rabbit euthanized 4 weeks after the placement of implantable resonator did not show any obvious accumulation of inflammatory cells or blood cells surrounding the sensor loops. Similar results were also evident as early as 7 days after the placement of implantable resonator with 6-mm length of transmission lines in the brain of rats. The head of the anesthetized rabbit was antiseptically treated with Betadine, and 70% alcohol scrubs. A small incision (2–3 cm) was made on the skin and burr holes were gently created by using 18-gauge needle on the skull at predefined coordinates (anterior–posterior from bregma, −2.0 mm; medial–lateral from midline, 4 and 8 mm on each hemisphere; dorsal–ventral from surface of skull, 15 and 10 mm in each hemisphere). The sensor loops, SL1 and SL4, were located in the parietal cortex, whereas SL2 and SL3 were located in the subcortex (basal forebrain/internal capsule) region of the brain. The position of the sensor loops can be altered depending on the coordinates of the compromised tissue after ischemic stroke. The sensor loops were placed at the desired depth and the coupling loop was placed on the skull below the skin, to allow inductive coupling with the external surface loop resonator of the EPR spectrometer (Figure 1B). The incision on the skin was closed with nonabsorbable 3-0 nylon suture and the rabbit was monitored for recovery as per the Institutional Animal Care and Use Committee protocol. The repeated measurements of brain \( pO_2 \) by EPR oximetry was started 72 hours (day 3) after the placement of implantable resonator and the measurements were repeated for 4 weeks.

**In Vivo EPR Oximetry**

EPR oximetry requires one-time implantation of the oxygen probes (LiPc or implantable resonator), but rest of the procedure for \( pO_2 \) measurement is entirely noninvasive and can be...
repeated as desired. The basis of oximetry is the paramagnetic nature of oxygen, which broadens the EPR signal of the probe in proportion to the amount of oxygen. EPR oximetry has unique capabilities and advantages compared with other techniques, such as (1) direct measurement of absolute pO2 in the tissue of interest, (2) pO2 is quantified through a physical interaction of oxygen with the probe (does not require oxygen consumption), (3) pO2 measurements can be made continuously and repeatedly as desired, without a confounding influence of previous measurements, (4) The oxygen sensors are metabolically inert and coated with Teflon, therefore, do not perturb the tissue microenvironment, including oxygen content, and (5) there is no other technique available at present to make repeated measurement of tissue pO2 without the need to reintroduce the probe for each measurement.

A 1.2-GHz EPR spectrometer equipped with a surface loop resonator and a set of gradient coil for multisite oximetry was used for monitoring brain pO2 in the rabbit. The anesthetized rabbit (2.5% isoflurane in 30% O2) was positioned in the EPR magnet and the external surface loop resonator was gently placed over the head region. A magnetic field gradient of 1.7 G/cm per ampere was used to separate the EPR spectra from each sensor loop for simultaneous multisite oximetry. The peak-to-peak line widths of the EPR spectra were used to determine pO2 by using the calibration of implantable resonator (Figure 1C and 1D). The rabbit was maintained at 38±1.0°C (monitored via a rectal probe) by keeping the animal under warm air during the EPR measurements. The EPR settings were as follows: incident microwave power, 0.4 to 1.2 mW; modulation frequency, 24 kHz; magnetic field center, 410 G; scan time, 10 s, scan range, 8 to 12 G, and modulation amplitude not exceeding one third of the line width. The implantable resonator appears as a signal void in T1-weighted MRI scans, which can be used to confirm their position in the brain of rabbits.

**Results**

Brain pO2 was measured for 20 to 25 minutes on day 3 and the measurements were repeated on days 5, 7, 14, 21, and 28 (Figure 2). The mean (SD) baseline pO2 at each site (SL1–SL4) in the brain on day 3 was 39.2 (2.2), 41.6 (1.4), 41.3 (1.7), and 43.6 (2.0) mm Hg, respectively, and only a modest variation was observed in the measurements repeated subsequently for ≤4 weeks. To mimic the ischemic situation with low levels of oxygen and test the response of implantable resonator, the breathing gas was switched to 15% O2 for 15 minutes and then returned to 30% O2, (Figure 3A). To test the potential effect of hyperoxia on brain pO2, the breathing gas was switched to carbogen (95% O2+5% CO2) for 20 to 25 minutes and then returned to 30% O2 (Figure 3B). These experiments were performed on days 7, 14, 21, and 28. The brain pO2 measured at 4 sites decreased by ≈30% from baseline in rabbit breathing 15% O2. However, brain pO2 measured at 4 sites increased significantly by ≥75% during carbogen breathing. An exponential quadratic function of time was used to determine minimum pO2 (PO2min) attained during 15% O2 on each day and the time to reach the PO2min (Tmin) (Figure 4). Similar analysis was used to determine maximal pO2 (PO2max) attained on each day and the time to reach maximum pO2 (Tmax) during carbogen inhalation. These analyses suggest that it took 10 to 14 minutes to reach a minimal or maximal pO2 when the breathing gas was switched from 30% O2 to 15% O2 or carbogen, respectively. A similar time scale was noted for the brain pO2 to return to the baseline level when the breathing gas was switched from 15% O2 or carbogen to 30% O2. We anticipate that such temporal pO2 information will be extremely useful in designing hyperoxic therapies to modulate oxygen levels in ischemic stroke.
emic stroke. The brain pO2 data presented here was obtained in the nonischemic contralateral brain can be used as internal experiment. A stable brain pO2 was observed from day 3, which the sensor loops can be modified as needed for a particular length of transmission lines, number, and distance between the region of interest, that is, infarct core and penumbra after ischemic loops should be carefully selected so that they are located in the gen levels and thus save vital tissue loss in ischemic stroke. The extent of increase in tissue pO2 that occur during the course of ischemic stroke, a capability, which was not available hitherto. Dynamic information of cerebral pO2 can be used to test and to optimize strategies for improving treatment outcome of ischemic stroke. EPR oximetry using implantable resonator in the brain of rabbits. The measurement of tissue pO2 be initiated as early as 3 days after the placement of implantable resonator in the brain of a rabbit. The pO2 measurements can be repeated as desired. A rapid decline in tissue pO2 during 15% O2 breathing potentially highlights the immediate changes in the oxygen levels that may occur in ischemic stroke. The extent of increase in tissue pO2 during carbogen breathing is encouraging and can be potentially used as a therapeutic strategy to improve oxygen levels and thus save vital tissue loss in ischemic stroke.

For the multisite oximetry approach, the position of sensor loops should be carefully selected so that they are located in the region of interest, that is, infarct core and penumbra after ischemic stroke. The design of the implantable resonator, including the length of transmission lines, number, and distance between the sensor loops can be modified as needed for a particular experiment. A stable brain pO2 was observed from day 3, which suggest that the experiments to investigate ischemic stroke can be initiated as early as 3 days after the placement of implantable resonator in the brain of rabbits. The measurement of tissue pO2 in the nonischemic contralateral brain can be used as internal control and investigate adaptive response of the brain to ischemic stroke. The brain pO2 data presented here was obtained in a rabbit to illustrate the capability of temporal monitoring by EPR oximetry. We are currently implementing this technique to investigate temporal changes in the brain pO2 in additional rabbits during hyperoxia and ischemic stroke.

Conclusions

We have demonstrated a direct and longitudinal measurement of absolute tissue pO2 at several sites simultaneously in the brain of rabbit by EPR oximetry using implantable resonators, a capability, which was not available hitherto. Dynamic information of cerebral pO2 can be used to test and to optimize strategies for improving treatment outcome of ischemic stroke. EPR oximetry with implantable resonators should also be useful to investigate the effect of other pathologies, such as traumatic brain injury and cold injury, on the oxygen levels in the brain of clinically pertinent animal models.

Sources of Funding

National Institutes of Health grants R21NS082585 to Dr Khan, R01EB004031 to Dr Kuppusamy, and the Electron Paramagnetic Resonance Center, Geisel School of Medicine at Dartmouth, Lebanon, NH.

Disclosures

None.

References


**Key Words:** carbogen • EPR oximetry • hyperoxia • implantable resonator • ischemia • pO2 • stroke
Deep-Tissue Oxygen Monitoring in the Brain of Rabbits for Stroke Research
Nadeem Khan, Huagang Hou, Clifford J. Eskey, Karen Moodie, Sangeeta Gohain, Gaixin Du, Sassan Hodge, William C. Culp, Periannan Kuppusamy and Harold M. Swartz

Stroke. published online January 22, 2015;
Stroke is published by the American Heart Association, 7272 Greenville Avenue, Dallas, TX 75231
Copyright © 2015 American Heart Association, Inc. All rights reserved.
Print ISSN: 0039-2499. Online ISSN: 1524-4628

The online version of this article, along with updated information and services, is located on the
World Wide Web at:
http://stroke.ahajournals.org/content/early/2015/01/22/STROKEAHA.114.007324.citation

Permissions: Requests for permissions to reproduce figures, tables, or portions of articles originally published in Stroke can be obtained via RightsLink, a service of the Copyright Clearance Center, not the Editorial Office. Once the online version of the published article for which permission is being requested is located, click Request Permissions in the middle column of the Web page under Services. Further information about this process is available in the Permissions and Rights Question and Answer document.

Reprints: Information about reprints can be found online at:
http://www.lww.com/reprints

Subscriptions: Information about subscribing to Stroke is online at:
http://stroke.ahajournals.org//subscriptions/